

Size-partitioned phytoplankton carbon and carbon-to-chlorophyll ratio from ocean colour by an absorption-based bio-optical algorithm

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1 **Size-partitioned phytoplankton carbon and**
2 **carbon-to-chlorophyll ratio from ocean-colour by**
3 **an absorption-based bio-optical algorithm**

4 **Shovonlal Roy^{*1}, Shubha Sathyendranath^{2,3} and Trevor Platt²**

5 ¹Department of Geography and Environmental Science & School of
6 Agriculture, Policy and Development, University of Reading, Whiteknights,
7 Reading RG6 6AB, U.K

8 ²Plymouth Marine Laboratory, Prospect Place, Plymouth PL1 3DH, U.K.

9 ³National Centre for Earth Observation, Plymouth Marine Laboratory,
10 Prospect Place, Plymouth PL1 3DH, U.K.

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*Corresponding author. E-mail: shovonlal.roy@reading.ac.uk

Abstract

The standing stock of phytoplankton carbon is a fundamental property of oceanic ecosystems, and of critical importance to the development of Earth System models for assessing global carbon pools and cycles. Some methods to estimate phytoplankton carbon at large scales from ocean-colour data rely on the parameterization of carbon-to-chlorophyll ratio, which is known to depend on factors such as the phytoplankton community structure, whereas other methods are based on the estimation of total particulate organic carbon (POC), and rely on the assumption that a known fraction of POC is made up of phytoplankton carbon. The carbon-to-chlorophyll ratio is also used in marine ecosystem models to convert between carbon and chlorophyll, a common requirement. In this paper we present a novel bio-optical algorithm to estimate the carbon-to-chlorophyll ratio, and the standing stocks of phytoplankton carbon partitioned into various size classes, from ocean colour. The approach combines empirical allometric relationships of phytoplankton size structure with an absorption-based algorithm for estimating phytoplankton size spectra developed earlier. Applying the new algorithm to satellite ocean-colour data from September 1997 to December 2013, the spatio-temporal variations of carbon-to-chlorophyll ratio and phytoplankton carbon across various size classes are computed on a global scale. The average annual stock of phytoplankton carbon, integrated over the oceanic mixed-layer depth, is estimated to be ~ 0.26 gigatonnes, with the size-partitioned stocks of 0.14 gigatonnes for picoplankton, 0.08 gigatonnes for nanoplankton and 0.04 gigatonnes for microplankton. The root-mean-square error and the bias in the satellite-derived estimates of picoplankton carbon, when compared with corresponding *in situ* data, are found to be 36.23 mgC m^{-3} and $-13.53 \text{ mgC m}^{-3}$, respec-

35 tively, on individual pixels. The regional uncertainties in the estimates of phytoplankton
36 carbon are calculated to be less than the relative uncertainties in other satellite-derived
37 products, for most parts of the global ocean, and can amplify only for certain oceano-
38 graphic regions. Although the new estimates of phytoplankton are of the same order
39 of magnitude as those based on existing models, our study suggests that a consensus
40 is yet to be built on the accurate sizes of the phytoplankton carbon pools; improved
41 satellite chlorophyll products, and better estimates of inherent optical properties would
42 be essential pre-requisites to minimising the uncertainties.

43 **Keywords**

44 Phytoplankton carbon; carbon-to-chlorophyll ratio; ocean colour; carbon-based size class;
45 picoplankton, nanoplankton, microplankton; phytoplankton size spectra.

46 **1 Introduction**

47 Although the standing stock of the autotrophic biomass (phytoplankton) in the ocean is only
48 a small fraction (less than 1%) of the Earth's photosynthetic biomass, approximately half
49 (~ 50 gigatonnes C) of the global annual carbon-fixation is accounted for by the oceanic au-
50 totrophs through primary production (Falkowski, 2012; Field et al., 1998). Therefore, for
51 understanding, estimating and monitoring the carbon dynamics in the ocean, it is impor-
52 tant to be able to make accurate measurements of the standing stocks of phytoplankton
53 carbon. However, major complexities in carbon estimation arise from phytoplankton commu-

54 nity composition; for example, the carbon content of a phytoplankton cell varies with species
55 and its morphological characteristics (e.g., large vs small cell size); it also depends on the
56 ambient light and nutrient conditions (Marañón, 2008; Marañón et al., 2013; Menden-Deuer
57 and Lessard, 2000). Another level of complexity in estimating phytoplankton carbon accu-
58 rately arises from uncertainties in parameterising the carbon-to-chlorophyll ratio (χ), which
59 is used to convert phytoplankton-carbon biomass to chlorophyll biomass in ecosystem mod-
60 els for comparison with satellite-derived chlorophyll data, and also in satellite algorithms
61 for estimating phytoplankton carbon from chlorophyll data (Sathyendranath et al., 2009).
62 A standard product from ocean-colour remote sensing is chlorophyll concentration (e.g.,
63 <http://oceandata.sci.gsfc.nasa.gov/>; <https://www.oceancolour.org/>). Marine biogeochemical
64 and ecosystem models (e.g., <http://pft.ees.hokudai.ac.jp/maremip/index.shtml>), on the other
65 hand, deal with phytoplankton biomass in carbon units and use a carbon-to-chlorophyll ra-
66 tio. The magnitude of carbon-to-chlorophyll ratio can vary over two orders of magnitude
67 depending on phytoplankton community composition and environmental conditions (Geider,
68 1987; Geider et al., 1998; Sathyendranath et al., 2009), and hence it may lead to significant
69 uncertainties in the conversions between the two measures. Furthermore, the retrieval of
70 phytoplankton carbon from remote sensing of ocean colour is also affected by the presence of
71 particulates, other than phytoplankton that contribute to the water-leaving radiance captured
72 by the sensors. Dissolved constituents such as coloured dissolved organic materials (CDOM)
73 that absorbs strongly in the blue wavelengths can also affect the remotely-sensed ocean colour
74 and interfere with chlorophyll-a retrievals, particularly in coastal and high latitudes. Owing
75 to these complexities, the estimation of phytoplankton carbon from remote sensing is recog-

76 nised as a non-trivial task, and it is essential to improve satellite-based algorithms for use in
77 carbon-cycle research (Behrenfeld et al., 2005; Kostadinov et al., 2016; Sathyendranath et al.,
78 2009).

79 Nevertheless, algorithms have been developed to compute particulate organic carbon
80 (POC) in the ocean from remotely-sensed ocean colour. For example, Stramski et al. (2008)
81 derived a band-ratio algorithm that uses the blue-to-green band ratio of remote-sensing re-
82 flectance to calculate the concentration of POC. This algorithm can then be used to compute
83 phytoplankton carbon by assuming a constant ratio of phytoplankton carbon to POC in the
84 ocean (Stramski et al., 2008). Behrenfeld et al. (2005) derived an empirical relationship to
85 compute phytoplankton carbon from particulate backscattering coefficients by assuming a
86 fixed ratio of 30% between phytoplankton carbon and POC. More recently, Kostadinov et al.
87 (2016) developed an algorithm to compute phytoplankton carbon from particulate backscat-
88 tering coefficient using allometric relationships for the POC particle size distribution and
89 assuming that the fraction of carbon in the living phytoplankton relative to that of POC is
90 $1/3$. Kostadinov et al. (2016) also computed the absolute and the fractional carbon biomass in
91 three size classes of phytoplankton, i.e., picoplankton (with diameter $0.5\text{-}2\ \mu\text{m}$), nanoplank-
92 ton (with diameter $2\text{-}20\ \mu\text{m}$) and microplankton (with diameter $20\text{-}50\ \mu\text{m}$), under these
93 assumptions. Although the existing algorithms may provide a mutually comparable estimate
94 (in order of magnitude) of total phytoplankton carbon in the global ocean, the underlying
95 assumption of a constant ratio of phytoplankton carbon and POC imposes significant un-
96 certainties in regional estimates of phytoplankton carbon and its spatial distributions. This

97 is important because the ratio of phytoplankton carbon to POC varies over a wide range,
98 from 14% to 85%, across a variety of oceanographic regions (Behrenfeld et al., 2005; DuRand
99 et al., 2001; Eppley et al., 1992; Gundersen et al., 2001; Kostadinov et al., 2016; Oubelkheir
100 et al., 2005; Redalje and Laws, 1981; Stramski et al., 2008). Furthermore, with the excep-
101 tion of Kostadinov et al. (2016), current algorithms are limited in their ability to retrieve
102 the carbon-based classification of phytoplankton functional types (PFT) or phytoplankton
103 size classes (PSC), though many methods are available to estimate the fractional chlorophyll
104 distribution across PFTs and PSCs (e.g., IOCCG, 2014). Given the importance and wide
105 applications of satellite-based PFTs, it is important to improve our understanding on phyto-
106 plankton carbon stocks in various PSCs and PFTs, through developing new algorithms based
107 on complementary bio-optical variables.

108 In this paper, we present a new bio-optical algorithm to estimate phytoplankton carbon
109 from remotely-sensed ocean-colour data, designed by targeting the photosynthetic phyto-
110 plankton cells directly. The algorithm builds on Roy et al. (2013), where we developed a
111 semi-analytical method to compute the exponent of the phytoplankton size spectrum from
112 the specific-absorption coefficient of phytoplankton (which depends on chlorophyll concentra-
113 tion and total absorption by phytoplankton), and derived the equivalent spherical diameter
114 of phytoplankton cells and the fractions of chlorophyll in any size class of phytoplankton,
115 in particular, those for picoplankton, nanoplankton and microplankton. Here, the method
116 is extended for computing carbon-to-chlorophyll ratio from ocean colour applicable to any
117 size class of phytoplankton, by combining analytically the allometric relationships between

118 phytoplankton cell size and carbon content with the size-spectrum algorithm of Roy et al.
119 (2013, 2011), and implementing them to estimate phytoplankton carbon in any size class.
120 The method is applied to ocean-colour data for the period 1997-2013, and is validated using
121 the available *in situ* data. Results are discussed in relation to the applicability of this method
122 to obtain independent remote-sensing-based measurements of phytoplankton carbon, and the
123 carbon budget, according to phytoplankton size. The results pave the way to improved im-
124 plementation of carbon-based growth models using satellite data for computation of primary
125 production in various PSCs.

126 2 Data

127 We used a continuous time series of ocean-colour data on global scale produced by
128 the European Space Agency's Ocean Colour Climate Change Initiative (OC-CCI) project
129 (<http://www.esa-oceancolour-cci.org>) through systematically merging the available satellite
130 data from three major sensors: NASA-SeaWiFS, NASA-MODIS-Aqua and ESA-MERIS. For
131 temporal consistency of OC-CCI products, and for algorithms selected for processing them,
132 please see Belo Couto et al. (2016); Brewin et al. (2015); Müller et al. (2015). We used the
133 global 4-km, level-3 mapped products from OC-CCI, the details of which can be found in
134 <http://www.esa-oceancolour-cci.org> (also in, Sathyendranath et al., unpublished ms). Fur-
135 ther, to validate the new algorithm we used a global dataset on pico-phytoplankton carbon
136 compiled by Buitenhuis et al. (2012) that included flow cytometry data obtained since the late
137 1980s during cruises throughout most of the world ocean, as a contribution to the MARE-

138 DAT World Ocean Atlas of Plankton Functional Types database. The details of the database
139 can be found in Buitenhuis et al. (2012) and in <http://doi.pangaea.de/10.1594/PANGAEA>.
140 We extracted a subset of this database to cover the time period from September 1997 to
141 December 2013, over which the satellite-based ocean-colour data were available. We further
142 obtained mixed-layer depths from Monthly Isopycnal & Mixed-layer Ocean Climatology (MI-
143 MOC, Schmidtko et al., 2013, <http://www.pmel.noaa.gov/mimoc/>), and remapped those to
144 OC-CCI 4-km grids using nearest-neighbour interpolation (using MATLAB2015b interpola-
145 tion routine).

146 3 Development of the bio-optical algorithm

147 3.1 Exponent of phytoplankton size spectra (ξ) from their absorp- 148 tion coefficients $a_{ph}(\lambda)$ following Roy et al. (2013)

149 The exponent of the phytoplankton size spectrum (ξ) can be computed from the absorption
150 coefficient of phytoplankton at 676 nm, $a_{ph}(676)$, using a method developed by Roy et al.
151 (2013). For the completeness of the methodology of this paper, we briefly describe below the
152 principal steps for retrieval of ξ , without fully reproducing it from Roy et al. (2013). In this
153 method, it was assumed that the particle size distribution of phytoplankton cells follows the
154 power law, so the number of phytoplankton cells per unit volume of seawater with a particle
155 diameter of D was expressed as $N(D) = kD^{-\xi}$, with ξ as the exponent of the phytoplank-
156 ton size spectrum, and k a constant related to the abundance of the total population. A

157 relationship was then derived between the concentration of chlorophyll-a (B in mg Chl m^{-3})
158 within a diameter range $[D_{min}, D_{max}]$ of phytoplankton cells and the exponent of the phyto-
159 plankton size spectrum, by considering that the concentration of chlorophyll-a within the size
160 interval (diameter range $[D_{min}, D_{max}]$) would be a product of the number of phytoplankton
161 cells within that size class, the volume of each cell, and the intracellular concentration of
162 chlorophyll-a (c_i). The quantity c_i (mg Chl-a m^{-3}) was parameterised as : $c_i = c_0 D^{-m}$, with
163 the parameters $c_0 = 3.9 \times 10^6$ ($\text{mg Chl-a m}^{-2.94}$) and $m = 0.06$ (dimensionless), which were
164 estimated earlier by Roy et al. (2011) using the *in situ* measurements published by Marañón
165 et al. (2007). The concentration of chlorophyll-a (B in mg Chl m^{-3}) within the set diameter
166 range was then expressed as a function of ξ as follows:

$$B = \int_{D_{min}}^{D_{max}} \left[\left(\frac{\pi}{6} D^3 \right) (c_0 D^{-m}) (k D^{-\xi}) \right] dD = \left(\frac{\pi}{6} k c_0 \right) \frac{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}}{4 - \xi - m}, \quad (1)$$

167 with the parameters k , c_0 and m described as above.

168 Next, the specific absorption coefficient of chlorophyll-a (a_{chl}^* , as distinct from the specific
169 absorption of phytoplankton a_{ph}^*) was expressed as a function of the cell diameter (D). To do
170 so, phytoplankton absorption coefficient (a_{ph}) at 676 nm was considered with the assumption
171 that at this wavelength the contribution from auxiliary pigments, and substances other than
172 chlorophyll-a would be negligible (Roy et al., 2011). At this wavelength, the specific absorption
173 coefficient of the cell material of phytoplankton was assumed to be equal to a_{ci}^* , the specific-
174 absorption coefficient of chlorophyll-a inside the cell, with units of $\text{m}^2 (\text{mg Chl-a})^{-1}$ (Roy et al.,
175 2011); and following Duyens (1956), the theoretical value of the chlorophyll-specific absorption
176 of phytoplankton cells of diameter D was expressed as: $a_{chl}^*(676, D) = [3 a_{ci}^* Q_a(\rho_c)]/2\rho_c$, with

177 Q_a as the dimensionless absorption efficiency of a cell given by $Q_a(\rho_c) = 1 + [2 \exp(-\rho_c)]/\rho_c +$
 178 $2[\exp(-\rho_c) - 1]/\rho_c^2$, and ρ_c as the dimensionless optical thickness of the cell given by $\rho_c =$
 179 $\rho_c(676, D) = a_{ci}^*(676) c_0 D^{1-m}$. The observed absorption coefficient of chlorophyll-a at 676 nm
 180 due to the phytoplankton cells in the prescribed diameter range was then expressed as:

$$a_{chl}(676) = \int_{D_{min}}^{D_{max}} \left[\left(\frac{\pi}{6} D^3 \right) (c_0 D^{-m}) (k D^{-\xi}) \times a_{chl}^*(676, D) \right] dD. \quad (2)$$

181 Using Eqs. (1) and (2), the specific absorption of chlorophyll-a at 676 nm, due to phytoplank-
 182 ton cells in the diameter range $[D_{min}, D_{max}]$, was obtained as:

$$\begin{aligned} a_{chl}^*(676) &= \frac{a_{chl}(676)}{B} = \frac{1}{B} \int_{D_{min}}^{D_{max}} \left[\left(\frac{\pi}{6} D^3 \right) (c_0 D^{-m}) (k D^{-\xi}) a_{chl}^*(676, D) \right] dD \\ &= \frac{4 - \xi - m}{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}} \int_{D_{min}}^{D_{max}} \left[D^{3-\xi-m} \times a_{chl}^*(676, D) \right] dD. \end{aligned} \quad (3)$$

183 Note that, $a_{chl}^*(676, D)$ on the right-hand side of the above equation is the theoretical value
 184 of the specific-absorption coefficient of chlorophyll-a at 676 nm, expressed as a function of
 185 the equivalent spherical diameter D of phytoplankton, as described above based on Roy
 186 et al. (2011). For remote-sensing applications, $a_{ph}^*(676)$ is obtained from ocean colour by
 187 an algorithm for inherent-optical properties (IOP), for example, the Carder et al. (1999)
 188 algorithm as implemented in Roy et al. (2013). Further, from $a_{ph}^*(676)$, the quantity $a_{chl}^*(676)$
 189 is calculated using the method of Roy et al. (2011). The quantity ξ is then estimated from
 190 Eq. (3) numerically, by using a non-linear optimization algorithm. For further details on the
 191 methodology, parameterisation and optimization algorithm associated with the retrieval of ξ ,
 192 the reader is referred to Roy et al. (2013, 2011).

193 **3.2 Relating ξ to phytoplankton carbon and carbon-to-chlorophyll**
 194 **ratio (χ) using allometric relationships**

195 Allometric relationships appear to hold for phytoplankton communities, as well as for other
 196 organisms (Marañón, 2008; Marañón et al., 2013; Menden-Deuer and Lessard, 2000; Peters,
 197 1983; Strathmann, 1967). Menden-Deuer and Lessard (2000) have reported allometric rela-
 198 tionships between the cellular content of phytoplankton carbon (C_{cell}) and cell volume (V_{cell})
 199 for morphologically different dinoflagellates, diatoms and other protist groups. The allometric
 200 relationships take the following canonical form:

$$C_{cell} = a V_{cell}^b, \quad (4)$$

201 where V_{cell} is the volume of a phytoplankton cell expressed in μm^3 , C_{cell} is expressed in
 202 pg C cell^{-1} , and the quantities a and b are constants, which should ideally remain unchanged
 203 for a given ecological community. The concentration of phytoplankton carbon (C_{total} , in
 204 mgC m^{-3}) contained in the cells within a diameter range $[D_{min}, D_{max}]$ can then be expressed
 205 as:

$$\begin{aligned} C_{total} &= \int_{D_{min}}^{D_{max}} [\text{number of cells}] \times [\text{carbon content within a cell}] dD, \\ &= \int_{D_{min}}^{D_{max}} (kD^{-\xi}) \left[10^{-9} a \left(10^{18} \frac{\pi}{6} D^3 \right)^b \right] dD, \\ &= 10^{-9} k a \left(10^{18} \frac{\pi}{6} \right)^b \left(\frac{D_{max}^{3b-\xi+1} - D_{min}^{3b-\xi+1}}{3b - \xi + 1} \right). \end{aligned} \quad (5)$$

206 We note that the values 10^{-9} and 10^{18} are associated with the conversions of units from
 207 picogram to mg, and m^3 to μm^3 , respectively. In the special case when $\xi \rightarrow (3b + 1)$,
 208 the denominator in Eq. (5) goes to zero; so, to avoid division by zero, a limit of $C_{total} \rightarrow$

209 $\left[10^{-9} k a \left(10^{18} \frac{\pi}{6}\right)^b \log_e \left(\frac{D_{max}}{D_{min}}\right)\right]$, is used. Equations (1) and (5) relate ξ to the concentration
 210 of total phytoplankton chlorophyll (B , mg m^{-3}) and the total phytoplankton carbon (C_{total} ,
 211 mg m^{-3}), respectively, from which the carbon-to-chlorophyll ratio (χ) of the mixed population
 212 can be calculated as

$$\chi = \frac{C_{total}}{B} = \frac{10^{-9} a \left(10^{18} \frac{\pi}{6}\right)^b}{(\pi/6) c_0} \left(\frac{D_{max}^{3b-\xi+1} - D_{min}^{3b-\xi+1}}{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}}\right) \left(\frac{4 - \xi - m}{3b - \xi + 1}\right). \quad (6)$$

213 We note that the only unknown parameter k appearing in both Eqs. (1) and (5) cancels out
 214 within the expression of carbon-to-chlorophyll ratio (6). Once the exponent ξ is computed
 215 from Eq. (3) following the description in the previous section, χ can be computed directly
 216 from Eq. (6). Therefore, the total phytoplankton carbon can be calculated simply as,

$$C_{total} = \chi B = \frac{10^{-9} a \left(10^{18} \frac{\pi}{6}\right)^b}{(\pi/6) c_0} \left(\frac{D_{max}^{3b-\xi+1} - D_{min}^{3b-\xi+1}}{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}}\right) \left(\frac{4 - \xi - m}{3b - \xi + 1}\right) B. \quad (7)$$

217 It is clear that the estimates of phytoplankton carbon, using the above equations for χ
 218 and C_{total} , would depend on accurate parameterisation of the allometric relationship between
 219 phytoplankton cell volume and cellular carbon. However, the allometric parameters a and b
 220 are reported to vary across phytoplankton groups (Menden-Deuer and Lessard, 2000). So,
 221 the estimates of mixed phytoplankton carbon would be biased if the allometric parameters
 222 corresponding to any one phytoplankton group were used (Fig. 1a). More explicitly, according
 223 to Menden-Deuer and Lessard (2000), if the allometric relationship for protists (green line
 224 in Fig. 1a) were used, phytoplankton carbon would be underestimated for small cells and
 225 overestimated for large cells; if that for diatoms (blue line in Fig. 1a) were used, phytoplankton
 226 carbon would be underestimated for large cells; and finally, if that for dinoflagellate (yellow
 227 line in Fig. 1a) were used, phytoplankton carbon would be overestimated for small cells.

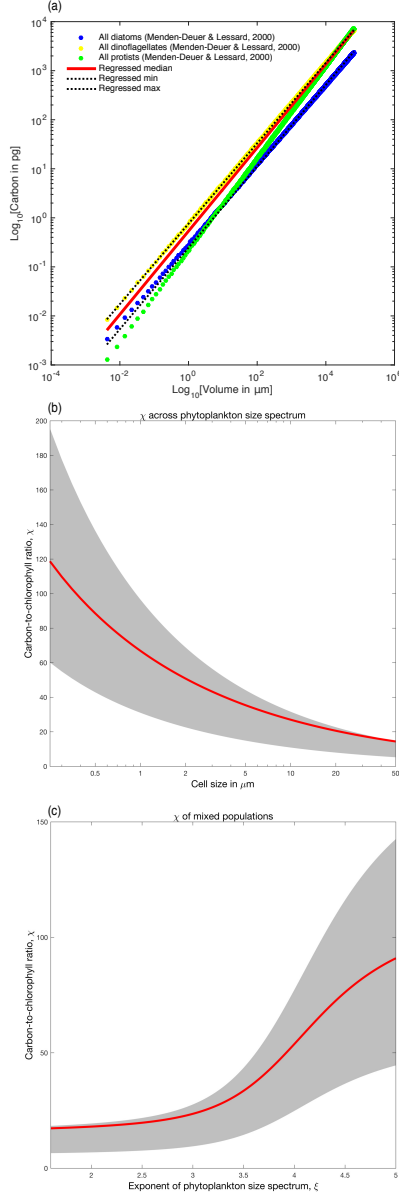


Figure 1: Reported and derived allometric relationships between phytoplankton carbon and their cell size. (a) Allometric carbon of diatoms (blue), dinoflagellates (yellow) and protists (green) reported by Menden-Deuer and Lessard (2000); and the allometric carbon for mixed phytoplankton a function of their cell volume derived by regression; the regressed median ($a = 0.54$, $b = 0.85$), and the lower ($a = 0.25$, $b = 0.83$) and upper ($a = 0.76$, $b = 0.82$) bounds are shown by red solid line, and two black-dotted lines respectively. (b)-(c) Derived relationship between carbon-to-chlorophyll ratio χ and phytoplankton size: (b) χ as a function of phytoplankton cell diameter for a homogenous population, calculated from Eq. (6) and the relationship between ξ to average cell diameter derived in Roy et al. (2013); and (c) χ as a function of the exponent of phytoplankton size spectrum ξ calculated from Eq. (6). In (b) and (c), the red lines represent the median of the allometric relationship shown in (a); and the grey areas represent the ranges of χ corresponding to the regressed minimum and maximum shown as black-dotted lines in (a).

228 Therefore, for calculating cellular carbon of mixed phytoplankton operationally, the allo-
229 metric parameters need to be established, which is not straight-forward. In a recent study,
230 Kostadinov et al. (2016) considered an approach in which four different allometric relation-
231 ships reported by Menden-Deuer and Lessard (2000) were used for two different parts of the
232 phytoplankton size spectrum. However, the allometric relationship is scale-free (as known
233 from allometric studies based on other species, e.g., Peters, 1983), and therefore, the al-
234 lometric parameters should remain unchanged across the size range of the phytoplankton
235 community. But deriving a new allometric relationship for phytoplankton based on *in situ*
236 data, applicable to all oceanographic regions and across all size ranges of mixed phytoplank-
237 ton, is out of the scope of this study, which aims at making a first estimate of phytoplankton
238 carbon using reported allometric relationships, and the new method. So, from an operational
239 perspective, we considered the various estimates of ‘*a*’ and ‘*b*’ reported by Menden-Deuer and
240 Lessard (2000) as independent observations, and derived, as described below, a continuous
241 allometric relationship with a view to applying them to mixed populations, assuming that
242 the populations are combinations of the phytoplankton groups for which the allometric re-
243 lationships were reported by Menden-Deuer and Lessard (2000). In this approach, we first
244 computed phytoplankton carbon over a broad range of cell volumes using the allometric rela-
245 tionships reported for protists, diatoms and dinoflagelletes, respectively (shown by the green,
246 blue and yellow dots, respectively in Fig. 1a). We next computed the median, minimum and
247 maximum of the three estimates of phytoplankton carbon, at each size, over the same range
248 of cell volumes (see, Fig. 1a). We then derived three allometric relationships between cell
249 volume and the median, minimum and maximum estimates of phytoplankton carbon, respec-

250 tively, using linear regression (the median is shown by solid red line, and the minimum and
251 maximum by dotted black lines in Fig. 1a). As expected, the revised allometric parameters,
252 corresponding to the regressed median ($a = 0.54$, $b = 0.85$, $r^2 > 0.95$), minimum ($a = 0.25$,
253 $b = 0.83$, $r^2 > 0.95$) and maximum ($a = 0.76$, $b = 0.82$, $r^2 > 0.95$), differed from the reported
254 allometric parameters corresponding to any particular phytoplankton group. However, the
255 regressed median line (red) in Fig. 1a would represent an approximate allometric relationship
256 for which the estimates of mixed-phytoplankton carbon would always be within the range of
257 estimates based on single phytoplankton groups. Furthermore, the minimum and maximum
258 estimates of the phytoplankton carbon at any size would be represented by the lower and
259 upper bounds for the allometric relationships (the dotted black lines in Fig. 1a) derived this
260 way from the group-specific allometric relationships.

261 The allometric parameters ‘ a ’ and ‘ b ’, derived by regression as above, can be incorporated
262 into the expression for carbon-to-chlorophyll ratio χ (Eq. 6) to describe the variations of χ
263 with phytoplankton size structure. For phytoplankton populations consisting of homogeneous
264 cells of the same size, the variation of χ as a function of the cell size of the population is shown
265 in Fig. (1b). When the population deviates from homogeneity and consists of cells of different
266 sizes, χ varies as a function of the exponent of size spectrum according to Fig. (1c). The
267 magnitude of χ decreases with increase in cell size (Fig. (1b, the black curve). For mixed
268 populations, χ increases with the exponent of phytoplankton size spectrum ξ (Fig. (1c, the
269 black curve). The shaded areas in Fig. (1b) and Fig. (1c) represent the lower and upper levels
270 of χ corresponding to the regressed-minimum and maximum of the allometric relationship.

271 The figures (1b-1c) show that the carbon-to-chlorophyll ratio of phytoplankton will be at
272 the higher end (e.g., $\chi > 100$) when the population is dominated by small cells, and would
273 decrease to a significantly lower value (e.g., $\chi < 20$) if the population were dominated by large
274 cells. These results are remarkably consistent, qualitatively, with empirically derived carbon-
275 to-chlorophyll ratios, e.g., those in Sathyendranath et al. (2009).

276 We next apply these relationships to derive analytical expressions for χ and phytoplankton
277 carbon for any given size class of phytoplankton population. Although we have used the
278 above allometric parameters for the rest of the calculations to obtain a first estimate of
279 phytoplankton carbon by our method, any improvement on the allometric relationships based
280 on new *in situ* data would improve our estimates of phytoplankton carbon, and it would be
281 straight-forward to incorporate any new parameter estimates into our method.

282 **3.3 Carbon-to-chlorophyll ratio (χ) and fractions of carbon for any** 283 **size class of phytoplankton**

284 Considering that the biomass of phytoplankton (in carbon units) is the sum of biomasses
285 in n non-overlapping size classes, the carbon biomass C_{ij} of a size class defined by the size
286 (diameter) range $[D_i, D_j]$ with $0 \leq i < j \leq n$, can be expressed as the product of the
287 carbon-to-chlorophyll ratio χ_{ij} and the chlorophyll concentration B_{ij} of that size class. Using
288 Eq. (6) and the expression for B_{ij} from Roy et al. (2013), the carbon content of any size class

289 C_{ij} can be expressed as,

$$C_{ij} = \chi_{ij} B_{ij} = \chi_{ij} \left(\frac{D_j^{4-\xi-m} - D_i^{4-\xi-m}}{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}} \right) B. \quad (8)$$

290 The total phytoplankton carbon can then be expressed as a sum of phytoplankton carbon
291 from n size classes,

$$C_{total} = \sum_{i=0, j=i+1}^{i=n-1, j=n} C_{ij} = \frac{B}{D_{max}^{4-\xi-m} - D_{min}^{4-\xi-m}} \sum_{i=0, j=i+1}^{i=n-1, j=n} \left[\chi_{ij} \left(D_j^{4-\xi-m} - D_i^{4-\xi-m} \right) \right], \quad (9)$$

292 where the carbon-to-chlorophyll ratio χ_{ij} of the size class $[D_i, D_j]$ follows directly from Eq. (6),

$$\chi_{ij} = \frac{10^{-9} a (10^{18} \pi/6)^b}{(\pi/6) c_0} \left[\frac{D_j^{3b-\xi+1} - D_i^{3b-\xi+1}}{D_j^{4-\xi-m} - D_i^{4-\xi-m}} \right] \left[\frac{4 - \xi - m}{3b - \xi + 1} \right]. \quad (10)$$

293 Further, the fractional phytoplankton carbon F_{ij} within any size class $[D_i, D_j]$ can be com-
294 puted as follows:

$$F_{ij} = \frac{C_{ij}}{C_{total}} = \frac{\chi_{ij} \left(D_j^{4-\xi-m} - D_i^{4-\xi-m} \right)}{\sum_{i=0, j=i+1}^{i=n-1, j=n} \left[\chi_{ij} \left(D_j^{4-\xi-m} - D_i^{4-\xi-m} \right) \right]}. \quad (11)$$

295 In particular, if $[D_0, D_1]$, $[D_1, D_2]$ and $[D_2, D_3]$ represent the ranges of cell diameters
296 corresponding to picoplankton, nanoplankton and microplankton respectively, the carbon-to-
297 chlorophyll ratio corresponding to the three size classes (χ_p , χ_n and χ_m) can be respectively
298 computed using Eq. (10) as follows:

$$\chi_p = \frac{10^{-9} a (10^{18} \pi/6)^b}{(\pi/6) c_0} \left[\frac{D_1^{3b-\xi+1} - D_0^{3b-\xi+1}}{D_1^{4-\xi-m} - D_0^{4-\xi-m}} \right] \left[\frac{4 - \xi - m}{3b - \xi + 1} \right]; \quad (12)$$

$$\chi_n = \frac{10^{-9} a (10^{18} \pi/6)^b}{(\pi/6) c_0} \left[\frac{D_2^{3b-\xi+1} - D_1^{3b-\xi+1}}{D_2^{4-\xi-m} - D_1^{4-\xi-m}} \right] \left[\frac{4 - \xi - m}{3b - \xi + 1} \right]; \quad (13)$$

300 and

$$\chi_m = \frac{10^{-9} a (10^{18} \pi/6)^b}{(\pi/6) c_0} \left[\frac{D_3^{3b-\xi+1} - D_2^{3b-\xi+1}}{D_3^{4-\xi-m} - D_2^{4-\xi-m}} \right] \left[\frac{4 - \xi - m}{3b - \xi + 1} \right]. \quad (14)$$

301 Moreover, using equation (11) and equations (12-14), the fractions of carbon for picoplankton
 302 (F_p), nanoplankton (F_n) and microplankton (F_m) can be computed as follows:

$$F_p = \frac{\chi_p (D_1^{4-\xi-m} - D_0^{4-\xi-m})}{\left[\chi_p (D_1^{4-\xi-m} - D_0^{4-\xi-m}) + \chi_n (D_2^{4-\xi-m} - D_1^{4-\xi-m}) + \chi_m (D_3^{4-\xi-m} - D_2^{4-\xi-m}) \right]}; \quad (15)$$

$$F_n = \frac{\chi_n (D_2^{4-\xi-m} - D_1^{4-\xi-m})}{\left[\chi_p (D_1^{4-\xi-m} - D_0^{4-\xi-m}) + \chi_n (D_2^{4-\xi-m} - D_1^{4-\xi-m}) + \chi_m (D_3^{4-\xi-m} - D_2^{4-\xi-m}) \right]}; \quad (16)$$

$$F_m = \frac{\chi_m (D_3^{4-\xi-m} - D_2^{4-\xi-m})}{\left[\chi_p (D_1^{4-\xi-m} - D_0^{4-\xi-m}) + \chi_n (D_2^{4-\xi-m} - D_1^{4-\xi-m}) + \chi_m (D_3^{4-\xi-m} - D_2^{4-\xi-m}) \right]}. \quad (17)$$

303 Consistent with the previous studies (Roy et al., 2013; Sieburth et al., 1978; Vidussi et al.,
 304 2001), the diameter bounds of pico-, nano-, and micro- size classes may be taken as $D_0 = 0.2$
 305 μm , $D_1 = 2 \mu m$, $D_2 = 20 \mu m$, and $D_3 = 50 \mu m$. Applying these limits to Eq. (10), the
 306 carbon-to-chlorophyll ratios of picoplankton, nanoplankton, and microplankton can be plot-
 307 ted as functions of the exponent of the phytoplankton size spectrum as in Fig. (2a). Com-
 308 pared with the carbon-to-chlorophyll ratio of the mixed population (black curve, Fig. 2a),
 309 carbon-to-chlorophyll ratio of picoplankton (blue curve, Fig. 2a) is always higher, but that
 310 of microplankton (red curve, Fig. 2a) is always lower, over the range values of ξ . On the
 311 other hand, the carbon-to-chlorophyll ratio of the nanoplankton (green curve, Fig. 2a) is less
 312 than that of the mixed population for low values of ξ , and is greater than that of the mixed
 313 population for the high values of ξ (Fig. 2a). The range of variation of carbon-to-chlorophyll
 314 ratio is the minimum for micro-size class, and maximum for pico-size group (Fig. 2a).

315 The proportions of phytoplankton carbon corresponding to the three size classes, when
 316 plotted as functions of ξ (the solid blue, green and red lines corresponding to pico-, nano-
 317 and micro classes, respectively in Fig. 2b), have shapes similar to those obtained for the

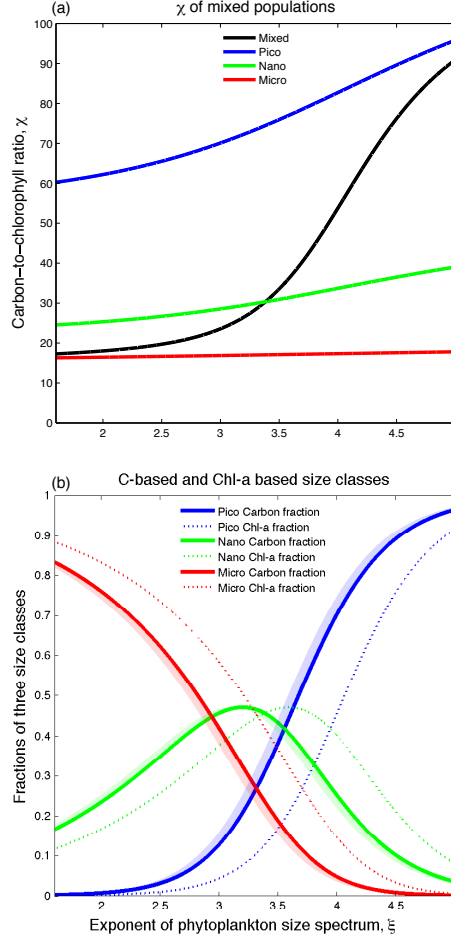


Figure 2: Carbon-to-chlorophyll ratio χ and phytoplankton carbon derived for various size classes of phytoplankton. (a) χ of mixed phytoplankton (black line, using Eq. 6), picoplankton (blue line, using Eq. 12), nanoplankton (green line, using Eq. 13) and microplankton (red line, using Eq. 14) plotted as functions of the exponent of phytoplankton size spectrum ξ . (b) Size-fractionated phytoplankton carbon and chlorophyll plotted as functions of ξ . The solid blue, green and red lines represent the fractions of phytoplankton carbon corresponding to picoplankton (using Eq. 15), nanoplankton (using Eq. 16) and microplankton (using Eq. 17); and the shaded area represents the corresponding ranges of carbon fraction. The dotted blue, green and red lines represent fractions of chlorophyll corresponding to picoplankton, nanoplankton and microplankton, as derived in Roy et al. (2013).

318 chlorophyll-proportions (the dotted lines, Fig. 2b, based on Roy et al., 2013). However, over
319 the range of ξ relevant for phytoplankton, the fraction of microplankton based on carbon is
320 lower than that based on chlorophyll (the solid and dotted red lines, Fig. 2b); and the fraction
321 of picoplankton based on carbon is higher than that based on chlorophyll (the solid and dotted
322 blue lines, Fig. 2b). On the other hand, the fraction of nanoplankton based on carbon is higher
323 than that based on chlorophyll for low values of ξ , but the relationship is reversed for higher
324 values of ξ (the solid and dotted green lines, Fig. 2b). We also note that the uncertainties in
325 allometric parameters result in relatively low uncertainties in the estimates of carbon-based
326 fractions of pico, nano and micro size classes (the blue, green and red shaded areas associated
327 with the corresponding solid lines in Fig. 2b indicate these uncertainties).

328 4 Results and discussion

329 4.1 Algorithm validation using *in situ* data

330 Ideally, it would require a large global dataset of *in situ* phytoplankton carbon to validate the
331 bio-optical method presented here. However, constraints on the availability of *in situ* data on
332 phytoplankton carbon limit the possibilities for algorithm validation. Nevertheless, we have
333 attempted a validation exercise using the available flow cytometry data on phytoplankton
334 compiled and reported by Buitenhuis et al. (2012) as contribution to the MAREDAT World
335 Ocean Atlas of Plankton Functional Types database. However, this database reported phy-
336 toplankton carbon for the pico-size group only, from 1980 onwards, over the world ocean.

337 Therefore, the validation exercise presented here is limited to *in situ* data on pico-size class,
338 and the statistics may not apply to phytoplankton carbon in other size classes or to the total
339 phytoplankton carbon, which would be a sum of carbon in all size classes. To maximise the
340 number of data points for validation, we have used the reported pico-carbon data over the
341 entire period of satellite coverage i.e., from September 1997 to December 2013, consistent
342 with the OC-CCI v2 satellite data. Given the short time-scale of phytoplankton turn over,
343 the satellite and *in situ* match-up would be most optimal on a daily scale. Compared with
344 the weekly or monthly products, the choice of daily products would minimise the possible
345 uncertainties that might arise due to time differences between the *in situ* and satellite ob-
346 servations. We thus computed pico-plankton carbon using our method on the daily maps,
347 and retrieved the spatially matched-up data points, which provided ~ 900 data points for
348 validation of pico-carbon.

349 The locations of the *in situ* measurements from the MAREDAT database taken for this
350 study are shown in Fig. (3a), and the validation results are shown in Fig. (3b-d). On a
351 linear scale, the Spearman's correlation (ρ) between the *in situ* picoplankton carbon and the
352 satellite-derived estimates of pico-carbon (in mgC m^{-3}) computed by our method is 0.57, $p <$
353 0.0001, where the root-mean squared error (RMSE) of the satellite-based estimates is 36.23
354 mgC m^{-3} . The data-density plot shows high density (red colour) of sample points below the
355 1:1 line (black line in Fig. 3b) suggesting that the satellite-derived picoplankton carbon values
356 are lower than the corresponding *in situ* estimates, and on a linear scale the bias is -13.53
357 mgC m^{-3} .

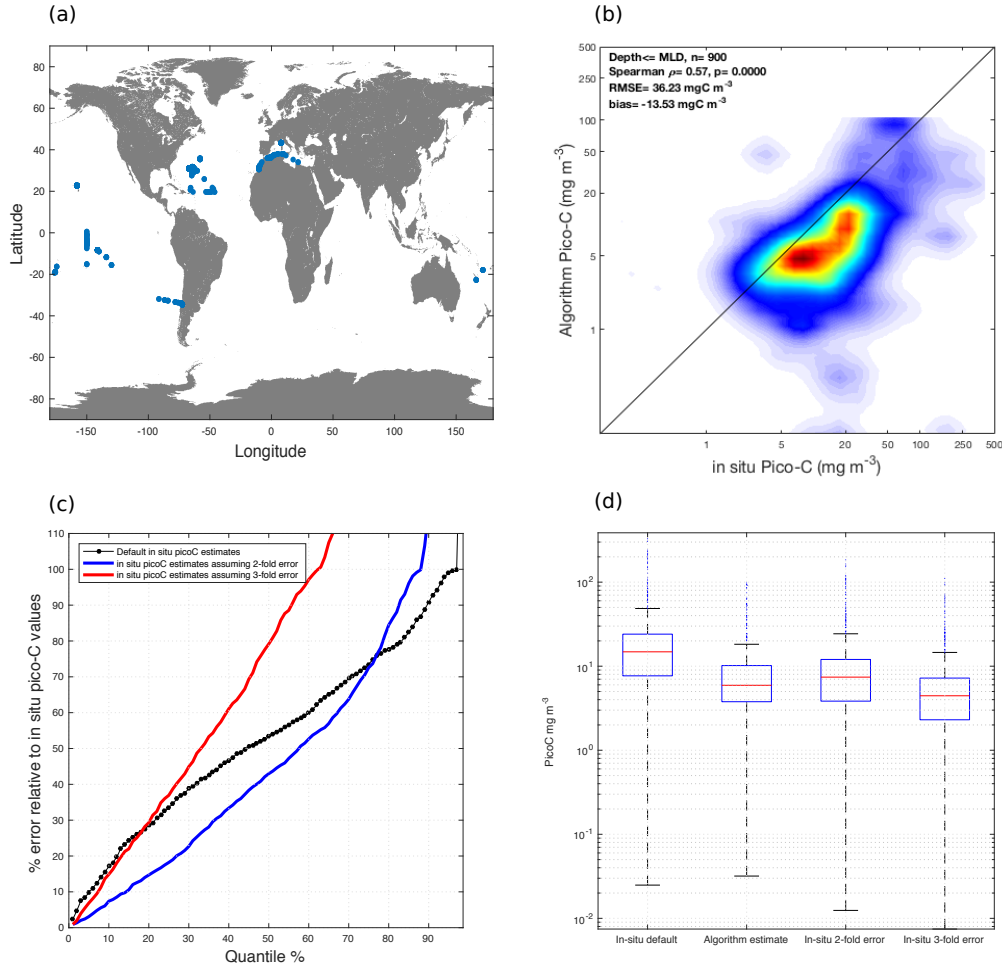


Figure 3: Validation of the computed phytoplankton carbon using *in situ* data. (a) Locations of the *in situ* data, which is a part of *in situ* samples from MAREDAT within the range of satellite coverage, i.e, 1997-2013. The compiled dataset represents *in situ* measurements of carbon for small-phytoplankton, $< 2 \mu\text{m}$ in diameter. Phytoplankton carbon for the corresponding size range was computed using Eq. (8) and Eq. (10). (b) Comparison plot for the observed and computed picoplankton carbon. The increased densities of the data points around the 1:1 line are evident in the high density (red colour) close to the 1:1 line. Lower densities are shown in blue. (c) Magnitudes of the relative error (in %) in estimation of picocarbon with respect to the reported *in situ* values presented for data quantiles. The black line indicates the error percentages for the default *in situ* values, whereas the blue and red lines show those for assumed 2 and 3 fold overestimation in the in situ calculations (these possibilities in MAREDAT are discussed by Buitenhuis et al. (2012)). (d) Box plots of the observed and satellite-derived values of picoplankton carbon corresponding to the default *in situ* values, and *in situ* values with possibilities of 2 and 3 fold overestimations.

358 The apparent underestimation of picoplankton carbon by the method presented here may
359 be due to uncertainties in satellite input, the allometric parameterisation or the uncertain-
360 ties in the *in situ* estimates. In particular, the *in situ* pico-carbon values in MAREDAT
361 were calculated assuming a set of fixed values of carbon per cell for the three picoplankton
362 species considered, and so the overall pico-carbon estimates from the *in situ* data are sub-
363 ject to uncertainties related to the cell-to-carbon conversion factors. Buitenhuis et al. (2012)
364 acknowledged that "there is considerable uncertainty in the conversion factors" (see Table 2
365 in Buitenhuis et al. (2012) for the ranges of conversion factors) in the reported MAREDAT
366 pico-carbon data, and further suggested that this factor may lead to significant overestima-
367 tion of *in situ* picoplankton carbon, which on a global scale could contribute to "a 2-3 fold
368 difference in the estimated picophytoplankton biomass" (Buitenhuis et al., 2012).

369 Taking these uncertainties in the *in situ* estimates into consideration, we have investigated
370 the uncertainty bounds for the satellite-based estimates: Fig. 3c shows three scenarios of the
371 percentages of error in the satellite-derived estimates relative to the *in situ* values over the
372 data quantiles. Corresponding to the default (reported) *in situ* estimates, the magnitude
373 of the relative errors in satellite-derived estimates are $< 34\%$ for a quartile of the data,
374 and $< 72\%$ for the three quartiles of the data (black line, in 3c). This scenario changes
375 significantly if the possible uncertainties in the *in situ* values are taken into account: for
376 example, corresponding to an overall 2-fold (or 3-fold) overestimation in the *in situ* data,
377 satellite-derived estimates are $< 18\%$ (or $< 35\%$) for a quartile of the data, and $< 70\%$ (or
378 $> 100\%$) for the three quartiles of the data (the blue and red lines, respectively, in 3c). Also,

379 the box plots (Fig. 3d) show that the median values and the spread and distributions of the
380 estimated and *in situ* picoplankton values differ between the default *in situ* values and the
381 alternative two scenarios: the median value of the estimated pico-carbon is lower than that for
382 the default *in situ* estimates, but the difference reduces considerably if we take into account
383 the possibilities of a 2-fold or 3-fold overestimation of the *in situ* pico-carbon, and in fact,
384 corresponding to a 3-fold *in situ* overestimation, the median of the satellite estimates is higher
385 than those for the *in situ* estimates (Fig. 3d).

386 Therefore, our satellite-based estimates show underestimation of picoplankton carbon with
387 respect to the reported *in situ* estimates, but the level of bias of the current estimates is also
388 subject to the uncertainties in the carbon-per-cell conversion factors applied to the *in situ*
389 data. The validation might also have been affected by the properties of the statistical dis-
390 tribution of the quantities under comparison; for example, the *in situ* picoplankton-carbon
391 data were computed in MAREDAT under the assumption of mean cell-to-carbon conversion
392 factors, whereas the algorithm, by design, considered the median of a number of allometric
393 relationships drawn from the literature for different taxa. So, the possibility of non-normality
394 in the *in situ* picoplankton-carbon distribution would impose a systematic bias, when con-
395 sidering the mean over the median. However, re-calculation of the *in situ* pico-carbon from
396 MAREDAT database to explore the unknown error characteristics is beyond the scope of our
397 study. The other sources of uncertainties in pico-carbon may also be associated with the un-
398 certainties in the satellite input, e.g., here we have used OC-CCI-version-2 data, which have
399 been re-processed with a view to reducing uncertainties. However, a new version of the data

400 (OC-CCI-version-3) has been released only recently, but we are yet to apply our method to
401 the updated version of the data. We further note that the RMSE and bias values presented
402 are based on picoplankton carbon data only, and uncertainties in phytoplankton carbon for
403 other classes would require further investigation.

404 4.2 C:Chl (χ) and phytoplankton carbon over global ocean

405 The average carbon-to-chlorophyll ratios (χ) computed over the global ocean using composite
406 monthly images from September 1997 to December 2013 vary over a wide range, from <20
407 in the coastal or case-2 waters to >90 in the open ocean and case-1 waters (Fig. 4a). These
408 results are consistent with our understanding that the low and high values of χ represent,
409 respectively, the areas dominated by large and small phytoplankton. The annually-averaged
410 standing stocks of phytoplankton carbon over the mixed layer vary from less than 1 mg m^{-3}
411 in the gyres to more than 500 mg m^{-3} in the case-2 and coastal waters (Fig. 4b). The stock
412 of phytoplankton carbon integrated over the mixed layer and globe is found to be ~ 0.26
413 GtC (Fig. 4b), with some monthly variation in the stock ranging from 0.24 to 0.29 GtC. The
414 smallest stock is observed in June with ~ 0.24 GtC and a maximum in September ~ 0.29 GtC,
415 with the autumn months having stocks of carbon greater than the annual average.

416 In a recent study, Kostadinov et al. (2016) have shown that the estimates of annual stock
417 of phytoplankton carbon depend on the estimation method, and can vary from 0.2 to 0.32
418 GtC, with the minimum due to Stramski et al. (2008): ~ 0.2 GtC, followed by the average
419 of some CMIP5 models (Taylor et al., 2012): ~ 0.22 GtC, Kostadinov et al. (2016): ~ 0.24

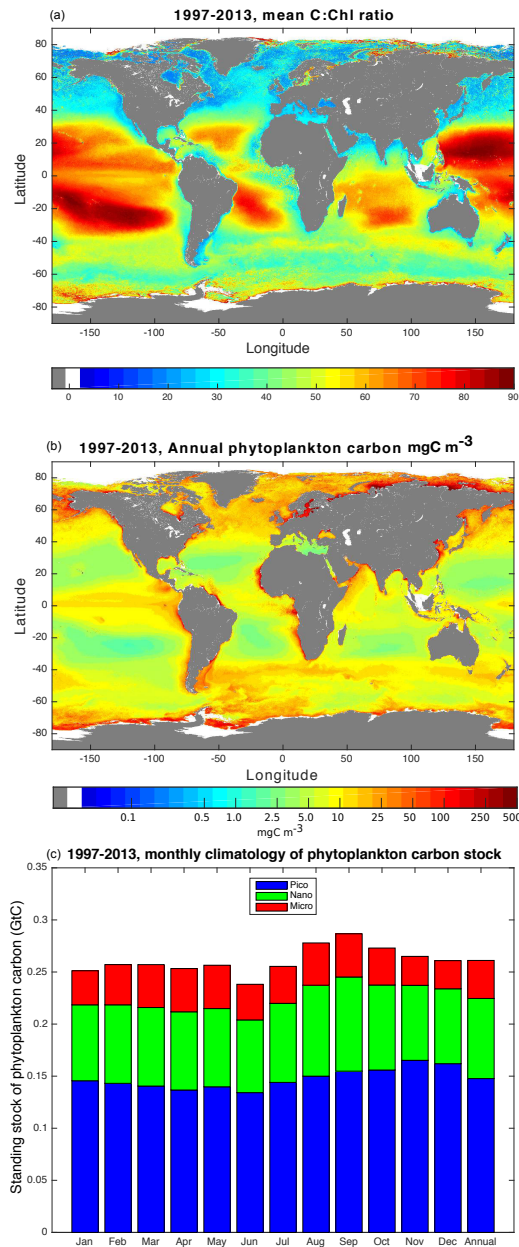


Figure 4: Global distribution of C:Chl and phytoplankton carbon estimated over 1997-2013 using monthly OC-CCI data. (a) Annual climatology of C:Chl over 1997-2013. (b) Annual climatology of phytoplankton carbon in the surface over 1997-2013. (c) Monthly climatology of the global estimates of phytoplankton carbon (in giganon, GtC) integrated over the mixed-layer depth. Blue, green and red colours indicate the proportions of phytoplankton carbon corresponding to pico-, nano- and micro- size groups.

420 GtC (with a range of 0.2 to 0.3 GtC) and Behrenfeld et al. (2005): ~ 0.32 GtC. Our estimate
421 of ~ 0.26 GtC (with a range of monthly variations between 0.24 and 0.29 GtC) is slightly
422 higher than those of Stramski et al. (2008), the average result for CMIP5 models reported by
423 Kostadinov et al. (2016), and the back-scattering-based method of Kostadinov et al. (2016),
424 but is lower than that reported by Behrenfeld et al. (2005).

425 **4.3 Size-partitioned phytoplankton carbon over the global ocean**

426 Using the equations derived in Section 3.3, phytoplankton carbon can be partitioned into any
427 number of size classes, and in particular, into the three broad size classes, e.g., pico, nano,
428 and micro. The annual average of phytoplankton carbon in the three size classes expressed
429 both as the percentages of total phytoplankton and in the units of mgC m^{-3} are shown in
430 Fig. (5).

431 The global distributions of the carbon-based phytoplankton size classes (i.e., the percent-
432 ages of carbon in three size classes in Fig. 5a-c), are generally similar to the corresponding
433 chlorophyll-based distributions reported in Roy et al. (2013). Pico-carbon stocks generally
434 dominate over those of nano- and micro-carbon for most of the open oceans, including the
435 gyres and the equatorial regions, with contributions ranging from $\sim 70\%$ to more than 90%
436 of total phytoplankton carbon (Fig. 5a). Converting the percentages into units of carbon,
437 the concentration of picoplankton carbon in these areas is generally within the range of 1-10
438 mgC m^{-3} (Fig. 5d). In most of the coastal waters and generally in the northern hemisphere,
439 the pico-carbon stocks are around 10-20% of the total phytoplankton carbon (Fig. 5a); how-

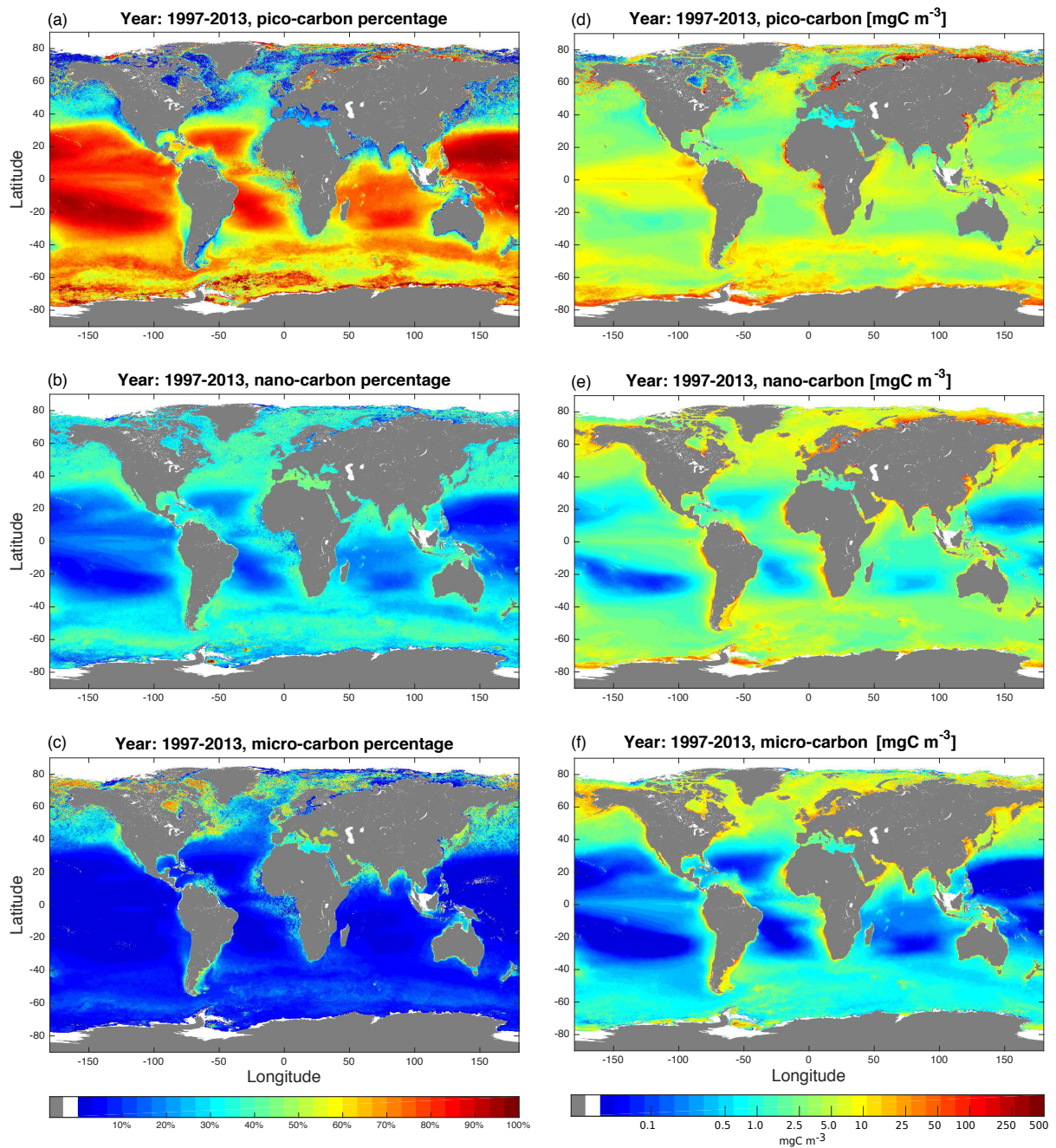


Figure 5: Global average distribution of phytoplankton carbon corresponding to pico-, nano- and micro- size groups estimated by averaging monthly values computed from OC-CCI data for the period September 1997 - December 2013. Carbon-based size classes of phytoplankton: Fractional (%) contributions of (a) picoplankton carbon, (b) nanoplankton carbon and (c) microplankton carbon to total phytoplankton carbon. Estimates of the concentrations of (d) picoplankton carbon, (e) nanoplankton carbon and (f) microplankton carbon in the surface in mgC m^{-3} .

440 ever, the range of pico-carbon may vary from 2 mgC m^{-3} to more than 100 mgC m^{-3} (Fig. 5d).
441 The stocks of nano-plankton carbon are 10-15% of total phytoplankton carbon in equatorial
442 gyres, and go up to 40-45% in the southern ocean, northern hemisphere and coastal oceans
443 (Fig. 5b). These percentages account for $\sim 2\text{-}3 \text{ mg m}^{-3}$ of nano-carbon in the equatorial gyres,
444 and $\sim 10\text{-}30 \text{ mgC m}^{-3}$ in the northern and southern oceans (Fig. 5e). The stocks of micro-
445 carbon, on the other hand, are estimated to be less than 20% in most of the equatorial and
446 southern ocean, except the coastal regions, and in the northern hemisphere, where its percent-
447 age contribution goes up to 70-80% (Fig. 5c). In the coastal oceans and northern hemisphere,
448 the concentration of micro-plankton carbon is estimated to be in the range $20\text{-}30 \text{ mgC m}^{-3}$,
449 whereas in the equatorial gyres it is below 0.5 mgC m^{-3} (Fig. 5f).

450 The global distributions of the size-partitioned phytoplankton carbon can be spatially inte-
451 grated over the mixed-layer depth to estimate their annual-mean stocks, which are $\sim 0.14 \text{ GtC}$
452 for picoplankton (with a monthly range of $0.13\text{-}0.16 \text{ GtC}$), $\sim 0.08 \text{ GtC}$ for nanoplankton (with
453 a monthly range of $0.07\text{-}0.09 \text{ GtC}$) and $\sim 0.04 \text{ GtC}$ for microplankton (with a monthly range
454 of $0.03\text{-}0.041 \text{ GtC}$) (Fig. 4c). These stocks of carbon in the three size classes constitute ap-
455 proximately 54% (with a monthly range of 53-62%), 31% (with a monthly range of 27-32%)
456 and 15% (with a monthly range of 10-16%) of the global stock of phytoplankton carbon,
457 respectively.

458 4.4 Sources and estimates of uncertainty

459 The estimates of phytoplankton carbon from the bio-optical algorithm presented here would
460 be subject to uncertainties from two sources: uncertainties associated with the remote sensing
461 products (chlorophyll-a and phytoplankton absorption, and hence satellite-derived values of
462 ξ); and the uncertainties in allometric parameterisation in the bio-optical model; but the two
463 uncertainty sources are independent of each other. We consider an overall uncertainty in ξ
464 arising from the uncertainties in satellite chlorophyll-a and phytoplankton absorption (based
465 on the uncertainty calculations by Roy et al., 2013). We then compute from Eq. (7) the
466 total relative sensitivity of the estimated phytoplankton carbon (i.e., $\frac{\Delta C_{total}}{C_{total}}$), as a combined
467 function of the individual relative sensitivities $\frac{\Delta \xi}{\xi}$, $\frac{\Delta a}{a}$, and $\frac{\Delta b}{b}$. In the following, we apply the
468 above sensitivity analysis to understand the uncertainties in the estimation. The uncertainties
469 presented below should be interpreted as model-based uncertainties; and not as those based
470 on the *in situ* observations (which was not possible due to lack of the size-partitioned data
471 on phytoplankton carbon).

472 The overall uncertainties in the estimates of phytoplankton carbon due to 0-25% un-
473 certainties in ξ (chosen based on Roy et al., 2013) and 20% uncertainties in the allometric
474 parameters a and b are presented on a contour map in Fig. (6a). The uncertainty level in
475 phytoplankton carbon is typically <30% over the range of ξ typically encountered at sea,
476 except for ξ values between 3.5 and 4, where the uncertainties can amplify up to 80-90%
477 corresponding to >20% uncertainty in satellite-derived ξ values (Fig. 6a). In other words, for
478 phytoplankton populations that are clearly dominated by either small cells (higher end of ξ)

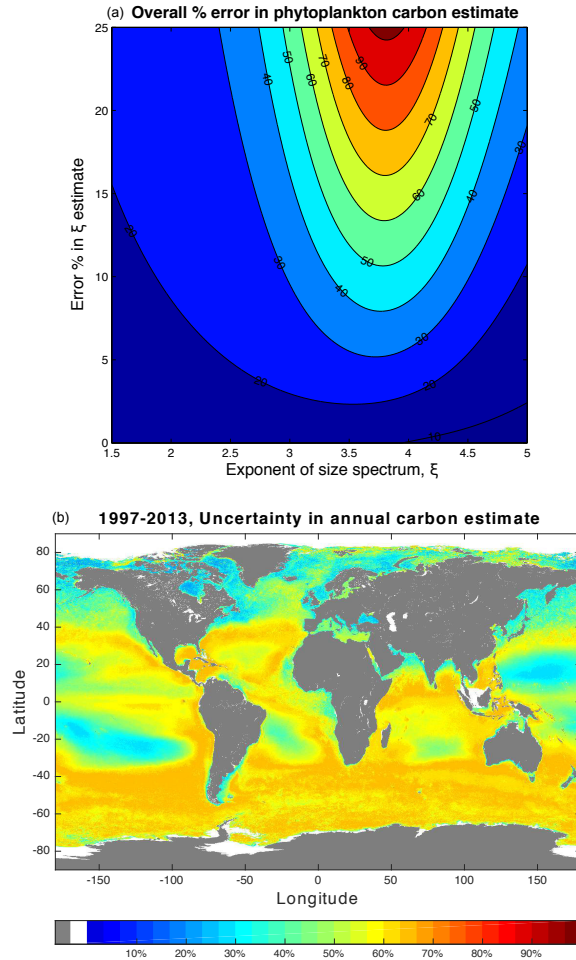


Figure 6: Level of uncertainties in phytoplankton carbon computed by the method proposed here. (a) Uncertainty in phytoplankton carbon estimates due to possible errors in estimating ξ (the exponent of phytoplankton size spectrum) and b (the exponent of allometric carbon relationship). The overall uncertainties in the estimates of phytoplankton carbon are shown over a possible uncertainty range 0-25% for ξ and an uncertainty level 20% for b . (b) Propagated uncertainties in the estimates of phytoplankton carbon corresponding to 25% uncertainty in ξ and 20% uncertainty in b over the global ocean for the period of 1997-2013.

479 or large cells (lower end of ξ), the uncertainties in estimating phytoplankton carbon will be
480 low (20-30%), but, for populations with no obvious dominance by large or small cells, the
481 uncertainties can be high (>30%).

482 On the global map, the propagation of uncertainties in phytoplankton carbon correspond-
483 ing to the higher ends of uncertainties in ξ (say, 25%), a and b (say, 20%) is presented for
484 1997-2013 (Fig. 6b). In most of the Northern hemisphere, in the subtropical gyres and in
485 the coasts, the uncertainties in phytoplankton carbon are within a range of 20-40% (Fig. 6b).
486 However, uncertainties in the Southern Ocean, and parts of Atlantic Ocean can go up to
487 50-70% (Fig. 6b). The lower and upper levels of the annual stocks of phytoplankton carbon
488 arising from regional-level uncertainties may vary between 0.12 GtC and 0.35 GtC; and those
489 for pico-, nano- and micro- carbon may vary in the ranges of [0.07, 0.2], [0.03, 0.09] and [0.01
490 0.04] GtC, respectively (Fig. 7). The monthly variations of the stocks are also remarkable:
491 the possibility exists of pico-carbon stock being larger or smaller than the default estimates,
492 whereas for microplankton-carbon, the uncertainties tend to lower the estimates, as evident
493 when taking into account regional uncertainties in phytoplankton carbon (Fig. 7).

494 5 Concluding remarks

495 Estimates of total concentration of carbon in phytoplankton and its fractions in various phyto-
496 plankton size classes from satellite-remote sensing can provide valuable information for ocean
497 biogeochemical and carbon-cycle research. However, the work in this direction has been ham-
498 pered by the absence of a remote-sensing signal that can be related directly to phytoplankton

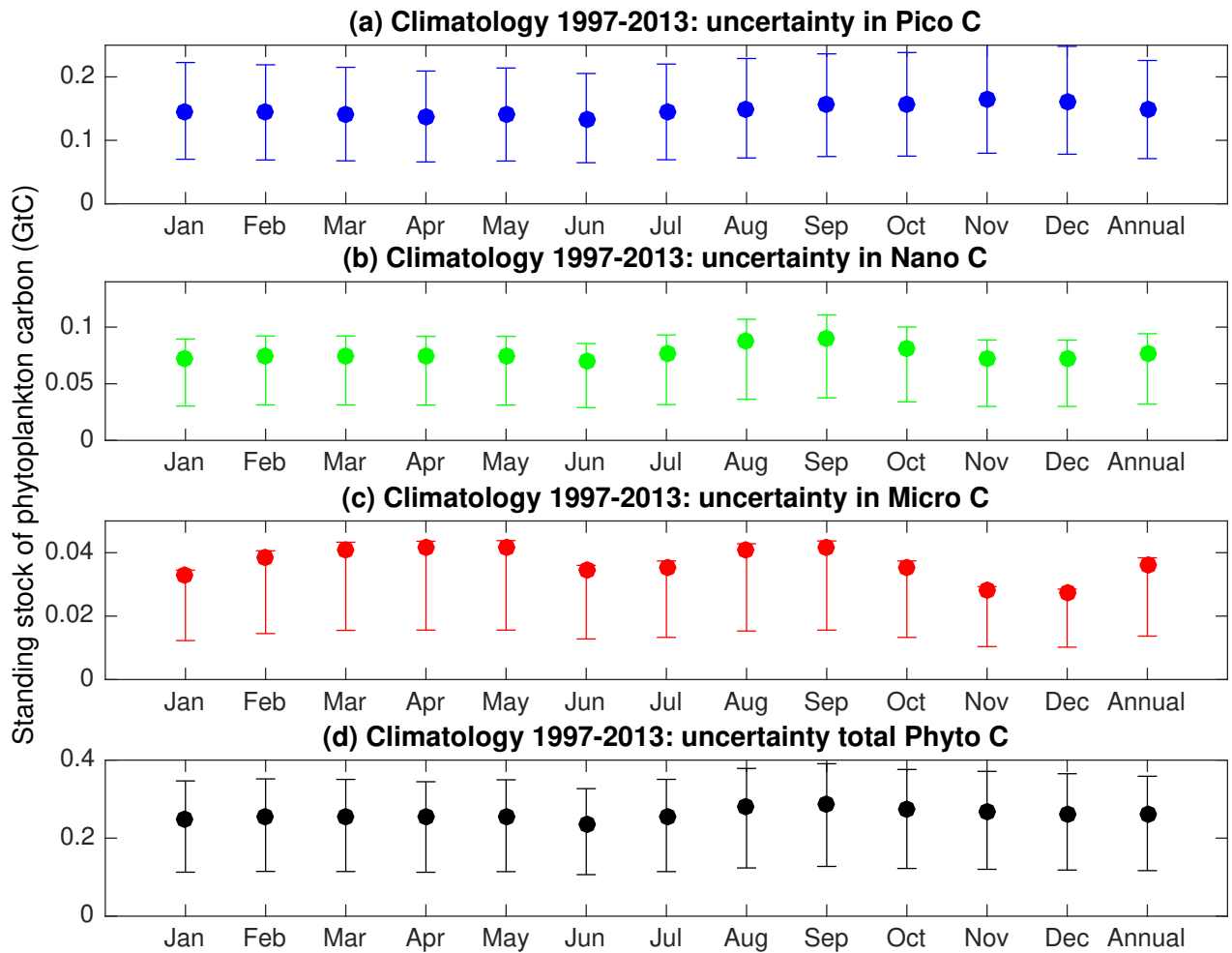


Figure 7: Estimates of uncertainties in the monthly and annual standing stocks of phytoplankton carbon. Monthly and annual climatologies of the standing stocks of (a) picoplankton carbon, (b) nanoplankton carbon, (c) microplankton carbon and (d) total phytoplankton carbon, plotted along with their corresponding uncertainty ranges (represented by vertical error bars) estimated assuming possible uncertainties in ξ and b parameterisation as in Fig. 6

499 carbon. Only a small number of studies have addressed this problem, and all the methods
500 proposed so far (Behrenfeld et al., 2005; Kostadinov et al., 2016; Stramski et al., 2008) have
501 relied on relating POC to back-scattering or to remote-sensing reflectance, and then ascribing
502 a fixed fraction of POC to phytoplankton. Though these approaches have met with reasonable
503 success, their weakness lies in the natural variability in the ratio of phytoplankton carbon to
504 POC, which the algorithms cannot account for.

505 Here we present a novel bio-optical algorithm that uses the absorption coefficient of phyto-
506 plankton from remote sensing along with the allometric relationship of cellular carbon content
507 to compute carbon-to-chlorophyll ratio, the standing stocks of phytoplankton carbon, and the
508 carbon-based proportions of phytoplankton size classes, in the global ocean. The basis of the
509 method is the bio-optical algorithm developed by Roy et al. (2013) to compute the exponent
510 of the phytoplankton size spectrum and the chlorophyll proportions at various size classes
511 from the absorption coefficient of phytoplankton in the red part of the absorption spectrum.
512 Extending the method of Roy et al. (2013), we have derived analytical expressions for combin-
513 ing phytoplankton absorption from remote sensing with the allometric relationship between
514 cell size and phytoplankton carbon. The new expressions enable computation of phytoplank-
515 ton carbon from satellite remote sensing based on the bio-optical fingerprints of the living
516 phytoplankton alone. By design, this absorption-based method does not rely on a systematic
517 relationship between phytoplankton carbon and POC (such as a constant ratio), as required
518 by the other methods that are available at present (Behrenfeld et al., 2005; Kostadinov et al.,
519 2016). Instead, by combining the estimates of phytoplankton carbon, based on the absorption

520 coefficient of pigment-containing phytoplankton cells (presented here), with the estimates of
521 POC from back-scattering or remote-sensing reflectances (Behrenfeld et al., 2005), we can
522 arrive at independent estimates of the ratio of phytoplankton carbon to POC. Such estimates
523 would be an immediate application of the method proposed here.

524 We have used the new method to compute phytoplankton carbon in the global ocean
525 on a monthly basis for the 1997-2013 period using OC-CCI time series data, and computed
526 monthly climatologies of the standing stock of phytoplankton carbon in the mixed layer, and
527 their annual averages. The new results are of the same order of magnitude, and comparable
528 with, those reported earlier (Behrenfeld et al., 2005; Kostadinov et al., 2016; Stramski et al.,
529 2008), though there are regional and seasonal differences. We have provided the RMSE and
530 bias of the estimates with respect to the *in situ* measurements of the picoplankton carbon, but
531 due to the unavailability of *in situ* data, we have been unable to estimate the uncertainties,
532 RMSE or bias for other phytoplankton size classes (e.g., micro- or nano- phytoplankton).
533 We also recognize that, as additional data become available, it would be interesting and
534 useful to carry out extensive inter-comparisons among the various methods for estimating
535 phytoplankton carbon.

536 With the availability of a variety of satellite-derived products, it has become increasingly
537 important to understand and quantify uncertainties associated with these products. For
538 example, the Global Climate Observing System (GCOS) has provided requirements for accu-
539 racy in ocean-colour data that can be used for climate studies (GCOS, 2011). Because our
540 method for estimation of carbon is semi-analytical, it is possible, as shown here, to quantify

541 analytically the uncertainties in carbon estimates, provided that the uncertainties in satellite-
542 derived chlorophyll and absorption coefficient are known. For illustration, we have provided
543 estimates of the uncertainties corresponding to 30% overall uncertainty (GCOS requirement)
544 in the satellite input, and we have identified the oceanographic regions where the carbon
545 estimates will be less (or more) sensitive to uncertainties in the inputs. These calculations
546 also provide insight into the error characteristics of phytoplankton carbon estimated by our
547 method, and suggest that the errors do not generally amplify, and that they become less for
548 more accurate retrievals of the satellite-based inherent optical properties. Another source of
549 uncertainty is the allometric parametrisation, and any change in the allometric parameters
550 would alter our estimates of phytoplankton carbon (as shown in the sensitivity results). How-
551 ever, implementation of any improved allometric parametrisation within this method would
552 be straight forward. Finally, we note that the uncertainties in the estimates of carbon in the
553 coastal oceans and at high latitudes may be high due to several reasons, e.g., high concen-
554 tration of CDOM, solar zenith angles, clouds or ice; and so this method, like many other
555 ocean colour algorithms, will be generally applicable to open oceans. Further investigations
556 should address its applicability to optically complex waters, and oceanic regions with complex
557 phytoplankton community structure, e.g., blooms of large chain-forming diatoms.

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568 References

- 569 Behrenfeld, M. J., Boss, E., Siegel, D. A. and Shea, D. M. (2005), ‘Carbon-based ocean
570 productivity and phytoplankton physiology from space’, *Global biogeochemical cycles* **19**(1).
- 571 Belo Couto, A., Brotas, V., Mélin, F., Groom, S. and Sathyendranath, S. (2016), ‘Inter-
572 comparison of oc-cci chlorophyll-a estimates with precursor data sets’, *International Journal*
573 *of Remote Sensing* **37**(18), 4337–4355.
- 574 Brewin, R. J., Sathyendranath, S., Müller, D., Brockmann, C., Deschamps, P.-Y., Devred,
575 E., Doerffer, R., Fomferra, N., Franz, B., Grant, M. et al. (2015), ‘The ocean colour climate
576 change initiative: Iii. a round-robin comparison on in-water bio-optical algorithms’, *Remote*
577 *Sensing of Environment* **162**, 271–294.
- 578 Buitenhuis, E. T., Li, W. K., Vaultot, D., Lomas, M. W., Landry, M., Partensky, F., Karl, D.,

579 Ulloa, O., Campbell, L., Jacquet, S. et al. (2012), ‘Picophytoplankton biomass distribution
580 in the global ocean’, *Earth System Science Data* **4**(1), 37–46.

581 Carder, K. L., Chen, F., Lee, Z., Hawes, S. and Kamykowski, D. (1999), ‘Semianalytic
582 moderate-resolution imaging spectrometer algorithms for chlorophyll a and absorption
583 with bio-optical domains based on nitrate-depletion temperatures’, *Journal of Geophys-
584 ical Research-Oceans* **104**(C3), 5403–5421.

585 DuRand, M. D., Olson, R. J. and Chisholm, S. W. (2001), ‘Phytoplankton population dy-
586 namics at the bermuda atlantic time-series station in the sargasso sea’, *Deep Sea Research
587 Part II: Topical Studies in Oceanography* **48**(8), 1983–2003.

588 Duyens, L. (1956), ‘The flattering of the absorption spectrum of suspensions, as compared to
589 that of solutions’, *Biochimica et Biophysica Acta* **19**, 1–12.

590 Eppley, R. W., Chavez, F. P. and Barber, R. T. (1992), ‘Standing stocks of particulate carbon
591 and nitrogen in the equatorial pacific at 150 w’, *Journal of Geophysical Research: Oceans*
592 **97**(C1), 655–661.

593 Falkowski, P. (2012), ‘The power of plankton’, *Nature* **483**, S17–S20.

594 Field, C. B., Behrenfeld, M. J., Randerson, J. T. and Falkowski, P. (1998), ‘Primary
595 production of the biosphere: integrating terrestrial and oceanic components’, *Science*
596 **281**(5374), 237–240.

597 GCOS, G. (2011), ‘Systematic observation requirements for satellite-based products for cli-
598 mate. 2011 update supplement details to the satellite 39 based component of the imple-

599 mentation plan for the global observing system for climate in support of the unfccc (2010
600 update)', Tech. rep., World Meteorological Organisation (WMO), 7 bis, avenue de la Paix,
601 CH- 1211 Geneva 2, Switzerland.

602 Geider, R. J. (1987), 'Light and temperature dependence of the carbon to chlorophyll a ratio
603 in microalgae and cyanobacteria: implications for physiology and growth of phytoplankton',
604 *New Phytologist* **106**(1), 1–34.

605 Geider, R. J., MacIntyre, H. L. and Kana, T. M. (1998), 'A dynamic regulatory model of
606 phytoplanktonic acclimation to light, nutrients, and temperature', *Limnology and Oceanog-*
607 *raphy* **43**(4), 679–694.

608 Gundersen, K., Orcutt, K. M., Purdie, D. A., Michaels, A. F. and Knap, A. H. (2001),
609 'Particulate organic carbon mass distribution at the bermuda atlantic time-series study
610 (bats) site', *Deep Sea Research Part II: Topical Studies in Oceanography* **48**(8), 1697–1718.

611 IOCCG, ed. (2014), *Phytoplankton functional types from Space.*, number 15, International
612 Ocean-Colour Coordinating Group, Reports of the International Ocean-Colour Coordinat-
613 ing Group (IOCCG).

614 Kostadinov, T., Milutinovic, S., Marinov, I. and Cabré, A. (2016), 'Carbon-based phytoplank-
615 ton size classes retrieved via ocean color estimates of the particle size distribution', *Ocean*
616 *Science Discussions* **12**, 561–575.

617 Marañón, E. (2008), 'Inter-specific scaling of phytoplankton production and cell size in the
618 field', *Journal of Plankton Research* **30**(2), 157–163.

- 619 Marañón, E., Cermeño, P., López-Sandoval, D. C., Rodríguez-Ramos, T., Sobrino, C., Huete-
620 Ortega, M., Blanco, J. M. and Rodríguez, J. (2013), ‘Unimodal size scaling of phytoplankton
621 growth and the size dependence of nutrient uptake and use’, *Ecology letters* **16**(3), 371–379.
- 622 Marañón, E., Cermeño, P., Rodríguez, J., Zubkov, M. V. and Harris, R. P. (2007), ‘Scaling of
623 phytoplankton photosynthesis and cell size in the ocean’, *Limnol. Oceanogr* **52**(5), 2190–
624 2198.
- 625 Menden-Deuer, S. and Lessard, E. J. (2000), ‘Carbon to volume relationships for dinoflagel-
626 lates, diatoms, and other protist plankton’, *Limnology and Oceanography* **45**(3), 569–579.
- 627 Müller, D., Krasemann, H., Brewin, R. J., Brockmann, C., Deschamps, P.-Y., Doerffer, R.,
628 Fomferra, N., Franz, B. A., Grant, M. G., Groom, S. B. et al. (2015), ‘The ocean colour
629 climate change initiative: II. spatial and temporal homogeneity of satellite data retrieval due
630 to systematic effects in atmospheric correction processors’, *Remote Sensing of Environment*
631 **162**, 257–270.
- 632 Oubelkheir, K., Claustre, H., Sciandra, A. and Babin, M. (2005), ‘Bio-optical and biogeochem-
633 ical properties of different trophic regimes in oceanic waters’, *Limnology and oceanography*
634 **50**(6), 1795–1809.
- 635 Peters, R. H. (1983), *The Ecological Implications of Body Size*, Cambridge University Press,
636 Cambridge.
- 637 Redalje, D. and Laws, E. (1981), ‘A new method for estimating phytoplankton growth rates
638 and carbon biomass’, *Marine Biology* **62**(1), 73–79.

639 Roy, S., Sathyendranath, S., Bouman, H. and Platt, T. (2013), ‘The global distribution of
640 phytoplankton size spectrum and size classes from their light-absorption spectra derived
641 from satellite data’, *Remote Sensing of Environment* **139**, 185–197.

642 Roy, S., Sathyendranath, S. and Platt, T. (2011), ‘Retrieval of phytoplankton size from bio-
643 optical measurements: theory and applications’, *Journal of The Royal Society Interface*
644 **8**(58), 650–660.

645 Sathyendranath, S., Brewin, R., Brockmann, C., Brotas, V., Ciavatta, S., Chuprin, A., Couto,
646 A., Doerffer, R., Dowell, M., Grant, M., Groom, S., Horseman, A., Jackson, T., Krasemann,
647 H., Lavender, S., Martinez Vicente, V., Mélin, Moore, T., Müller, D., Regner, P., Roy, S.,
648 Steinmetz, F., Swinton, J., Taberner, M., Thompson, A., Valente, A., Zühlke, M., Brando,
649 V., Feldman, G., Franz, B., Frouin, R., Gould, Jr, R., Hooker, S., Kahru, M., Mitchell,
650 M., Muller-Karger, F., Sosik, H., Voss, K., Werdell, J. and Platt, T. (2016), ‘Creating
651 an ocean-colour time series for use in climate studies: the experience of the ocean-colour
652 climate change initiative’, **Unpublished manuscript**.

653 Sathyendranath, S., Stuart, V., Nair, A., Oka, K., Nakane, T., Bouman, H., Forget, M.-H.,
654 Maass, H. and Platt, T. (2009), ‘Carbon-to-chlorophyll ratio and growth rate of phyto-
655 plankton in the sea’, *Marine Ecology Progress Series* **383**(7), 73–84.

656 Schmidtko, S., Johnson, G. C. and Lyman, J. M. (2013), ‘MIMOC: A global monthly isopyc-
657 nal upper-ocean climatology with mixed layers’, *Journal of Geophysical Research: Oceans*
658 **118**(4), 1658–1672.

- 659 Sieburth, J. M., Smetacek, V. and Lenz, J. (1978), ‘Pelagic ecosystem structure: Het-
660 erotrophic compartments of the plankton and their relationship to plankton size fractions
661 1’, *Limnology and Oceanography* **23**(6), 1256–1263.
- 662 Stramski, D., Reynolds, R. A., Babin, M., Kaczmarek, S., Lewis, M. R., Röttgers, R., Scian-
663 dra, A., Stramska, M., Twardowski, M., Franz, B. et al. (2008), ‘Relationships between the
664 surface concentration of particulate organic carbon and optical properties in the eastern
665 south pacific and eastern atlantic oceans’, *Biogeosciences* **5**(1), 171–201.
- 666 Strathmann, R. (1967), ‘Estimating the organic carbon content of phytoplankton from cell
667 volume or plasma volume’, *Limnology and Oceanography* **12**(3), 411–418.
- 668 Taylor, K. E., Stouffer, R. J. and Meehl, G. A. (2012), ‘An overview of cmip5 and the
669 experiment design’, *Bulletin of the American Meteorological Society* **93**(4), 485–498.
- 670 Vidussi, F., Claustre, H., Manca, B. B., Luchetta, A. and Marty, J.-C. (2001), ‘Phyto-
671 plankton pigment distribution in relation to upper thermocline circulation in the eastern
672 mediterranean sea during winter’, *Journal of Geophysical Research: Oceans (1978–2012)*
673 **106**(C9), 19939–19956.