

Snail blocks the cell cycle and confers resistance to cell death

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The Snail zinc-finger transcription factors trigger epithelial-mesenchymal transitions (EMTs), endowing epithelial cells with migratory and invasive properties during both embryonic development and tumor progression. During EMT, Snail provokes the loss of epithelial markers, as well as changes in cell shape and the expression of mesenchymal markers. Here, we show that in addition to inducing dramatic phenotypic alterations, Snail attenuates the cell cycle and confers resistance to cell death induced by the withdrawal of survival factors and by pro-apoptotic signals. Hence, Snail favors changes in cell shape versus cell division, indicating that with respect to oncogenesis, although a deregulation/increase in proliferation is crucial for tumor formation and growth, this may not be so for tumor malignization. Finally, the resistance to cell death conferred by Snail provides a selective advantage to embryonic cells to migrate and colonize distant territories, and to malignant cells to separate from the primary tumor, invade, and form metastasis.

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The Snail family members Snail and Slug trigger epithelial-mesenchymal transitions (EMTs) during embryonic development and tumor progression (Nieto 2002). During development, they are involved in the ingression of the early mesodermal cells at gastrulation and in the delamination of the neural crest from the neural tube (Nieto 2002). Functional analyses in the chick (Nieto et al. 1994) and a striking interchange in the expression patterns at the sites of EMT in chicken and mouse embryos (Sefton et al. 1998) suggested that Slug induces the transition in the chick, whereas Snail triggers EMT in the mouse. Indeed, in mammalian cells *Snail* induces EMT and represses *E-cadherin* transcription (Batlle et al. 2000; Cano et al. 2000), and *Snail* mutant mice die at gastrulation due to a defective EMT and maintained *E-cadherin* expression (Carver et al. 2001). The analysis of the gene family in all major vertebrate groups indicated that *Snail* is higher in the gene hierarchy controlling neural crest development in fish, amphibians, and mammals (Locascio et al. 2002; Aybar et al. 2003).

Snail is also involved in the EMT that takes place concomitant with the acquisition of invasive properties in tumors (Nieto 2002; Thiery 2002). It is expressed in the invasive cells of tumors induced in the skin of mice (Cano et al. 2000) and in biopsies from patients with ductal breast carcinomas (Cheng et al. 2001; Blanco et al.

2002), gastric cancer (Rosivatz et al. 2002), and hepatocellular carcinomas (Sugimachi et al. 2003). *Snail* appears as an early marker of the malignant phenotype and behaves as a prognostic factor (Blanco et al. 2002).

The process of EMT implies a dramatic phenotypic change that includes the loss of epithelial markers, the gain of mesenchymal markers, and changes in cell shape. Because Snail is able to induce a complete EMT (Batlle et al. 2000; Cano et al. 2000), it must have many targets. Indeed, together with *E-cadherin*, other direct targets for Snail repression that have been identified include the epithelial *Mucin-1* (Guaita et al. 2002) and the components of the tight junctions *claudins* and *occludin* (Ikenouchi et al. 2003). Snail is upstream of molecules involved in the degradation of the basement membrane and extracellular matrix such as metalloproteinase 2 (MMP-2; Yokohama et al. 2003), the mesenchymal markers vimentin and fibronectin (Cano et al. 2000; Guaita et al. 2002), and other transcription factors such as ZEB-1 and LEF-1 (Guaita et al. 2002). Although a direct link between Snail expression and cytoskeletal proteins has not been reported, RhoB, a small GTPase involved in cytoskeletal actin rearrangements, lies downstream of Slug during chick neural crest delamination (Del Barrio and Nieto 2002).

By analyzing epithelial cells transfected with *Snail* and mouse and chick embryos, we show here that Snail also regulates cell-cycle progression and survival. Snail regulates components of the early to late G1 transition and the G1/S checkpoint, including the repression of *Cyclin D2* transcription and the increase in p21/Cip1. Concomi-

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tantly, Snail confers resistance to the lethal effects of serum depletion or TNF- α administration by activating the MAPK and PI3K survival pathways. These survival properties confer a selective advantage to the invasive and migratory cells during both embryonic development and tumor dissemination.

Results

Snail impairs cell-cycle progression

When stably expressed in different epithelial cell lines, both mouse and human Snail dramatically decreased cell growth, prompting us to analyze cell proliferation and cycle progression of such cells. When transfected with *Snail* (MDCK-Snail), MDCK cells underwent a complete EMT (Cano et al. 2000) and incorporated lower levels of BrdU after 24 h in culture (25% of that observed in mock-transfected cells, Fig. 1A). Analysis by FACS (Fig. 1B) indicated that the vast majority of Snail-expressing cells (93%) was in the G0/G1 phase of the cell cycle in basal conditions after 72 h in culture. However, these cells can respond to mitogens and proliferate, as observed 16 h after they were replated with 10% serum (Fig. 1B). The percentage of cells in G0/G1 was again much higher in MDCK-Snail transfectants (61% vs. 36%) after 24 h in culture and at all other times analyzed (Fig. 1B). To check whether Snail-expressing cells had difficulties in progressing through the G1/S checkpoint, we analyzed the expression of the Cip/Kip proteins, essential to inhibit the activity of the cdk2–Cyclin E complex, and responsible for the hyperphosphorylation of the retinoblastoma protein (Rb) and the subsequent progression from late G1 to the S phase (Ortega et al. 2002). Although the levels of p27 (Kip1) remained low in both MDCK-Mock and MDCK-Snail cells, Snail greatly increased the expression of p21 (Cip1) as shown in basal conditions and after 8 h (Fig. 2A) and 12 h in culture (Fig. 2B). Nevertheless, a decrease was observed after 16 h, coincident with Rb hyperphosphorylation (Fig. 2A) and entry into S phase (Fig. 1B). When we analyzed the levels of p21 at later times, we found that they increased again

in Snail-expressing cells after 24 h and were always maintained at higher levels than in control transfectants (Fig. 2B). Thus, p21 expression is tightly regulated and maintained at a high level except for during short periods of time when cells are exposed to a high concentration of mitogens and respond by progressing through the cycle.

Snail represses Cyclin D2 transcription

The G1/S checkpoint requires the activity of Cyclin E and the activity of the Cyclins D helps to progress from early to late G1 passing the restriction point (R) where cells are committed to another round of division. Thus, we decided to analyze the expression of Cyclins D and E in the two cell lines. Although we could not detect significant differences in the amount of Cyclin E (data not shown) or Cyclin D3 (Fig. 3A) in MDCK Snail-expressing versus nonexpressing cells, we observed a decrease in the levels of Cyclins D1 and D2 (Fig. 3A). In addition, we also found that the expression of their partner, the cyclin-dependent kinase cdk4, was decreased in MDCK-Snail cells (Fig. 3A). These results indicate that Snail-expressing cells, with limited amounts of Cyclins D, have an impairment to progress through the restriction point.

After we had observed that Snail-expressing cells contain low levels of Cyclins D, we wanted to assess whether they can be direct targets of Snail transcriptional repression. Although we found that the levels of *Cyclin D1* mRNA were lower in cells expressing Snail compared to mock-transfected cells, those of *Cyclin D2* were strongly down-regulated (Fig. 3B). Because the *Cyclin D2* promoter contains two E-box consensus for Snail binding (Mauhin et al. 1993) that are conserved among mouse, rat, and human (Bouchard et al. 1999), we analyzed the effect of Snail on *Cyclin D2* transcription. We expressed mouse *Snail* cDNA together with reporter constructs of the human *Cyclin D2* promoter in the keratinocyte cell line MCA3D, previously used to study the proximal *E-cadherin* promoter (Cano et al. 2000). Snail repressed the wild-type promoter activity of *Cyclin D2* to ~55% of its activity (Fig. 3C) but did not affect that

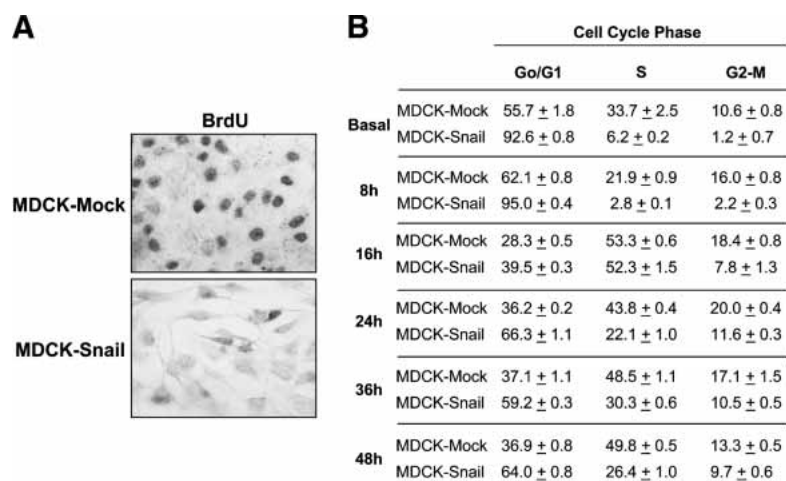


Figure 1. Snail expression impairs cell proliferation. (A) BrdU incorporation after a 1-h pulse in MDCK cells stably transfected with Snail (MDCK-Snail) or the empty vector (MDCK-Mock). Bright-field images after 24 h in culture. (B) FACS analysis of the cell cycle in MDCK-Mock and MDCK-Snail cells after different times in culture.

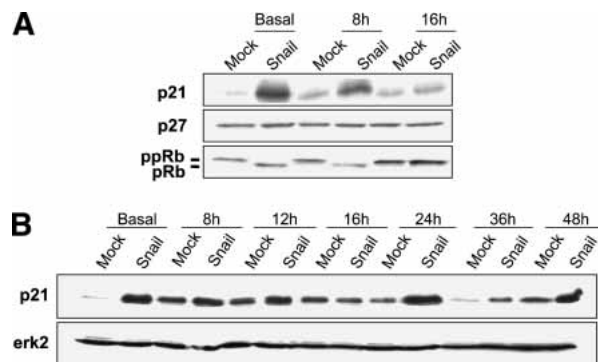


Figure 2. Snail alters the expression of proteins involved in progression from G1 to S phase. Western blot analysis of MDCK-Mock and MDCK-Snail cells. (A) Levels of the G1 checkpoint molecules, the cdk inhibitors p21 and p27, and the degree of Rb phosphorylation. (B) Levels of p21 after different times in culture. Rb, retinoblastoma protein; pRb, hypophosphorylated state; ppRb, hyperphosphorylated state. Representative experiments are shown ($n = 4$).

of promoter constructs carrying either a mutated or deleted distal E-box (located at -1600). Reporter activity was significantly decreased (to $\sim 40\%$) when the proximal E-box (located at -1400) was mutated or deleted. As this construct is insensitive to Snail when the distal box is intact, both E-boxes are needed to mediate repression. The proximal box and sequences located immediately 3' to it must be essential for promoter activity, because the -1303 D2-Luc deletion provoked a 10-fold decrease in activity (Fig. 3C). Similar results were obtained when human Snail cDNA constructs were used (data not shown). Thus, Snail can repress the activity of the *Cyclin D2* promoter in epithelial cells. The presence of the proximal E-box (-1400) is needed for the distal box (-1600) to repress, and the region between -1400 and -1300 that includes the proximal box acts as an enhancer.

Snail and the cell cycle in the mouse embryo

Having seen that Snail repressed *Cyclin D2* expression in cultured cells, we set out to determine whether the same could occur during embryonic development. Thus, we compared the expression of *Cyclins D1* and *D2* with that of *Snail* in 8.5-d postcoitum (dpc) mouse embryos (Fig. 4). At this developmental stage, *Snail* is expressed in regions undergoing EMT such as the premigratory neural crest and the primitive streak (Nieto et al. 1992; Smith et al. 1992; Sefton et al. 1998), and in mesodermal derivatives including the decondensing somites and the allantois (Fig. 4A,D,G,I). *Cyclin D2* transcripts were readily detected at high levels in the neural plate (Fig. 4F) and the neural tube (Fig. 4I), whereas they were absent from *Snail*-expressing regions (Fig. 4F,I,L), evidencing a clear inverse correlation in their expression patterns. *Cyclin D1* was also expressed in the neural tube (Fig. 4E,H) and absent from regions expressing high levels of *Snail* tran-

scripts such as the allantois (Fig. 4K). However, *Cyclin D1* transcripts were observed in regions where *Snail* was only moderately expressed, including the decondensing somites (Fig. 4H). Thus, our data are compatible with Snail being a strong repressor of *Cyclin D2* transcription, and are in agreement with the results obtained from Northern analysis of both *Cyclin D1* and *D2* and with those of the *Cyclin D2* promoter activity in epithelial cells.

The impairment in cell-cycle progression in Snail-expressing cells was due not only to the maintained down-regulation of *Cyclin D2* transcription but also to the changes in the expression of additional components of the G1/S checkpoint. Indeed, the expression of Snail was correlated with an increase of the proportion of cells in G0/G1 (Fig. 1). To assess whether a similar change occurred in vivo, we compared the pattern of *Snail* expression with that of BrdU incorporation in mouse embryos to visualize cells in the S phase of the cycle (Fig. 5A,B). In agreement with the data from cell cultures, in the regions of the embryo where *Snail* was expressed, a much lower proportion of nuclei accumulated BrdU. In general, it was possible to discern a complementary pattern of *Snail* expression and BrdU incorporation within the em-

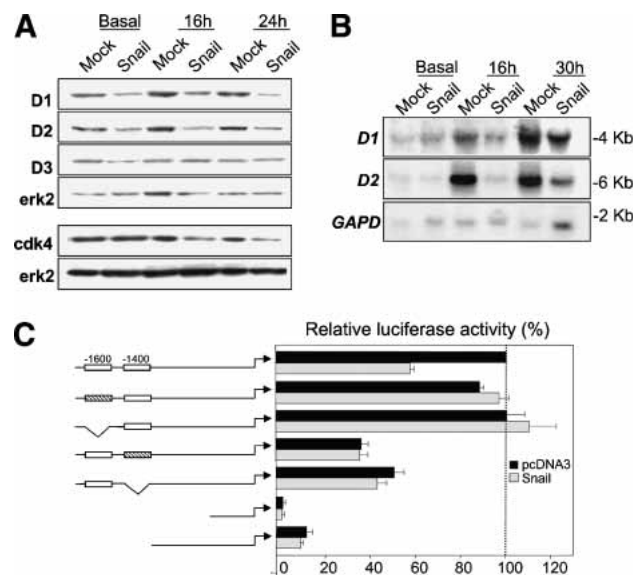


Figure 3. Snail represses *Cyclin D2* transcription. (A) Analysis of D cyclins and their partner cdk4. Immunoblotting of total erk2 was used as a control of gel loading. (B) Analysis of *Cyclins D1* and *D2* transcription by Northern blot of RNA extracted from MDCK-Mock and MDCK-Snail cells after different times in culture. The *GAPDH* probe was used as a control of loading. (C) Activity of the *Cyclin D2* promoter. Luciferase reporter constructs carrying the wild-type human *Cyclin D2* promoter (-1624) or independent deletions/mutations in the two E-boxes were transfected into MCA3D cells together with a mouse Snail expression vector or the empty vector (pcDNA3) as a control. Luciferase activity was assayed 40 h after transfection. Activity is expressed relative to that of the wild-type construct. Results are the mean values \pm S.E. of duplicates from four independent experiments.

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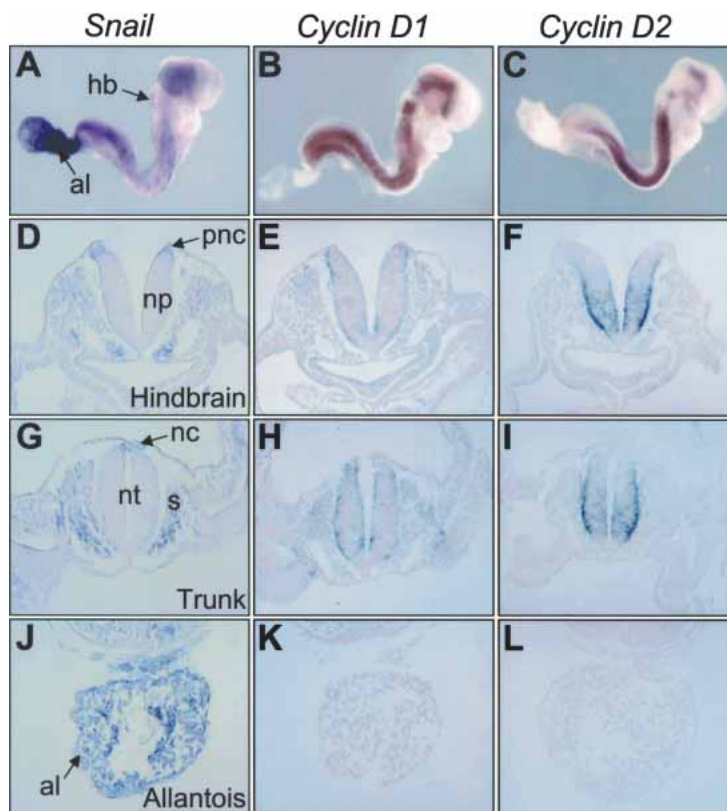


Figure 4. An inverse correlation exists between *Cyclin D2* and *Snail* expression in mouse embryos. Whole-mount in situ hybridization of 8.5-dpc mouse embryos (A–C) and transverse paraffin sections of the same embryos taken at the level of the posterior hindbrain (D–F), the trunk (G–I), and the allantois (J–L). *Snail* expression can be observed at the edges of the neural plate (D, pnc) corresponding to premigratory crest cells undergoing EMT. *Snail* expression is maintained in crest cells after delamination (G, nc), and it is also apparent in the decondensing somites (G, s) and in the allantois (J, al). An inverse correlation between *Snail* and *Cyclin D2* transcripts is readily observed in all the tissues analyzed (cf. D, G, J and F, I, L). (K) Although this correlation is not so striking for *Snail* and *Cyclin D1*, note that *Cyclin D1* expression is not detected in regions with high levels of *Snail* transcripts such as the allantois. (al) Allantois; (hb) hindbrain; (nc) neural crest; (np) neural plate; (nt) neural tube; (pnc) premigratory neural crest; (s) somite.

bryo (Fig. 5A,B). On the whole, as BrdU incorporation increased, *Snail* expression decreased (Fig. 5, cf. A and B [along the proximodistal axis of the allantois], C and D [in the section taken through the forebrain]). We also analyzed the phosphorylation of histone H3 to quantify the cells undergoing mitosis (Prigent and Dimitrov 2003) in the developing nervous system of 8.5-dpc mouse embryos. We observed high levels of phospho-H3 in the ventricular surface of the neural epithelium, where nuclei undergo mitosis (Fig. 5G,H). When quantified, at both the head and trunk levels, an approximately fivefold decrease in labeling was observed in the *Snail*-expressing regions compared to similar-sized regions located in the intermediate or ventral areas. These results indicate that *Snail*-expressing cells are difficult to find undergoing DNA synthesis or mitosis both in culture and in embryos.

Snail confers resistance to serum depletion-induced cell death

In several studies of hepatocytes (Valdés et al. 2002) and cancer cells progressing towards malignancy (Thiery 2002; Siegel and Massagué 2003), the transition to the mesenchymal phenotype is associated with a reduced susceptibility to apoptosis. Because *Snail* induces a complete EMT in MDCK cells (Batlle et al. 2000; Cano et al. 2000), accompanies tumor malignancy (Blanco et al. 2002; Sugimachi et al. 2003), and is induced in hepatocytes undergoing EMT (Spagnoli et al. 2000; Valdés et al.

2002), we tested whether *Snail* conferred resistance to cell death. Two main pathways can provoke apoptosis, the stress pathway triggered by developmental cues or intracellular damage (γ -irradiation, cytokine deprivation, etc.), and the activation of death receptors such as those of the TNF family mediated by extracellular signals, among them TNF- α , Fas ligand, and Trail.

To analyze the response to cellular stress, cells were maintained in the absence of serum. MDCK-*Snail* cells survived for at least 7 d, whereas a large amount of cell death was seen in mock-transfected cells after 2–3 d of serum deprivation (Fig. 6A). After 48 h in culture, labeling with propidium iodide showed that whereas mock transfectants had $10.3 \pm 0.1\%$ positive nuclei, only $3.8 \pm 0.2\%$ of the nuclei of *Snail*-expressing cells were labeled. Caspase-3 is involved in the cell death response and was three times more active in mock-transfected cells than in the corresponding *Snail*-expressing cells (Fig. 6B).

To check whether *Snail* may confer resistance to the death induced by developmental cues, we compared the pattern of cell death with that of *Snail* expression in mouse embryos (Fig. 6C). Although a clear pattern of cell death has been identified in rhombomeres (r) 2, 3, and 5 in the developing hindbrain of the chick (Graham et al. 1996; Ellies et al. 2000), the situation is not so clear in the mouse (Trainor et al. 2002). By comparing with the known expression of *Krox-20* in the presumptive rhombomeres (pr) 3 and 5 at this developmental stage (Fig. 6C), we observed numerous dying cells in pr2, 3, 5, and 6

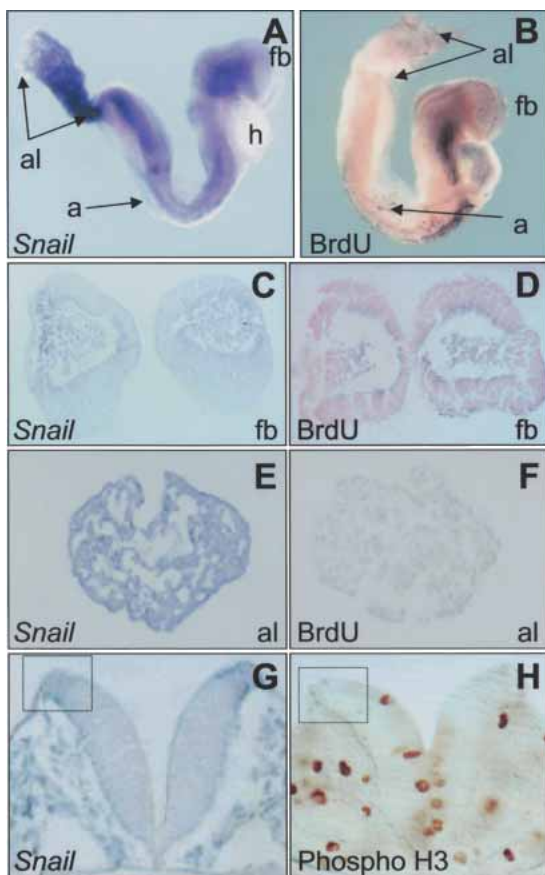


Figure 5. An inverse correlation exists between proliferation and *Snail* expression in mouse embryos. The embryos in *A* and *B* show a side-by-side comparison between *Snail* expression and BrdU incorporation as a measure of cell proliferation in the whole embryo in culture. An overall complementary pattern is observed, that can be better examined in the sections taken at the level of the forebrain (*C,D*) and the base of the allantois (*E,F*). *G* and *H* show sections at the trunk level to compare *Snail* expression with histone H3 phosphorylation, taken as a measure of cells undergoing mitosis. The squares mark the *Snail*-expressing region of the neural epithelium. (a) Amnion; (al) allantois; (fb) forebrain; (h) heart; (hb) hindbrain.

(NBS; Fig. 6C). Conversely, pr4 and the anterior spinal cord are regions where cell death was not apparent, coinciding with areas of *Snail* expression (Fig. 6C, high magnification). The absence of *Snail* expression from other regions of intense cell death was also apparent in the midbrain and forebrain, and in other tissues outside of the central nervous system such as the developing heart (Fig. 6C). In contrast, mesenchymal tissues of the head, including the migratory neural crest cells, showed high levels of *Snail* transcripts and no cell death. In summary, an inverse correlation between cell death and *Snail* expression existed in the embryos analyzed (Fig. 6C).

Given that *Snail*-expressing cells were resistant to serum depletion, we studied the activity of different survival pathways in these cells. Both the MAPK and PI3K pathways are highly active in *Snail*-expressing cells (Fig.

7A,B). In agreement with the higher levels of PI3K activity, we also found a much greater phosphorylation of its downstream effector Akt (Fig. 7B). Both the MEK/Erk and PI3-K/Akt pathways can mediate the up-regulation of Bcl-x_L expression (Ramljak et al. 2003), a death-inhibitory member of the Bcl-2 family that blocks the stress-induced release of cytochrome c from the mitochondria. Thus, we looked at the expression of Bcl-x_L in MDCK-*Snail* cells and found that it was increased at all times analyzed (Fig. 7C). These data indicate that the activation of the MEK/Erk and PI3-K/Akt pathways may explain the survival properties associated with *Snail* expression following serum depletion.

Snail protects from TNF- α -induced cell death

Because *Snail* expression conferred resistance to stress-induced cell death, we checked whether *Snail*-expressing cells might also be resistant to that induced by proapoptotic signals that activate the death receptor pathway such as TNF- α . Indeed, whereas mock-transfected MDCK cells died within 24 h of treatment, MDCK-*Snail* cells survived (Fig. 8A). Caspase-8 is specifically recruited to the death receptors upon ligand binding and is then auto-activated initiating the apoptotic pathway. Accordingly, we found that treatment with TNF- α induced the activation of caspase-8 in mock-transfected cells. An approximately fivefold decrease in the activity of caspase-8 was observed in *Snail*-expressing cells after 24 h of treatment compared with that of mock transfectants. This result substantiates that the death observed in MDCK cells was mediated by the activation of this pathway (Fig. 8B). As expected, in both cell types the activity of the initiator, caspase-8, correlated with that of the effector, caspase-3 (Fig. 8C), confirming that the expression of *Snail* protects the MDCK cells from TNF- α -induced death.

Slug behaves in the chick as *Snail* in the mouse

We showed previously that the *Snail* family member *Slug* induces EMT in the chick embryo (Nieto et al. 1994) and that in general, the expression patterns of these two family members are interchanged at the sites of EMT in birds and mammals (Sefton et al. 1998). Indeed, the cells undergoing EMT in the neural tube and the primitive streak express *Snail* in the mouse and *Slug* in the chick. Nevertheless, *Snail* can induce EMT when ectopically expressed in the chick embryo hindbrain (Del Barrio and Nieto 2002), indicating that the two proteins can be functionally equivalent during embryonic development. However, even though they can be functionally equivalent, only one gene is expressed in each corresponding embryo and thus, *Snail* could not play a role in the control of cell division or survival in the neural tube of the chick, because it is not expressed there. Thus, this functional equivalence and the striking interchange in the expression patterns led us to analyze whether the family member expressed in the chick, *Slug*, can regulate cell-cycle progression and survival in the neural tube.

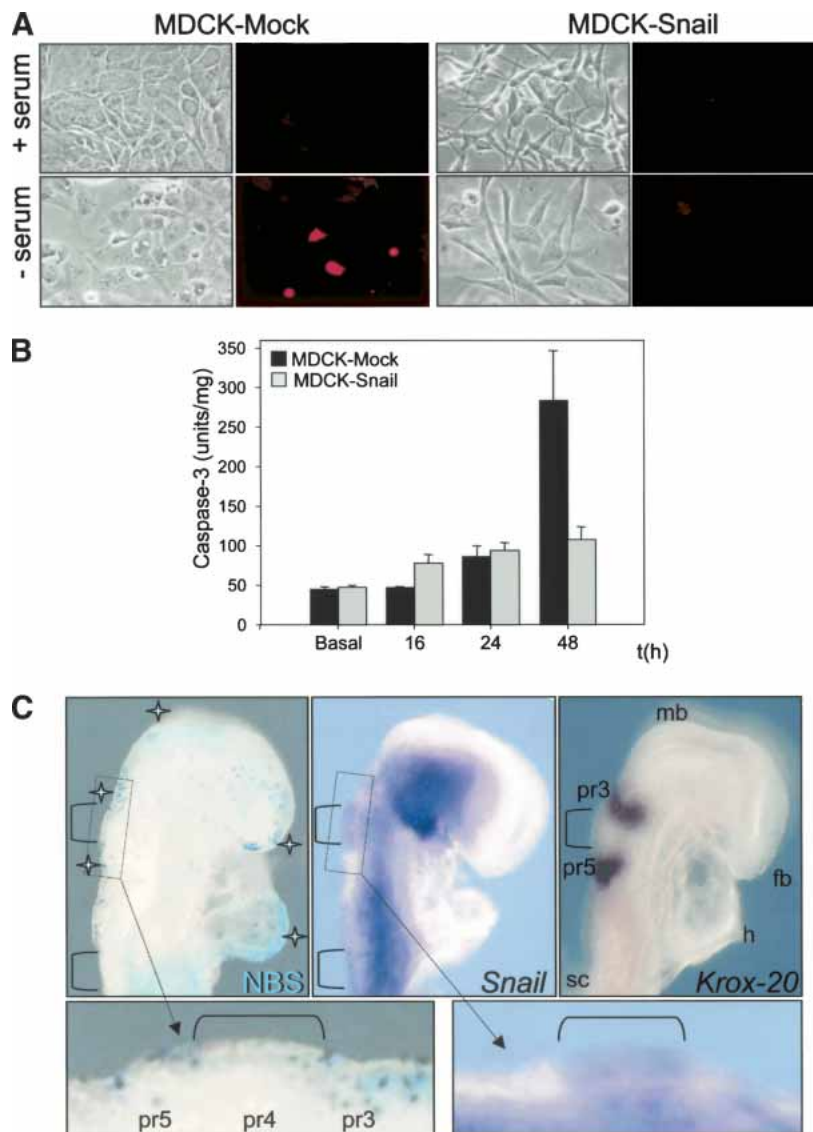


Figure 6. Snail confers resistance to apoptosis induced by serum deprivation. (A) Cell viability was assessed by propidium iodide staining 48 h after serum depletion. (B) Caspase-3 activity at different times after serum removal represented as mean values \pm S.E. from three independent experiments carried out with duplicate dishes. Note the low levels of activity in *Snail*-expressing cells 48h after serum deprivation compared to the mock-transfected cells. (C) Cell death visualized using Nile Blue Sulphate staining is compared side-by-side with *Snail* expression in the head of an 8.5-dpc mouse embryo. The pattern of cell death assessed by Nile Blue Sulphate (NBS) staining (stars) is complementary to that of *Snail* (brackets). A similar embryo hybridized with *Krox-20* to indicate the relative position of pre-rhombomeres (pr) 3 and 5 in the hindbrain to help compare the pattern of cell death and *Snail* expression. The inverse correlation can be better assessed in the high-power photographs. (fb) Forebrain; (h) heart; (mb) midbrain; (sc) anterior spinal cord.

To analyze the cell cycle we measured both BrdU incorporation and phospho-H3 expression in chick embryos as we had performed for the mouse. We observed high levels of cell proliferation in the basal half of the neural tube epithelium, where the nuclei undergo DNA synthesis. In contrast, and as previously described (Burstyn-Cohen and Kalcheim 2002), the levels of incorporation were much lower in the dorsal region of the neural tube at the level of the epithelial somites (Fig. 9A). Interestingly, the dorsal neural tube is occupied by the premigratory neural crest, which expresses high levels of *Slug* transcripts (Fig. 9B). Quantification of BrdU-positive cells indicated that the proportion of cells in the S phase of the cycle was approximately three times less in the *Slug*-expressing area compared to that found in nonexpressing regions of a similar size (data not shown). When we quantified the cells containing phospho-H3 all along the dorsoventral axis of the neural tube, we found that in the *Slug*-expressing area, positive cells were only

~15% of those found in adjacent regions (data not shown). Thus, with respect to the cell cycle, *Slug*-expressing cells in the developing neural tube of the chick embryo behave as *Snail*-expressing cells in the mouse embryo and similarly, they are difficult to find undergoing DNA synthesis or mitosis.

As already mentioned, cell death has been analyzed in detail in the chick embryonic hindbrain (Ellies et al. 2000), with cell death in neural crest cells from r3 and r5 being crucial for the patterning of the branchial region (Graham et al. 1996; Ellies et al. 2002; Trainor et al. 2002). We carried out a careful analysis of cell death by both TUNEL and NBS staining and compared the data with the expression of *Slug*. We found that r4, which is naturally protected from cell death, expressed much higher levels of *Slug* transcripts than the adjacent rhombomeres (Fig. 9C–E). This result is reminiscent of the data regarding cell death and *Snail* expression from mouse embryos (see Fig. 6C).

Taking advantage of the amenability of the chick embryo to experimental manipulation, we further studied the role of *Slug* in promoting cell survival by overexpressing it in the neural tube in ovo. High levels of GFP (and *Slug*, data not shown) expression could be achieved by co-electroporation of *Slug*- and GFP-encoding vectors into the right-hand side of the developing neural tube (Fig. 9F). Several streams of electroporated neural crest cells were observed migrating from the hindbrain (Fig. 9F). NBS staining of the same embryo shows that the area of *Slug* overexpression presents a dramatic reduction of the naturally occurring cell death (Fig. 9G,H). Rescue of cell death could be observed in 53% of the electroporated embryos ($n = 30$), indicating that *Slug* can act as a survival factor in the chick embryo hindbrain. It is worth noting here that the amount of naturally occurring cell death in the different rhombomeres correlates with the balance in the expression of the death inducer BMP (Graham et al. 1994) and the survival factor *Slug*. Indeed, r4 expresses high levels of *Slug* and very low levels of *BMP4*. This balance also explains why in our overexpression experiments the death of r2 cells (with low levels of both *BMP4* and *Slug*) can be more effectively rescued by *Slug* overexpression than that of r5 (which expresses very high levels of *BMP4*; data not shown).

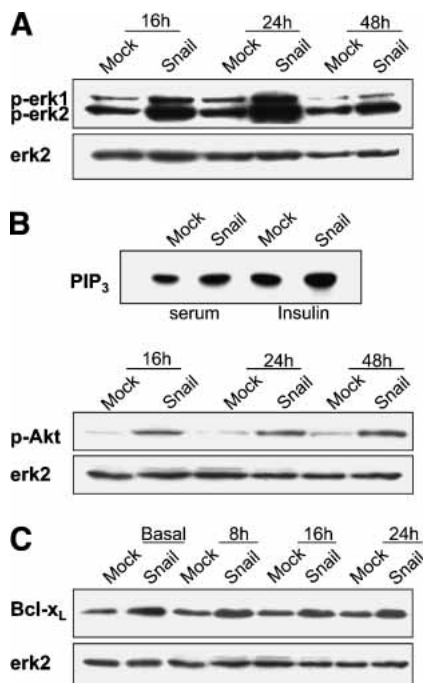


Figure 7. Snail activates survival pathways. Molecules from different survival pathways were analyzed in cells cultured in the absence of serum and collected at different times. The levels of active ERKs (phospho-erk1 and phospho-erk2; A), active Akt (B), and Bcl-x_L (C) were analyzed by Western blot and found to be increased in Snail-expressing cells. Total erk2 was used as a control for gel loading. (B) Higher levels of PI3K activity were also detected in Snail-expressing cells as analyzed by thin-layer chromatography, compatible with the higher levels of phosphorylation found for Akt.

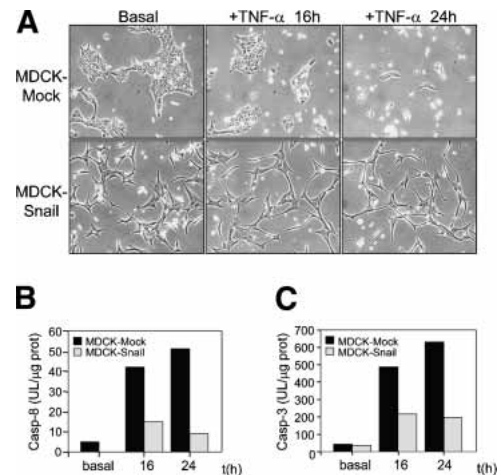


Figure 8. Snail confers resistance to the cell death induced by TNF- α . Mock and MDCK-Snail-expressing cells were treated with TNF- α (5 ng/mL) after being pretreated with cycloheximide (0.5 μ g/mL for 30 min) to prevent the induction of the survival protein NF κ B. (A) Photographs of the cultures taken after 16 or 24 h of treatment. (B,C) The activity of the death receptors-specific caspase-8 and effector caspase-3, respectively, are shown from one representative experiment. Note the low activity of both caspases in Snail-expressing cells, explaining the healthy appearance observed in A.

Although some of the migratory *Slug*-expressing cells pass close to the otic vesicles, the dying cells indicated by black stars in Figure 9H are nonelectroporated ectodermal cells (as assessed in sections, data not shown). These results highlight the specificity of this rescue in the electroporated cells, and confirm that *Slug* confers resistance to cell death in the developing embryo.

Discussion

Snail genes impair cell-cycle progression in cultured cells and in developing embryos

The epithelial mesenchymal transition (EMT) confers migratory and invasive properties to epithelial cells, critical for the generation of cells that originate at a distance from their final destination (Nieto 2002) during embryonic development. When the transition to a mesenchymal phenotype occurs in the adult, it is usually associated with pathological processes such as tumor progression (Thiery 2002). Concomitant with the acquisition of motility, cells undergo dramatic changes in cell adhesion properties and cell shape. Snail induces a complete EMT in epithelial cells concurrent with the aforementioned changes (Batlle et al. 2000; Cano et al. 2000) and is associated with the process of EMT in both physiological and pathological conditions (Nieto 2002; Thiery 2002; Yáñez-Mo et al. 2003). The conversion to mesenchyme implies a profound reorganization of the cytoskeleton that may be incompatible with a highly proliferative state. Here we demonstrate that cell proliferation is impaired in Snail-expressing cells, supporting this

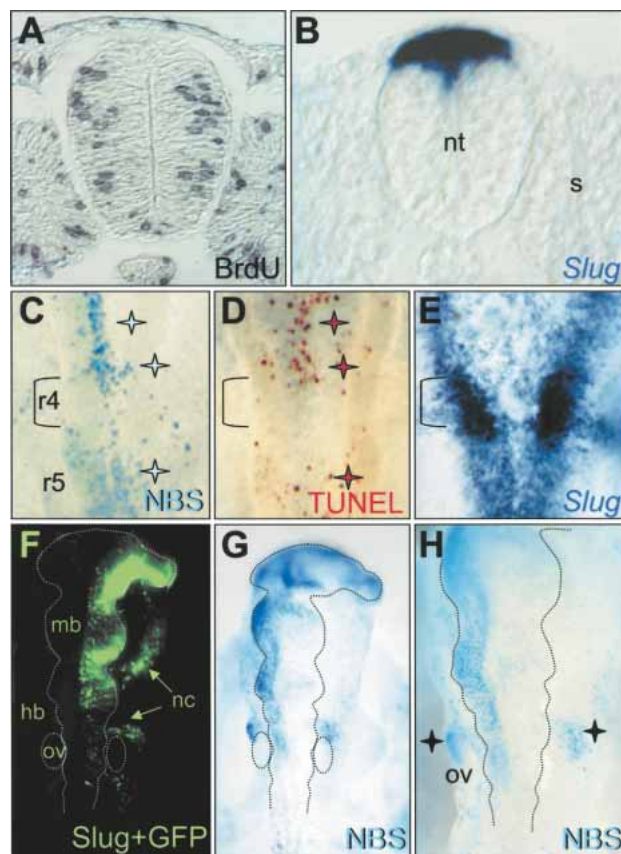


Figure 9. Slug expression correlates with little proliferation and protects the developing neural tube from physiological cell death in the chick. (A) In ovo BrdU incorporation was analyzed in transverse sections. One such section from the trunk of a stage 11 chicken embryo is shown. (B) A similar section hybridized with a *Slug* probe. Note the absence of BrdU in the premigratory neural crest, showing high levels of *Slug* expression. (C,D) The pattern of cell death in the hindbrain region of a stage 12 chick embryo as assessed by NBS and TUNEL staining, respectively. Compare the pattern of cell death (blue and brown stars in C,D) with that of *Slug* transcripts (E). As previously described, r4 shows very few apoptotic cells, coinciding with high levels of *Slug* transcripts (brackets). (F–H) An embryo electroporated with plasmids containing chick *Slug* and *GFP* cDNAs at stage 8 and analyzed 15 h later (stage 12). (F) GFP (and thus, *Slug*) expression is observed in the right-hand side of the neural tube and in cells migrating from it. (G) NBS staining of the same embryo shows a striking decrease in cell death in the side where *Slug* is overexpressed. (H) A higher-magnification picture that allows a better assessment of the region protected from cell death. The dotted lines demarcate the borders of the neural tube and the otic vesicle. The black stars indicate a region of ectodermal cell death that appears symmetrical on both sides of the embryo (see text). (hb) Hindbrain; (mb) midbrain; (nc) neural crest; (nt) neural tube; (ov) otic vesicle, (s) somite.

hypothesis. Further evidence that profound morphological changes are incompatible with high proliferation can be seen in the low BrdU incorporation in the premigratory neural crest (Burstyn-Cohen and Kalcheim 2002; the present study), in the mitotic arrest in the cells of the ventral furrow during *Drosophila* gastrulation (Foe

1989), the low proliferation in the invasive front of carcinomas (Jung et al. 2001), and the link between high p21 expression and poor prognosis in breast carcinomas (Yang et al. 2003). Significantly, Snail is expressed and triggers EMT in all of these territories and circumstances (for review, see Nieto 2002).

We have found that Snail impairs the transition from early to late G1 by maintaining low levels of Cyclins D and can block the G1/S transition by maintaining high levels of p21. However, Snail-expressing cells can respond to mitogenic signals by transiently decreasing p21 expression, which favors the transition to the S phase. Interestingly, neural crest cells synchronously enter into the S phase upon delamination from the neural tube (Burstyn-Cohen and Kalcheim 2002), which occurs after the process of EMT has been completed. Thus, when Snail (and *Slug* in the chick) induces EMT in the premigratory neural crest cells, it probably blocks proliferation, synchronizing them in G1 and allowing morphological changes to occur. Subsequently, cells will enter S phase upon delamination. Interestingly, the bladder epithelial carcinoma cell line NBT-2 is receptive only to FGF-induced EMT at G1 (Bonneton et al. 1999), and lymphocyte migration and hepatoma cell invasion occur only in G1 (Ratner 1992; Iwasaki et al. 1995). Thus, although a deregulation/increase in cell division is crucial for tumor formation and growth, this is not so for tumor malignization. The behavior of invasive cells allows tumor proliferation to be dissociated from malignancy.

Snail genes protect cultured cells and embryos from cell death induced by both the retrieval of survival factors and apoptotic signals

Selective cell death is of crucial importance for sculpting the embryo and maintaining tissue homeostasis. However, deregulation of programmed cell death can be critical in pathological processes such as cancer. Several studies have correlated the conversion to a mesenchymal phenotype with cell survival. Indeed, EMT protects fetal hepatocytes from the death induced by TGF- β (Valdés et al. 2002). Interestingly, TGF- β has multiple effects on cellular behavior, from inducing growth arrest and death to triggering EMT, survival, and tumor progression (Siegel and Massagué 2003). Indeed, when expressed in the skin of mice, TGF- β inhibits the formation of benign tumors but increases the frequency of invasive spindle carcinomas (Cui et al. 1996). Members of the TGF- β superfamily including TGF- β s and BMPs induce Snail/*Slug* expression in several systems such as hepatocytes (Spagnoli et al. 2000; Gotzmann et al. 2002; Valdés et al. 2002), epithelial and mesothelial cells (Peinado et al. 2003; Yáñez-Mo et al. 2003), and in the developing embryo (Dickinson et al. 1995; Liem et al. 1995; Romano and Runyan 2000; Piedra and Ros 2002). When *Snail* genes are induced by TGF- β , EMT is triggered and cells become resistant to TGF- β -induced cell death (Valdés et al. 2002), consistent with Snail activating the Mek/Erk and PI3K/Akt survival pathways. TGF- β is also capable of inducing the progression towards invasive car-

cinoma (Cui et al. 1996; Siegel and Massagué 2003), and this is usually explained by the tumor cells overcoming the TGF β -induced growth arrest (Siegel and Massagué 2003). However, in light of our data, it seems more likely that these invasive tumor cells overcome cell death and not growth arrest. Indeed, TGF- β induces EMT in epithelial cells and hepatocytes concomitant with the inhibition of apoptosis but without altering the growth retardation effect (Lehmann et al. 2000; Valdés et al. 2002; Peinado et al. 2003).

With respect to embryonic development, we show here that the expression of *Snail* genes in the chick and the mouse are inversely correlated with cell death in different developing tissues. Significantly, *Slug* overexpression protects the neural crest from the naturally occurring cell death in the chick hindbrain. Interestingly, members of the BMP family induce *Snail* or *Slug* expression in the hindbrain (Nieto 2002), and BMPs are also responsible for the cell death induced in particular rhombomeres (Graham et al. 1996). This situation appears to be similar to the response of hepatocytes to TGF- β , where this factor induces cell death in half of the cell population and induces EMT and resistance to TGF- β -induced cell death in the other half (Valdés et al. 2002). TGF- β -induced EMT and survival in both hepatocytes and epithelial cells occurs concomitantly with *Snail* induction (Valdés et al. 2002; Peinado et al. 2003). Altogether, these data are in agreement with the notion that high levels of *Snail/Slug* expression in both mouse and chick hindbrains are sufficient to overcome the BMP-induced cell death.

Other members of the *Snail* superfamily are thought to mediate cell survival. In *Caenorhabditis elegans*, *Ces-2* represses the *Scratch* homolog *Ces-1* (Metzstein and Horwitz 1999), promoting the physiological death of a particular class of neurons. In humans, a translocation converts the *Ces-2* homolog (HLF) in an activator that in turn induces *Slug* expression, leading to aberrant cell survival and leukemogenesis (Inukai et al. 1999). Furthermore, hematopoietic progenitors in *Slug* null mutant mice show an increased sensitivity to the death induced by γ -irradiation (Inoue et al. 2002; Pérez-Losada et al. 2003). However, there is no indication that *Scratch* protects cells in *C. elegans* physiology (Metzstein and Horwitz 1999; Thellmann et al. 2003), or that *Slug* promotes survival in physiological circumstances in mammals. The evidence in these cases points to an anti-apoptotic function following DNA damage (Inoue et al. 2002; Pérez-Losada et al. 2003). It thus seems likely that *Snail* in mammals and *Slug* in avians may be more efficient in conferring resistance to cell death, with the other family member maintaining certain activity of this ancestrally inherited property. The ancestral condition associated with the *Snail* superfamily is also supported by the fact that *Scratch* is the member acting as such in *C. elegans*, as the nematode does not seem to express any functional *Snail* protein (Manzanares et al. 2001).

In conclusion, *Snail* favors changes in cell shape rather than proliferation in cells that become migratory, in accordance with the often neglected low rates of prolifera-

tion observed at the invasive front of tumors. Concomitantly, *Snail* offers protection from both stress-induced cell death and that provoked by pro-apoptotic signals. In this way, *Snail* confers a selective advantage to invasive cells to migrate through hostile territories. This resistance to cell death is essential in the embryo for migratory cells to reach their final destinations and in the adult for malignant cells to disseminate and form metastasis.

Materials and methods

Cell lines and antibodies

Canine MDCK (Madin-Darby canine kidney) and mouse epidermal keratinocyte MCA3D cells were grown in Dulbecco's modified Eagle medium (DMEM) and Ham's F12, respectively, supplemented with 10% fetal calf serum (FCS). The following antibodies were used: Polyclonal antisera against Cyclins D1, D2, and D3, cdk4, p21, p27, Erk2, and Bcl-x (Santa Cruz Biotechnology), anti-phospho Akt (New England Biolabs), and Phosphohistone-3 (Upstate Biotechnology); monoclonal p21/Cip1 and retinoblastoma protein (PharMingen BD Biosciences) and anti-MAPK (p44/p42; Cell Signaling Technology) antibodies. As secondary antibodies, both peroxidase-conjugated (anti-mouse and anti-rabbit) were used (Bio-Rad Laboratories) together with a biotinylated anti-rabbit serum (Vector).

Embryos

Mouse embryos were obtained from natural matings of Balb-C mice from the animal facility at the Cajal Institute. Fertilized hen eggs were obtained from Granja Santa Isabel, Córdoba, Spain. The age of mouse embryos was determined as days post-coitum (dpc), the day on which the vaginal plug was detected being designated 0.5 dpc. Eggs were incubated and opened, and the embryos staged according to Hamburger and Hamilton (1951).

Analysis of cell proliferation through BrdU incorporation and phosphohistone-3 immunohistochemistry

To measure the percentage that incorporated 5-bromo-2'-deoxyuridine (BrdU), cells were grown on coverslips in DMEM plus 10% FCS. BrdU (10 μ M in PBS) was added at different times, and cells were fixed and stained 1 h after addition according to the Detection kit II (Roche). Cells that had incorporated BrdU were visualized and quantified using a Leica DMR microscope. For in ovo labeling, BrdU (10 mM) containing fast green (0.25 μ L/mL) in PBS was injected into chick embryos 1 h before sacrificing. Mouse embryos were dissected in DMEM plus 10% FCS at 8.5 dpc, leaving the extraembryonic membranes intact. BrdU (10 mM) was immediately injected into the amniotic cavity, and the embryos were cultured for 1 h. They were fixed overnight in 4% paraformaldehyde at 4°C, dehydrated through a series of ethanol, and rehydrated. In the case of chick embryos, after fixation, dehydration, and rehydration, immunohistochemistry was carried out on 15- μ m paraffin sections (Fibrowax, BDH). After paraffin removal with HistoClear, the chick embryo sections were rehydrated, and both the chick sections and mouse embryos were processed to develop BrdU incorporation following the manufacturer's instructions (Roche).

Cells undergoing mitosis were identified by the presence of phospho-histone-3 (Prigent and Dimitrov 2003) in 50- μ m vibratome sections obtained from mouse and chicken embryos. The

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sections were incubated overnight with PBT (PBS, 0.5% Triton X-100) at 4°C and subsequently treated with 0.1% H₂O₂ for 4 h at room temperature. After washing, they were blocked with 10% FCS and 1 mg/mL BSA in PBT for 3 h at 4°C. Incubation with primary and secondary (biotinylated) antibodies was performed overnight at 4°C. After washing, the sections were developed with the ABC kit (Pierce).

Analysis of DNA content by flow cytometry

Cells were detached from dishes with trypsin, fixed in 70% ethanol (-20°C) for 1 min, and treated with RNase (1 mg/mL) for 15 min at 37°C. After propidium iodide staining (0.05 mg/mL in PBS 15 min at room temperature in the dark), the cellular DNA content was evaluated in a FACS flow cytometer (Becton-Dickinson). For computer analysis, only signals from single cells were considered (10,000 cells/assay).

Western blots and immunoprecipitation assays

Cells were scraped off the plates after washing with cold PBS and lysed at 4°C in the following buffer: 20 mM Tris-HCl at pH 7.4, 10 mM EDTA, 100 mM NaCl, 1% Triton X-100, 1 mM NaF, 100 mM β-glycerophosphate, 1 mM EGTA, 5 mM NaP_i, 5 μg/mL leupeptin, 1 mM sodium o-vanadate, and 1 mM PMSF. Proteins were separated by SDS-PAGE on 12% or 7.5% (for retinoblastoma protein), and transferred to PVDF membranes (Millipore) that were then blocked in TTBS (TBS plus 0.05% Tween-20) containing 5% nonfat dried milk. The membranes were incubated for 2 h with the corresponding primary antibody at room temperature in blocking solution. After washing, they were incubated with peroxidase-conjugated anti-mouse or anti-rabbit immunoglobulin for 1 h at room temperature (1:3000). Antibody binding was visualized by ECL (Amersham Biosciences).

For immunoprecipitation assays to detect levels of p21 and p27, the cells were lysed and the proteins purified as above. Equal amounts of total protein (400 μg) were incubated with 1 μg of the appropriate antibody (p21/Cip1 or p27/Kip1) and protein A-agarose beads (Sigma) for 3 h at 4°C, and the beads were washed twice with lysis buffer, centrifuged, resolved by SDS-PAGE, and subjected to immunoblot analysis as described above.

RT-PCR analysis and primer sequence

Poly(A)⁺ mRNA was isolated from MDCK cells using the Microfast Track isolation kit (Invitrogen) and treated with DNase I before cDNA synthesis. Reverse transcription was carried out as described (Sefton et al. 1998), and PCR to amplify coding fragments for canine Cyclins D1 and D2 was performed over 35 cycles at an annealing temperature of 55°C using primers as follows. For Cyclin D1, primers derived from the mouse sequence were used: forward, 5'-CTGCGAAGTGGAGACCAT CCG-3'; and reverse, 5'-GTCCGGGTCACACTTGATGAC-3' (mouse-specific primers). For Cyclin D2, degenerate primers were used: forward, 5'-GAA/GGAA/GC/AGITAT/CT/CTIC CICAA/GTG-3'; and reverse, 5'-GAA/GTACATIGCA/GAAT/CTTA/GAAA/GTC-3'. The amplified fragments were subcloned in pGEMT-easy and sequenced. After sequencing, dog-specific primers were designed and after a similar amplification protocol, the fragment was subcloned, sequenced, and used for Northern blot analysis.

Northern blots

Total RNA was isolated as described by Chomczynski and Sacchi (1987). For each assay, 20 μg of denatured RNA was used per

lane. The coding fragments for canine Cyclins D1 and D2 obtained by RT-PCR as described above were labeled using the Rediprime II kit (Amersham Biosciences). A human GAPDH (glyceraldehyde-3-phosphate dehydrogenase) probe was used as a control for the quantity of RNA.

Transient transfections and promoter analysis

To analyze the *Cyclin D2* promoter, mouse epidermal keratinocyte MCA3D cells were cotransfected with 40 ng of Renilla vector and 400 ng of pGL3 vector (Promega) containing the *Cyclin D2* promoter fused to the Luciferase reporter gene together with 25 ng of human SNAIL or mouse Snail complete coding sequences in pZeo (Invitrogen) and pcDNA3 (Invitrogen) vectors, respectively. Control transfections were performed with the empty plasmids (Mock). Transfections were carried out using Lipofectamine (Life Technologies). Luciferase and Renilla activities were assayed using the dual-luciferase reporter system kit (Promega), and the activity normalized to that of the promoter cotransfected with the control vectors. Human D2-Luc (1624 base pairs) and the deletions -1303 D2-Luc and -444 D2-Luc were kindly provided by Brad H. Nelson (Martino et al. 2001). The D2-Luc (-1624) was used to delete and mutate the two E-boxes present in the human *Cyclin D2* promoter with a Quickchange Site Directed Mutagenesis Kit (Stratagene). The core sequence 5'-GCACGTGC-3' contained in the two E-boxes of the human *Cyclin D2* promoter was independently mutated to 5'-TTACGTGC-3'. These two point mutations (GC to TT) abolish the E-box and were shown to derepress the proximal mouse E-cadherin promoter (Cano et al. 2000).

In situ hybridization

Whole-mount in situ hybridization was carried out in chick and mouse embryos at several stages of development as described (Nieto et al. 1996). Digoxigenin-labeled probes were synthesized from the full-length cDNA of mouse *Snail* and chick *Slug*. Probes for the mouse *Cyclins D1* and *D2* were synthesized from cDNAs obtained from the mouse epithelial cell line NMuMG and amplified by RT-PCR with the primers and conditions described above. Following hybridization, the embryos were incubated with alkaline phosphatase-conjugated anti-digoxigenin antibody. The alkaline phosphatase activity was detected by incubation with NBT/BCIP substrates (Roche). After hybridization, embryos were fixed in 4% paraformaldehyde in PBS, washed, and photographed in whole mount under a Leica M10 dissecting scope. Subsequently, mouse embryos were embedded in paraffin (Fibrowax), sectioned at 15 μm, and photographed using a Leica DMR microscope.

Cell death analysis in cell lines and embryos

Cells were grown on coverslips in 6-cm cell culture dishes and stained with propidium iodide (0.05 mg/mL) in PBS for 15 min at room temperature in the dark. Only cells with altered membrane permeability were stained and could be visualized using a Leica DMR microscope.

For the analysis of apoptosis in embryos, DNA fragmentation was detected using the TUNEL in situ cell death detection kit (Roche) or by staining with Nile Blue Sulphate (NBS; Sigma) that marks dying cells. For TUNEL detection, whole embryos were fixed in 4% paraformaldehyde in PBT at 4°C, dehydrated and treated for 2 h at room temperature in 100% methanol plus 1% H₂O₂, and washed several times in methanol. After rehydration the embryos were digested with proteinase K (10 μg/mL for 3 min) at room temperature, washed, and fixed for 30 min in

4% paraformaldehyde. After several washes, they were treated with the reaction mix for 2 h at 37°C, washed, blocked for 2 h with blocking solution (KTBT 0.1% Triton X-100, 15% FCS, 0.7% blocking powder from Roche), and incubated with the POD-converter (1 h at 37°C). Embryos were developed in the dark with DAB (3,3'-Diaminobenzidine, Sigma) containing 0.03% H₂O₂. After washing, embryos were photographed and embedded in 0.5% gelatin (Sigma) prior to being sectioned at 40 µm on a vibratome. NBS staining was carried out on whole embryos immediately after dissection. The embryos were incubated in NBS (20 µg/mL in PBS) containing 0.1% Tween 20 for 30 min at room temperature, briefly washed in PBS, and immediately photographed in 4% PFA.

Analysis of caspase-3 and caspase-8 activities

Cells attached to the dish and those in the supernatant were collected and lysed at 4°C in 5 mM Tris-HCl at pH 8.0, 20 mM EDTA, and 0.5% Triton X-100. Ac-DEVD-AMC and Ac-IETD-AFC were used as substrates to measure the enzymatic activities of caspase-3 and caspase-8, respectively (Herrera et al. 2001). Analyses were performed in a Luminescence Spectrophotometer (Perkin-Elmer LS-50). A unit of caspase activity was defined as the amount of active enzyme necessary to produce an increase of 1 arbitrary luminescence unit in 2 h. The protein concentration of the cell lysates was determined with the Bio-Rad assay kit, and the results are presented as units of caspase activity per µg of protein.

PI3-kinase activity

After solubilization of cells in lysis buffer (10 mM Tris-HCl, 5 mM EDTA, 50 mM NaCl, 30 mM NaPPi, 50 mM NaF, 100 µM sodium o-vanadate, 1% Triton X-100 at pH 7.6) containing leupeptin (10 µg/mL), aprotinin (10 µg/mL) and 1 mM PMSF, lysates were clarified by centrifugation, and the proteins were immunoprecipitated with a monoclonal anti-Tyr-Phosphate antibody (Py72). Immunoprecipitates were used to analyze PI 3-kinase activity by in vitro phosphorylation of phosphatidylinositol as described (Valdés et al. 2004).

Chick embryo electroporation

In ovo electroporation was essentially carried out as described (Del Barrio and Nieto 2002) with the following modifications: pCX-EGFP construct (1 mg/mL; Ikawa et al. 1995) was co-electroporated with pCX-Slug (1.5 mg/mL) containing the full-length chick Slug cDNA or with the empty pCX vector as a control. The DNAs were injected into stage 8 chick hindbrains in ovo and electroporated using two 50-msec 10 V pulses. The embryos were allowed to develop for a further 14–16 h. In all the experiments the control side was to the left. Embryos were photographed in ovo to record GFP expression and processed for apoptosis analysis.

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