

Milestones in *Bacillus subtilis* sporulation research

Eammon P. Riley¹, Corinna Schwarz², Alan I. Derman² and Javier Lopez-Garrido^{2,*}

¹ Division of Biological Sciences, University of California, San Diego, La Jolla, CA, USA.

² Max Planck Institute for Evolutionary Biology, Plön, Germany.

* Corresponding Author:

Javier Lopez-Garrido, August-Thienemann-Str. 2, 24306 Plön, Germany; Phone: +49 4522 763 216;

E-mail: lopezgarrido@evolbio.mpg.de

ABSTRACT Endospore formation has been a rich field of research for more than a century, and has benefited from the powerful genetic tools available in *Bacillus subtilis*. In this review, we highlight foundational discoveries that shaped the sporulation field, from its origins to the present day, tracing a chronology that spans more than one hundred eighty years. We detail how cell-specific gene expression has been harnessed to investigate the existence and function of intercellular proteinaceous channels in sporulating cells, and we illustrate the rapid progress in our understanding of the cell biology of sporulation in recent years using the process of chromosome translocation as a storyline. Finally, we sketch general aspects of sporulation that remain largely unexplored, and that we envision will be fruitful areas of future research.

doi: 10.15698/mic2021.01.739

Received originally: 17.07.2020;

in revised form: 21.10.2020,

Accepted 03.11.2020,

Published 27.11.2020.

Keywords: *Bacillus subtilis*, sporulation, genetics, SpoIII^E, SpoIII^A, SpoIII^Q, sporulation history.

Abbreviations:

SASP – small acid-soluble protein,

Spo⁻ – asporogenous and oligosporogenous mutants,

Spo⁺ – sporulating strains,

SRS – SpoIII^E Recognition Sequence,

TCA – tricarboxylic acid,

TZM – 2,3,5-triphenyltetrazolium chloride.

INTRODUCTION

Since their discovery, bacterial endospores (a.k.a. spores) have attracted the attention of the scientific community and, at times, even the general public. This should not come as a surprise, as some properties of endospores could well be in the headlines of the popular press. Bacterial endospores are among the most resilient cell-types known, and can survive for very long periods of time without any nutrients. In fact, it has been claimed that viable endospores were isolated from the gut of extinct bees buried in Dominican amber more than 25 million years ago [1], evoking the imagery of Jurassic Park [2]. Furthermore, the use of *Bacillus anthracis* (anthrax) spores in biological warfare and, more recently, in bioterrorist attacks has brought the bacterial spore to the attention of modern society [3]. The study of endospore formation, however, is full of fascinating discoveries that, while subtle enough to escape the attention of the mainstream media, have contributed to shape our current perception of bacterial cells. This review aims to discuss some of the key findings that have shaped the sporulation field, through the lens of bacterial genetics.

A BRIEF HISTORY OF ENDOSPORE FORMATION

Arguably the most important milestone in endospore research was the actual discovery of endospores. The first reported observation of bacterial endospores dates back to 1838, when Christian Gottfried Ehrenberg noted refractile bodies inside bacterial cells [4]. It was not until nearly four decades later that endospores started to be characterized in seminal studies by Ferdinand Cohn and Robert Koch. Cohn initiated the study of the resistance properties of spores with the observation that *B. subtilis* spores survive periods of boiling [5]. Koch, in collaboration with Cohn, followed the sporulation-germination cycle in *B. anthracis* [6], and realized that spores and vegetative cells are different cellular forms of the same bacterial species, and that cells can interconvert between these two forms, from vegetative cells to spores via sporulation, and from spores to vegetative cells via germination. Their hand-drawn sketches of these processes are shown in **Fig. 1**.

At the same time, John Tyndall independently postulated the existence of latent bacterial forms able to survive extreme temperatures. He described a sterilization technique (now designated “tyndallization”) that eradicates

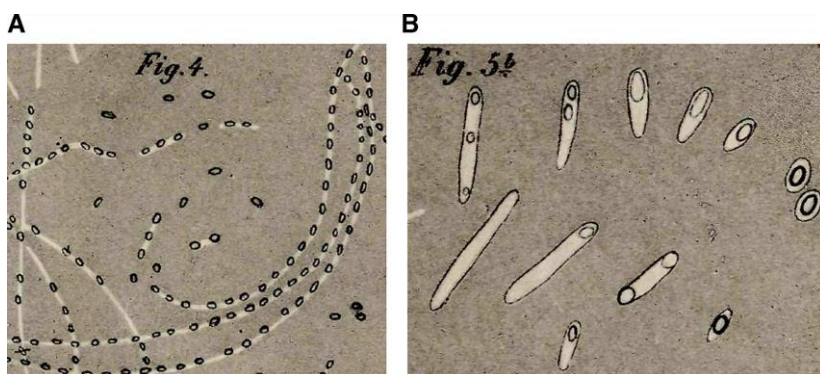


FIGURE 1: Early depictions of sporulation and germination in *Bacillus* spp. In one of his most well-known studies [6], Robert Koch investigated the etiology of anthrax. **(A)** Koch followed the sporulation process in *B. anthracis*. He placed a slice of spleen containing the *Bacillus* into cow sera or aqueous humor, and incubated the specimens at 35–37°C in an incubator that he himself had constructed. During the incubation, he observed the cells growing into string-like structures. After 10–15 h, these strings contained light refracting bodies, which he identified as spores. Koch depicted the structures in this sketch and likened them to fragile strings of pearls. The strings gradually

decomposed, the spores were released and sank to the bottom of the droplet where they accumulated and could be kept for weeks. **(B)** Koch followed spore germination and outgrowth, and determined that spores can form viable cells. He fixed dried spores on a slide and incubated the sample with aqueous humor. After 3–4 hours, he observed various stages of germination and outgrowth, which are depicted in Cohn's sketch. Koch described spores as egg-shaped structures, enclosed by a thin layer of protoplasm that he called bright matter. He hypothesized that during outgrowth this bright matter stretched and became the vegetative cell, whereas the spore remained at one cell pole, lightened, and shrank, before it dissociated and disappeared. He proposed that the spore core consists of an oily substance necessary for the cell to resume vegetative growth.

spores from culture media through the application of intermittent heating periods that kill vegetative cells, interspersed with periods of no heat that permit the spores that survive to germinate into vegetative cells, which are then killed during subsequent heating periods [7].

Throughout the next few decades, researchers attempted to characterize the process of endospore formation in different bacterial species, but it was difficult for them to achieve a unified understanding due to the limitations of the optical microscopy and staining methods available at that time [8, 9]. Some researchers, led by Koch, thought that spores were formed through the growth of a single refractile granule inside the cytoplasm of the vegetative cell [6]. A related model postulated that spores were not the result of the growth of a single granule, but of the fusion of multiple sporogenic granules inside the vegetative cell [10]. These putative sporogenic granules were probably cytoplasmic inclusion bodies that are present in various species of endospore forming bacteria, and are unrelated to spore formation. A different model, which comports better with our current perception of endospore formation (see below), proposed that spores were formed through the condensation of part of the vegetative cell protoplasm, which gained refractility over time [11]. A shared feature of these models, however, was that spore formation happened close to a single cell pole.

The advent of electron microscopy allowed for a more comprehensive cytological model for sporulation to be developed. In the late 1950's and early 60's, Young and Fitz-James provided detailed cytological descriptions of endospore formation in *Bacillus cereus* [12, 13]. Electron microscopy studies on sporulation in other *Bacillus* and *Clostridium* species were published shortly thereafter [14–16]. By the mid 60's, the field had achieved a consensus view of the cytological transformations leading to endospore formation, which are highly conserved among different species.

In 1965, Antoinette Ryter defined six different stages of sporulation [17], based on her own studies of *B. subtilis* and on those of Young and Fitz-James in *B. cereus*, and this classification was later expanded to eight stages [18, 19]. An up-to-date description of the different stages in *B. subtilis*, including details derived from fluorescence microscopy and genetic studies, is shown in **Fig. 2**. It is important to note that this classification represents discrete cytological steps of a process that is actually continuous. Thus, not only do several steps occur concomitantly, but also details about the transition between different stages are often neglected. For example, the transition between stages II and III happens through a phagocytosis-like process called engulfment, in which the membrane of the mother cell migrates around the forespore, generating the cell within a cell that is a hallmark of endospore formation (**Fig. 2**); the synthesis of the different spore layers is also gradual and starts shortly after polar septation (**Fig. 2**) [20]. Nevertheless, the classification proposed by Ryter represents a conceptual framework that facilitated the subsequent genetic dissection of spore formation.

B. SUBTILIS AND THE GENETIC ANALYSIS OF SPORULATION MUTANTS

During the first half of the 20th century, sporulation was studied simultaneously in different bacterial species. Two major discoveries, however, positioned *B. subtilis* to be the ideal model system for characterizing the process of sporulation during the second half of the century: in 1958 John Spizizen reported that *B. subtilis* can be transformed with DNA [21]; shortly thereafter, Thorne and Takahashi independently isolated phages capable of mediating generalized transduction in the same organism [22–25]. These powerful tools opened new avenues for genetic studies, and several laboratories turned their attention to *B. subtilis* to perform thorough genetic analyses of sporulation.

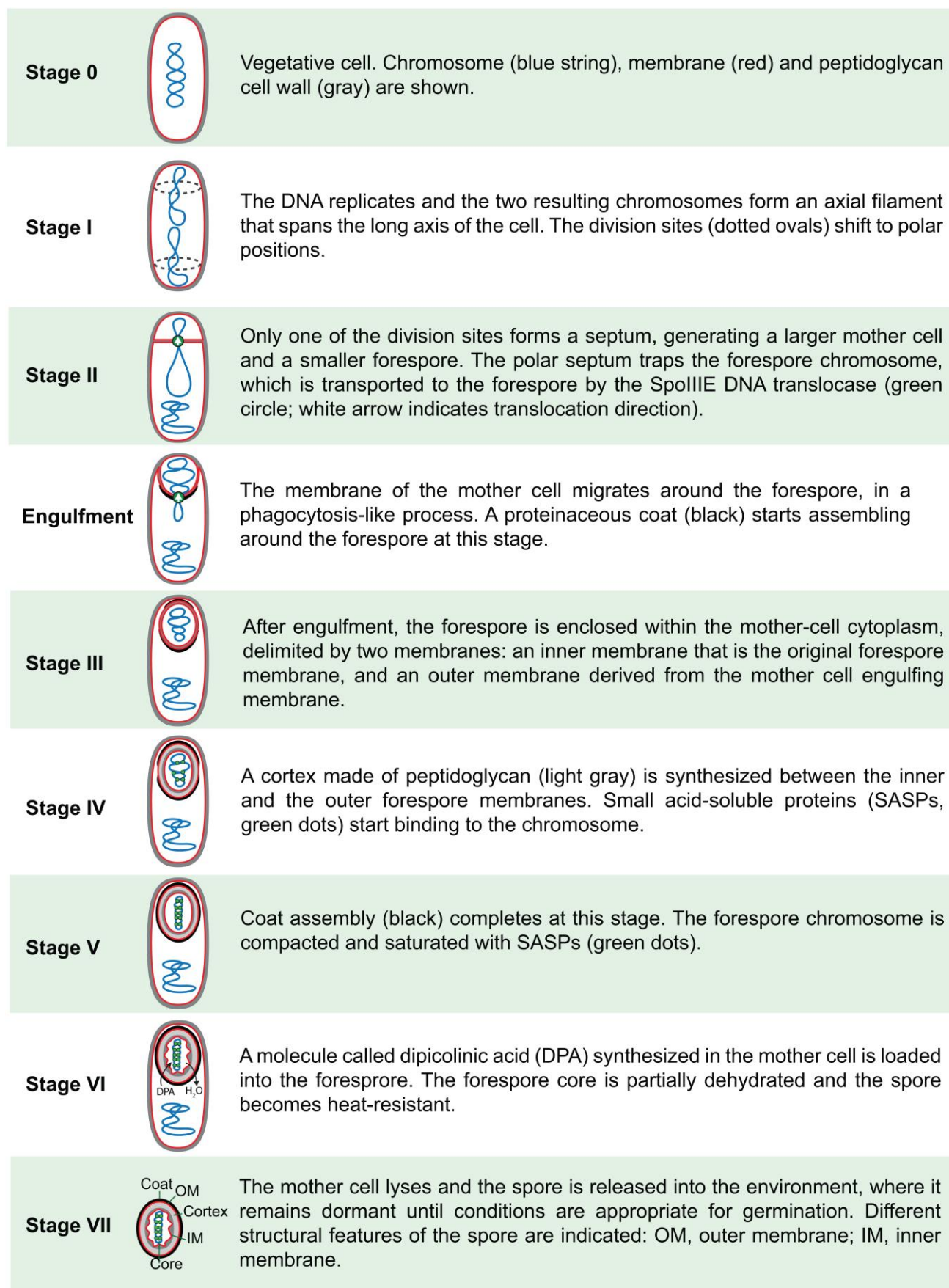


FIGURE 2: Sporulation stages in *B. subtilis*.

Mutants unable to form endospores (asporogenous), or with significantly reduced ability to form spores (oligosporogenous) have been reported for different species of endospore-forming bacteria ever since the inception of sporulation research [26]. The defining characteristic of oligosporogenous mutants is that the spores produced by them yield vegetative cells with equally low ability to produce spores. In contrast, asporogenous mutants, by definition, do not form viable spores, unless they accumulate secondary mutations that suppress the asporogenous phenotype. Such spores yield vegetative cells with significantly increased ability to form new spores compared to the asporogenous parental strain. For simplicity, we will refer to both asporogenous and oligosporogenous mutants as Spo⁻, and strains able to sporulate efficiently will be referred to as Spo⁺.

Researchers quickly devised simple, forward genetic screens to isolate Spo⁻ mutants in large numbers, and mapped the mutations using classical genetic methods such as transformation and transduction [27, 28]. Those studies were facilitated by the development of simple tests to distinguish Spo⁺ and Spo⁻ strains. Spo⁻ mutants form white colonies in certain rich media, whereas strains able to sporulate efficiently form brown colonies. The brown color is due to the accumulation of a pigment whose synthesis requires a component of the spore coat, CotA [29, 30]. Secondly, the heat resistance of the spores provides a definitive test to distinguish Spo⁺ and Spo⁻ strains [31]. Determining the linkage between mutations conferring the Spo⁻ phenotype and different markers in the chromosome led to the understanding that there are multiple sporulation (*spo*) loci, scattered around the chromosome [27, 28, 32–34].

The initial analysis of Spo⁻ mutants indicated that sporulation was blocked at different stages in different Spo⁻ mutants [31], suggesting that each sporulation stage was controlled by a dedicated set of genes. This was formally confirmed in 1966, when Ryter and collaborators performed a cytological characterization of *B. subtilis* strains carrying different *spo* mutations using electron microscopy, and classified them according to the stage at which sporulation was blocked [18]. This study led to the establishment of a specific nomenclature for the *spo* loci that was formal-

ized in a classic review by Piggot and Coote in 1976 [19]. The *spo* loci were defined as those, that when mutated, impaired the ability of the cell to form spores, without having a noticeable effect on vegetative growth. The loci were classified into different categories (*spo0*, *spoll*, *spolll*, *spolV*, and so on), according to the stage at which sporulation was blocked in the mutants (Fig. 2). No distinction was made between stages 0 and I, and mutations that caused blockage in either stage were globally referred to as *spo0*. A capital letter was used to distinguish different loci, mutations in which produced blockages at the same stage. Thus, for example, *spollIA* and *spollIE* refer to two distinct loci that, when mutated, produce a blockage at stage III. In some cases, it was later discovered that a specific locus was actually an operon containing several genes. In those cases, the different genes of the operon were assigned a second capital letter (for example, the *spollIA* locus actually consists of an operon of at least eight genes, which are named in order from *spollIAA* to *spollIAH*).

In addition to the *spo* genes, three more developmental gene categories were described: *ger*, *cot* and *ssp* (Fig. 3). Many *ger* genes were identified in elegant genetic screens for mutations that rendered spores unable to germinate properly [35–40]. These screens relied on an agar plate assay in which colonies formed by mutagenized cells were treated with heat or chloroform vapor to kill vegetative cells [35]. The remaining spores were then overlaid with agar containing two key components: (i) nutrients to induce germination, and (ii) 2,3,5-triphenyltetrazolium chloride (TZM), which turns red when reduced by metabolically-active cells. Spores are metabolically dormant, and are unable to reduce TZM to produce the red pigment, but upon germination become metabolically active and can reduce TZM to give the colonies a pink coloration. Colonies containing germination-deficient spores fail to reduce TZM, and therefore, do not produce the red pigment. While some of the *ger* genes identified in germination screens encode proteins involved exclusively in germination (for example, the genes encoding germinant receptors, such as those in the *gerA* and *gerB* operons), others encode proteins that are actually required for proper spore development, generating some overlap between *ger* and *spo* genes (for instance, *gerM* is required for proper sporulation, but

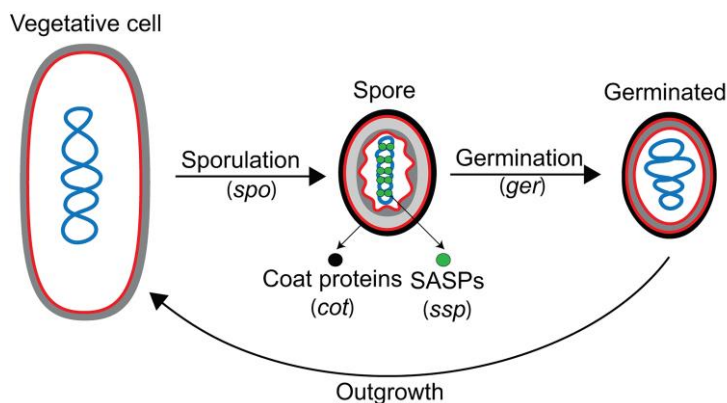


FIGURE 3: Developmental loci in *B. subtilis*. Diagram of the sporulation-germination cycle, with the different classes of developmental loci noted (in parentheses). The *spo* loci are required for spore formation, but not for vegetative growth. The *ger* loci are required for proper spore germination. After germination, metabolism is reactivated and the spore transforms into a growing vegetative cell in a process called outgrowth, which is thought to depend on the same pathways that control vegetative growth. The *cot* and *ssp* loci were identified after the isolation and characterization of coat and small acid-soluble proteins (SASPs), respectively, from spores. In addition to these categories, there are hundreds of developmentally-regulated genes whose roles in sporulation and germination are not yet understood. Membranes, red; DNA, blue; cell wall, gray; spore cortex, light gray; spore coat, black; SASPs, green circles.

it was first identified in a screen for germination mutants [41, 42]).

The two major remaining categories of developmental genes, *cot* (for coat) and *ssp* (for spore-specific protein), were identified by reverse genetics, after the isolation and biochemical characterization of coat proteins and small acid-soluble proteins (SASPs) [29, 43–47] from spores, respectively (Fig. 3). SASPs are abundant spore proteins that saturate and protect the spore chromosome [45]. Since individual SASPs and most coat proteins are dispensable for *B. subtilis* sporulation under standard laboratory conditions [29, 46, 48, 49], they were missed in the genetic screens that led to the identification of *spo* genes.

The discovery and characterization of developmental genes accelerated after the rise of DNA cloning and sequencing technology. Researchers were able to clone *spo* genes by selecting for chromosomal DNA fragments that complemented the respective *spo* mutations. From these clones, *lacZ* fusions were generated, and these in turn allowed for the study of the *spo* genes *in vivo*. New genetic tools, such as Tn917-based transposition mutagenesis [50–52] and integrational plasmid vectors [52–55], further facilitated the identification and manipulation of *spo* loci. During the late 1970's and 80's, many developmental loci were cloned and sequenced, their expression characterized, and the role of the proteins they encode started to be deciphered.

CELL-SPECIFIC GENE EXPRESSION

Three main conclusions arose from the initial genetic analysis of sporulation: first, most developmental genes are not expressed in growing cells, and their expression is induced at different times during sporulation; second, there is a dependency hierarchy in developmental gene expression, such that the expression of genes that participate in later events in the sporulation pathway tends to depend on the expression of genes that participate in earlier events; third, the majority of the sporulation genes are expressed in only one of the two cells required to form a spore, either the mother cell or the forespore. This last conclusion was achieved by using fractionation techniques to separate forespore and mother-cell protoplasts [56–58], and by the implementation of genetic strategies to determine the cell in which different *spo* genes were required [59, 60]. In one such genetic strategy, developed by Lencastre and Piggott [59] and illustrated in Fig. 4, Spo⁻ mutants were transformed with genomic DNA from the wild-type (Spo⁺) strain at the start of sporulation, thereby rendering the Spo⁻ mutants capable of forming heat-resistant spores. The parental Spo⁻ mutants fell into two categories, depending on the ability of the resultant spores to produce additional spores following germination: mutants in the first category produced spores that were homogeneously Spo⁺; mutants in the second category produced a heterogeneous population of Spo⁺ and Spo⁻ spores. The *spo* genes mutated in the first category of parental strains were inferred to be required in the forespore, and the ones mutated in the second were inferred to be required in the mother cell. The logic under-

lying this interpretation is as follows (Fig. 4): of the two cells required to form a spore, only the forespore retains its chromosome in the mature spore, since the mother cell lyses upon sporulation completion (Fig. 2). Thus, *spo* genes required in the forespore must be complemented in the forespore chromosome and the wild-type version of the gene will therefore always present in mature spores, and these will give rise to a homogenous population of Spo⁺ transformant spores. But if a *spo* gene is required in the mother cell and not in the forespore, then sporulating cells in which only the mother-cell chromosome is transformed with the wild-type allele will be able to form mature spores, but the non-transformed forespore chromosome would still contain the mutated version of the *spo* gene, and the spores would remain Spo⁻.

Results from these biochemical and genetic analyses underscored that different genetic programs were activated in the two cells, at different times during sporulation. From here, the focus moved on to deciphering the genetic regulation of sporulation.

In 1969, it was discovered that bacterial RNA polymerases contain a factor, called Sigma (σ), that is necessary for promoter recognition and transcription initiation [61, 62]. This finding led researchers to hypothesize the existence of alternative σ factors, which could direct RNA polymerase to specific subsets of promoters. The existence of such alternative σ factors was first demonstrated in *B. subtilis* phage SP01 [63], which encodes σ factors that drive expression from middle and late phage promoters. The first bacterial alternative σ factor to be discovered was σ^{37} [64], a general-stress response σ factor of *B. subtilis*. Initially, the name of the different σ factors was based on their molecular weight, in kDa. Thus, σ^{37} was ~37 kDa, as determined by the apparent size of the purified protein in an acrylamide gel. However, the discovery of alternative σ factors with similar sizes or that migrated aberrantly in acrylamide gels accelerated over the following years, which led researchers in the *B. subtilis* field to replace the molecular weight designation with a letter reflecting order of discovery [65, 66]. Thus, σ^{37} was renamed σ^B , the name by which it is commonly known today, and the genes encoding the σ factors were named *sig* followed by their corresponding letter.

Shortly after the discovery of σ^B , it was shown that the sporulation regulatory program also relies on alternative σ factors that are activated in the mother cell or in the forespore at different developmental stages. The first sporulation-specific σ factor to be discovered was σ^E (originally referred to as σ^{29}) [67]. σ^E was discovered using predominantly biochemical and molecular biology techniques [67]. The protein was isolated from RNA polymerase purified from sporulating cultures, and was shown to change the promoter specificity of RNA polymerase *in vitro*. Not surprisingly, it was subsequently shown that σ^E was encoded by a previously identified *spo* locus, *spolIGB* (later renamed as *sigE*) [68, 69]. After σ^E , three additional sporulation σ factors were identified by means of both sequence similarity with previously identified σ factors, and *in vitro* transcription assays of sporulation genes: σ^F [70, 71], σ^G [72–

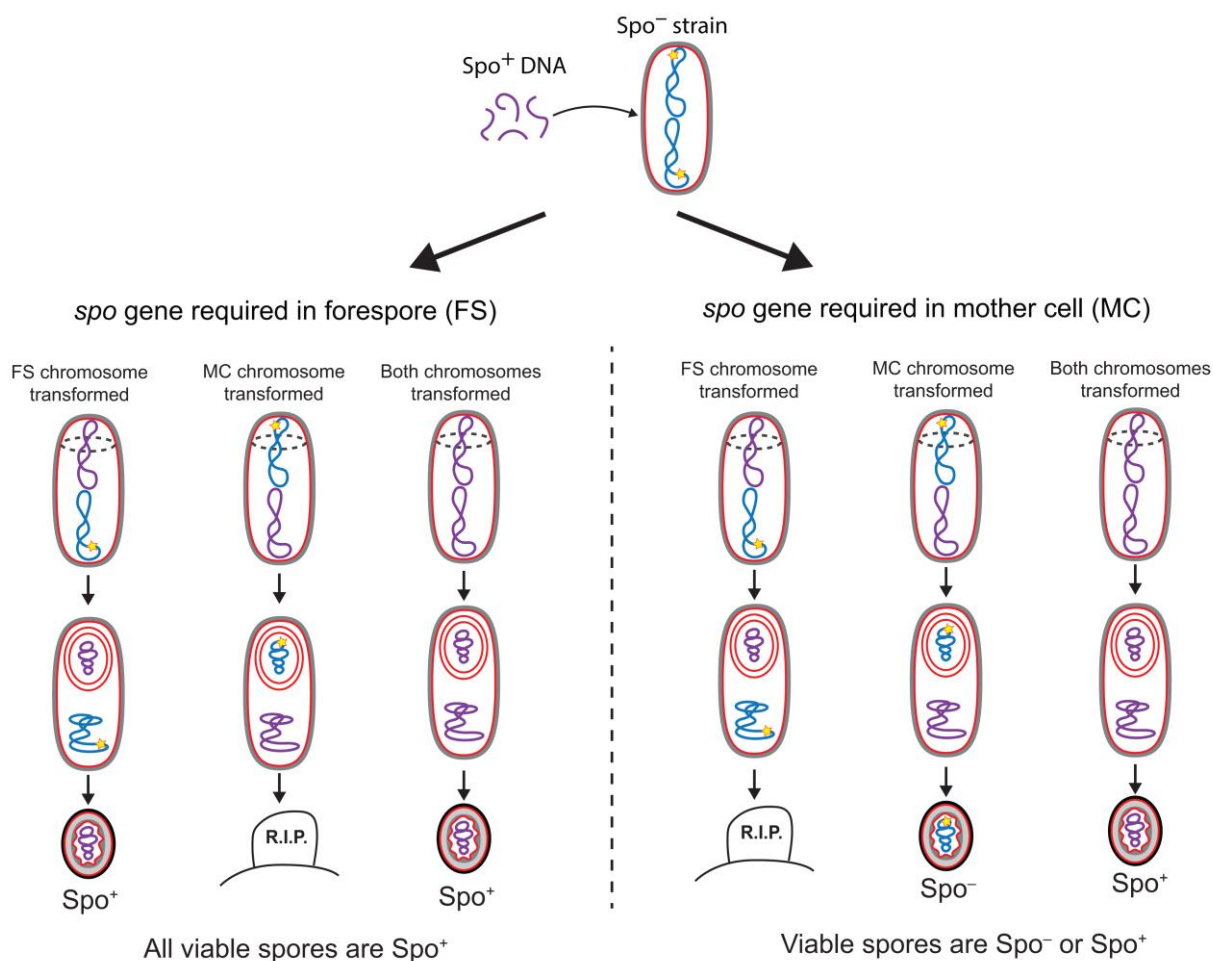


FIGURE 4: Genetic strategy to identify the cell in which a *spo* gene is required. A *Spo*⁻ strain with a mutation in a known *spo* gene (yellow star) is depicted in Stage I (see Fig. 2) with its two chromosomes, one that will be packed into the forespore (upper chromosome) and one that will remain in the mother cell and be destroyed when the mother cell lyses (lower chromosome). The strain is transformed with genomic DNA from a *Spo*⁺ strain at the onset of sporulation. Transformed chromosomes are drawn in purple, non-transformed chromosomes, in blue. Either or both chromosomes are capable of being transformed to *Spo*⁺. If the *spo* gene is required in the forespore (**left panel**), transformation of the forespore chromosome to *Spo*⁺ rescues the process and generates spores that can germinate and go on to sporulate again (left). But transformation of the mother-cell chromosome to *Spo*⁺ with no accompanying transformation of the forespore chromosome leaves the forespore chromosome *Spo*⁻ so the cells cannot complete the process, and no spores are produced (center). Hence only *Spo*⁺ spores are generated if the *spo* gene is required in the forespore. If the *spo* gene is required in the mother cell (**right panel**), transformation of the mother-cell chromosome to *Spo*⁺ rescues the process (center and right), but only if the forespore chromosome is transformed as well will the spores that are produced be able to germinate into cells that can go on to sporulate again (right). Hence, both *Spo*⁺ and *Spo*⁻ spores can be generated if the *spo* gene is required in the mother cell.

74] and σ^k [75]. In *B. subtilis*, the activation of the different sporulation σ factors follows a hierarchical order in which different σ factors are sequentially activated in alternating cellular compartments, and the activation of a later factor depends on the activation of the previous one (Fig. 5): first, σ^f becomes active in the forespore shortly after polar separation, followed by σ^e in the mother cell. Roughly coincident with engulfment completion, σ^g is activated in the forespore and, finally, σ^k is activated in the mother cell. The variety of mechanisms involved in the sequential activation of these σ factors includes phosphorelays, proteolytic processing, and even a gene splicing event [76–79].

After the discovery of the sporulation-specific σ factors, many of the developmental genes previously identified were assigned to different regulons, depending on the σ factor controlling their expression. Microarray technology allowed researchers to discover additional genes under the control of sporulation σ factors [80, 81]. Together the cell-specific σ factors control the expression of more than 500 different genes [82, 83], the majority of which are not expressed during vegetative growth. Over the course of the past few decades, the study of sporulation has focused on deciphering the function of genes that are under the control of sporulation-specific σ factors. But the functions of nearly 1/3 of these genes remain a mystery, and for the

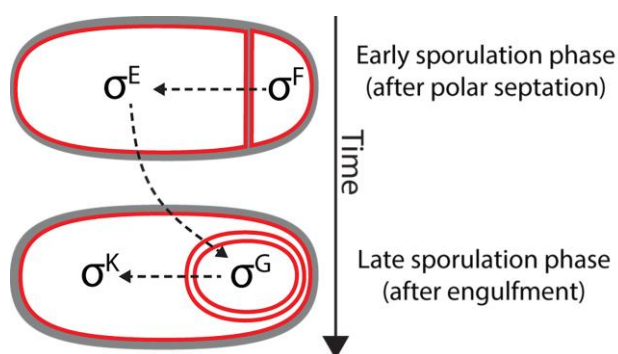


FIGURE 5: Cascade of sporulation-specific σ factors in *B. subtilis*. Immediately after polar septation, σ^F is activated in the forespore. σ^F activation triggers an intercompartmental signaling cascade that leads to the activation of σ^E in the mother cell. Together σ^E and σ^F control gene expression during engulfment. Roughly coincident with the completion of engulfment, σ^F is replaced by σ^G in the forespore, which leads to the subsequent activation of σ^K in the mother cell. σ^G and σ^K control gene expression in the forespore and the mother cell, respectively, after engulfment.

majority of sporulation genes with assigned function, we have only a superficial knowledge of how specific gene products contribute to spore formation. We envision that the study of developmentally-regulated genes will continue to be a fruitful area of research for years to come.

CELL-SPECIFIC GENE EXPRESSION AS A GENETIC TOOL

Beyond its central importance to the developmental program of sporulation, cell-specific gene expression has come to provide a powerful genetic tool for the study of different aspects of spore formation. After the discovery of the sporulation-specific σ factors, many researchers carefully characterized promoters regulated by the different σ factors (i.e. [84–92]), enabling these promoters to be co-opted for expression of heterologous genes in either the mother cell or the forespore, at different developmental stages. Cell-specific gene expression has been used to study many of the fundamental features of spore formation such as the temporal compartmentalization of the different sporulation σ factors [93], the organization of the chromosomes after sporulation initiation [94], and the transport of the forespore chromosome from the mother cell to the forespore [95–101], among other things. In order to illustrate the power of this approach, we describe three experiments that have helped to disentangle, at least partially, the function of the sporulation loci *spoIIIA* and *spoIIQ* (Fig. 6):

Mother-cell proteins encoded in the *spoIIIA* operon (which contains eight genes, *spoIIIAA* to *spoIIIAH*) together with the forespore protein SpoIIQ form a trans-envelope multimeric complex (henceforth A-Q complexes) that spans the mother-cell and forespore membranes (Fig. 6A) [102, 103]. Mutants lacking *spoIIIA* or *spoIIQ* are asporogenous, as they fail to activate the late forespore σ factor, σ^G [104, 105]. Some of the SpoIIIA proteins show sequence and structural homology to components of various secre-

tion systems from Gram-negative bacteria [103, 106–108], and one of them, SpoIIAG, has been shown to form multimeric rings *in vitro* [109, 110], leading to the proposal that A-Q complexes constitute proteinaceous channels that connect the mother cell and the forespore, through which molecules required for σ^G activation might be transferred between the two cells. Meisner and collaborators developed a compartmentalized biotinylation assay to test this idea (Fig. 6A) [111]. The authors used mother cell- and forespore-specific promoters to express the gene encoding *E. coli* biotin ligase (*birA*) in either cell, and fused biotin acceptor peptides to the extracellular domains of A and Q proteins. They found that BirA produced in the forespore could biotinylate the extracellular domain of SpoIIAH, which supported the idea that A-Q complexes are channels that are open at least on the forespore side.

In a separate study, Camp and Losick took advantage of cell-specific gene expression to study whether A-Q channels were specifically required for σ^G activation, or if they also played a role in the expression of σ^G -independent genes (Fig. 6B) [112]. They monitored the activity of the heterologous RNA polymerase, T7 RNAP, when produced in the forespore from a σ^F -dependent promoter by placing a *lacZ* gene under the control of a T7 RNAP-dependent promoter. In the presence of A-Q, β -galactosidase activity could be detected at the onset of sporulation, and it increased gradually over time until it reached a plateau that was maintained throughout spore formation. In mutants lacking functional A-Q complexes, β -galactosidase activity was also detected at the onset of sporulation, but dropped off later in sporulation, at roughly the time when σ^G would be normally activated. These observations indicated that A-Q plays a general role in regulating forespore gene expression at late sporulation stages, rather than affecting σ^G activity specifically. The authors proposed that A-Q channels constitute a feeding tube through which the mother cell provides metabolic resources to the forespore in order to maintain forespore biosynthetic activities at late sporulation stages [112].

We have recently accumulated more evidence that the mother cell nurtures the forespore, by implementing a cell-specific protein degradation system that also relies on cell-specific gene expression (Fig. 6C and D) [113, 114]. The system uses modified *ssrA** tags from *E. coli* fused to the C-terminus of target proteins [114], and the cognate *E. coli* SspB adaptor protein produced under the control of mother-cell or forespore-specific promoters [113] (note that *E. coli* SspB is not related to the sporulation *ssp* genes described previously in this manuscript; to avoid confusion, *E. coli* SspB will be referred to as SspB^{Ec}). When SspB^{Ec} is produced, it binds to *ssrA** and delivers the target protein to the endogenous *B. subtilis* protease ClpXP for degradation (Fig. 6C and D) [114]. We are using this platform to systematically assess the requirement of different metabolic pathways in the mother cell and in the forespore for sporulation to progress normally. Our results indicate that the tricarboxylic acid (TCA) cycle is required in the mother cell, but is largely dispensable in the forespore (Fig. 6E) [115]. We have also observed that protein synthesis in the fore-

spore requires TCA cycle activity in the mother cell, suggesting that the mother cell produces TCA cycle-derived metabolic precursors, such as amino acids, for protein synthesis in both the mother cell and the forespore (Fig. 6E) [115]. These findings indicate that mother-cell metabolic

precursors are transported to the forespore to support protein synthesis, in line with the feeding tube model proposed by Camp and Losick [112].

Overall, cell-specific gene expression has been a fantastic addition to the list of genetic tools available in *B. subtilis*.

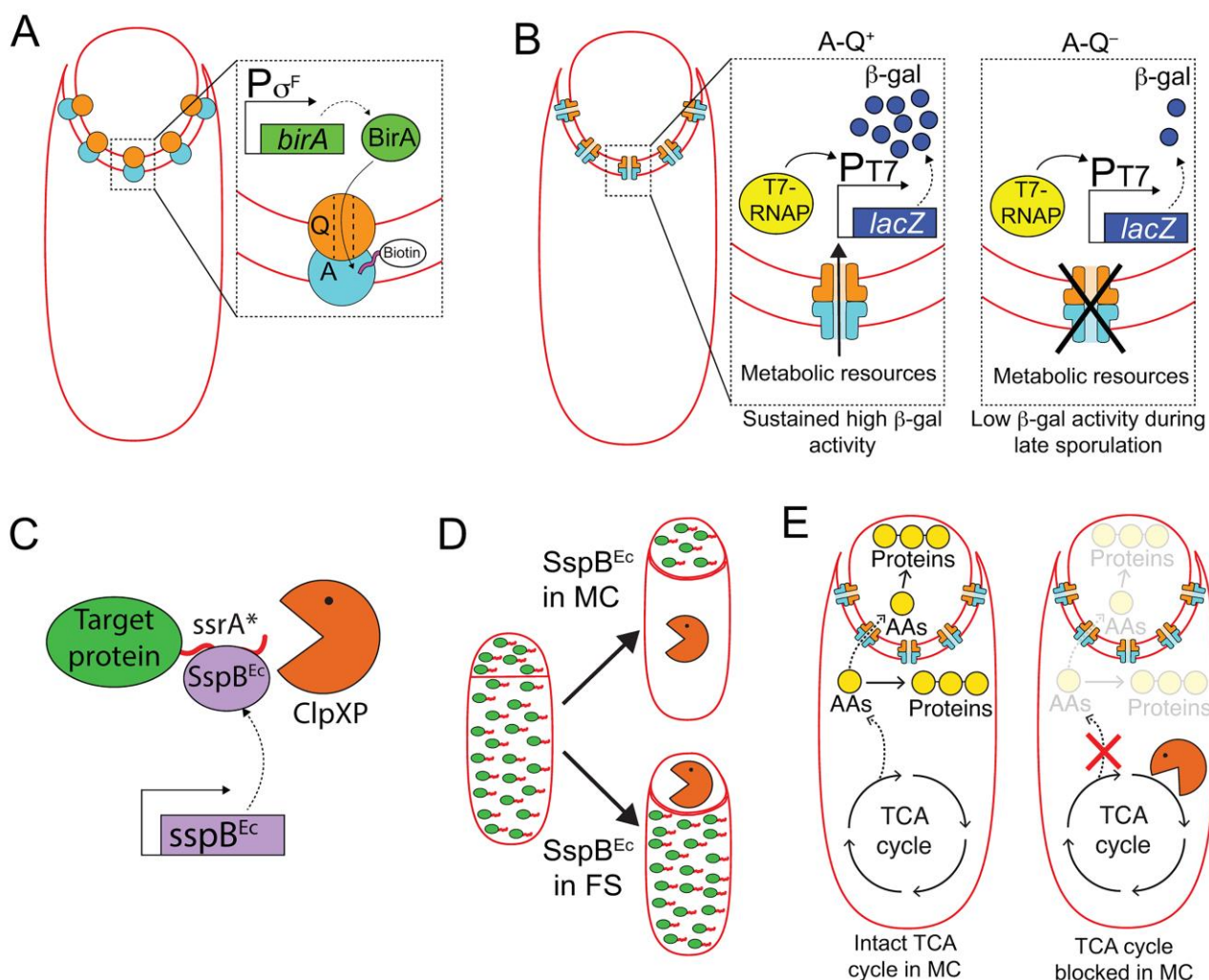


FIGURE 6: Harnessing cell-specific gene expression to study the function of A-Q complexes. (A) The forespore protein SpoIIQ (Q, orange circle) and the mother-cell proteins encoded in the *spoIIIA* operon (A, light blue circle), form trans-envelope complexes that bridge the forespore and mother-cell membranes during engulfment. Zoomed in panel: Biotin ligase (BirA, green) produced in the forespore is able to biotinylate a biotin acceptor peptide (pink) fused to the extracellular domain of the A protein, SpoIIIAH [111], indicating that A-Q complexes are channels (see panel B), and that the channel pore is open on the forespore side and large enough for BirA to reach the extracellular domain of SpoIIIAH. (B) The activity of a heterologous RNA polymerase (T7 RNAP, yellow) produced in the forespore under σ^F control is monitored by the accumulation of β -galactosidase (β -gal, dark blue circles) produced through the expression of a *lacZ* gene under the control of a T7 RNAP-dependent promoter. In the presence of A-Q channels (A-Q⁺, left zoomed in panel), sustained β -galactosidase activity is detected throughout sporulation. In the absence of A-Q channels (A-Q⁻, right zoomed in panel), β -galactosidase activity drops at later sporulation stages. Camp and Losick [112] proposed that A-Q channels constitute feeding tubes through which the mother cell transfers metabolic resources to the forespore to maintain biosynthetic activities at late sporulation stages. (C) The *ssrA*^{*}/*SspB*^{Ec} inducible protein degradation system [114]. *SspB*^{Ec} (purple) binds to *ssrA*^{*} (red) fused to the C-terminus of target proteins (green), and delivers the target proteins to the endogenous *B. subtilis* protease ClpXP (orange pacman) for degradation. (D) Cell-specific degradation of target proteins during sporulation is achieved by expressing *sspB*^{Ec} from mother cell- or forespore-specific promoters [113]. Target proteins represented by green ovals tagged with *ssrA* (red line); degradation represented by orange pacman. (E) Left cell: Mother-cell TCA cycle provides metabolic precursors, such as amino acids (AAs, yellow circles), to support protein synthesis in both the mother cell and the forespore [115]. Mother-cell metabolic precursors could be transported to the forespore via A-Q channels, in keeping with the feeding tube model. Right cell: Degradation of TCA cycle enzymes in the mother cell blocks protein synthesis in both the mother cell and the forespore [115].

We are looking forward to seeing new creative approaches using those tools to explore the inner workings of sporulating bacteria.

THE GENETIC BASIS OF CELLULAR DYNAMICS

The study of sporulation has helped to overturn the traditional dogma that bacterial cells lack a defined internal organization owing to the absence of membrane-bound organelles. Over the last 25 years, we have obtained precise descriptions of how proteins are redistributed during sporulation to mediate dynamic cellular processes. Advances in our understanding of sporulation during this period have been driven by the development of a wealth of new imaging technologies, particularly in fluorescence microscopy, which have allowed the field of bacterial cell biology to emerge. We illustrate the rapid progress made in this field by focusing on a single protein, SpoIIIE.

Since the mid 20th century, it was recognized that the two chromosomes resulting from a replication event prior to sporulation initiation are segregated during sporulation, such that one remains in the mother cell and the other is packed into the small forespore [12]. In 1994, Wu and Errington [116] reported that mutations in *spoIIIE*, one of the *spo* loci that was identified during the genetic analysis of sporulation mutants, prevented complete segregation of the forespore chromosome, such that only ~30% of the chromosome was present in the forespore with the rest still in the mother cell (**Fig. 7A**). They inferred that the polar septum trapped the forespore chromosome, and that SpoIIIE mediated the translocation of the chromosome across the septum into the forespore.

Further studies showed that SpoIIIE is a membrane-anchored FtsK-like protein that has a large C-terminal cytoplasmic motor domain [117] with DNA-dependent ATPase activity, that is capable of tracking along DNA in the presence of ATP [118]. More recently, *in vitro* studies with purified SpoIIIE motor domains have shown that they assemble into hexameric rings [119, 120], with an inner diameter large enough to accommodate a double-stranded DNA molecule [119]. Single molecule experiments using optical tweezers have allowed the characterization of the inter-subunit coordination [121] and of the mechanochemistry [122] of the hexameric rings as they move along DNA molecules, providing exquisite details of the mechanism of chromosome translocation at the molecular level.

SpoIIIE assembles into a translocation complex at the septal midpoint during the early stages of sporulation (**Fig. 7B**). This was shown first in fixed cells using immunofluorescence microscopy [117] and later in living cells with a SpoIIIE-GFP fusion and fluorescent membrane dyes that are compatible with live cell imaging [123] (**Fig. 7B**). More recently, studies of the assembly of the SpoIIIE translocation complex with super-resolution optical microscopy techniques [101, 124, 125] have revealed that SpoIIIE oligomers localize to the leading edge of the polar septum as it forms, and ultimately assemble a stable translocation complex at the septal midpoint. In mutants that produce thicker than normal polar septa, super-resolution micros-

copy has made it possible to resolve two different foci within the SpoIIIE complex (**Fig. 7C**), suggesting that the complex consists of two subcomplexes of opposite polarity, one anchored to the mother cell and the other to the forespore septal membrane. Each subcomplex contains enough SpoIIIE monomers to assemble at least two hexameric rings [101], which would allow the two arms of the chromosome to be translocated in parallel from the mother cell to the forespore (**Fig. 7D**) [98].

Assembly of the SpoIIIE translocation complex requires that the DNA be trapped at the septum [126], and the complex is in turn required to mediate septal membrane fission while the chromosome is translocated [124]. This has led to the proposal that the mother-cell and forespore SpoIIIE subcomplexes pair to form a continuous channel spanning both septal membranes (**Fig. 7D**). In support of this idea, it has been shown that both forespore and mother-cell subcomplexes are required to keep the septal membranes separated during chromosome translocation [101]. However, the mother-cell subcomplex by itself is sufficient to mediate the transport of the chromosome into the forespore [95, 101]. If the SpoIIIE mother-cell subcomplex is absent, the forespore subcomplex can transport the chromosome out of the forespore and into the mother cell [95, 101], indicating that the SpoIIIE translocation complex can, in principle, function as a bidirectional motor. During sporulation, however, the SpoIIIE translocation complex always transports the chromosome from the mother cell to the forespore (**Fig. 7D**). The mechanism that determines translocation directionality has been at least partially elucidated by means of a combination of *in vivo*, *in vitro* and *in silico* approaches. The motor domain of SpoIIIE is able to recognize specific, highly-skewed octameric sequences in the DNA called SpoIIIE Recognition Sequences (SRSs), which are present in each arm of the chromosome but on opposite strands [97]. Since the *B. subtilis* chromosome is circular and the two chromosome arms are trapped at the septum, the opposite orientation of SRSs on each arm might serve as a cue to determine translocation directionality. In fact, it has been shown that the interaction of SpoIIIE with the SRS in the orientation preferentially encountered by SpoIIIE as each chromosome arm is directed into the forespore stimulates SpoIIIE ATPase activity [120, 127, 128]. This might account for the direction of translocation and the simultaneous transport of both chromosome arms to the forespore.

In addition to guaranteeing that the forespore receives a full complement of genetic material, SpoIIIE-mediated chromosome translocation also contributes to the generation and maintenance of forespore shape during engulfment [100]. Packaging an entire chromosome into the small forespore compartment leads to a high turgor pressure, which inflates the forespore like a balloon and distends the forespore membranes (**Fig. 7E**).

While *spoIIIE* is among the best characterized *spo* loci, the field has made significant progress in the understanding of a myriad of dynamic processes associated with sporulation, such as the formation of the polar septum [129–131], the migration of the mother-cell membrane around

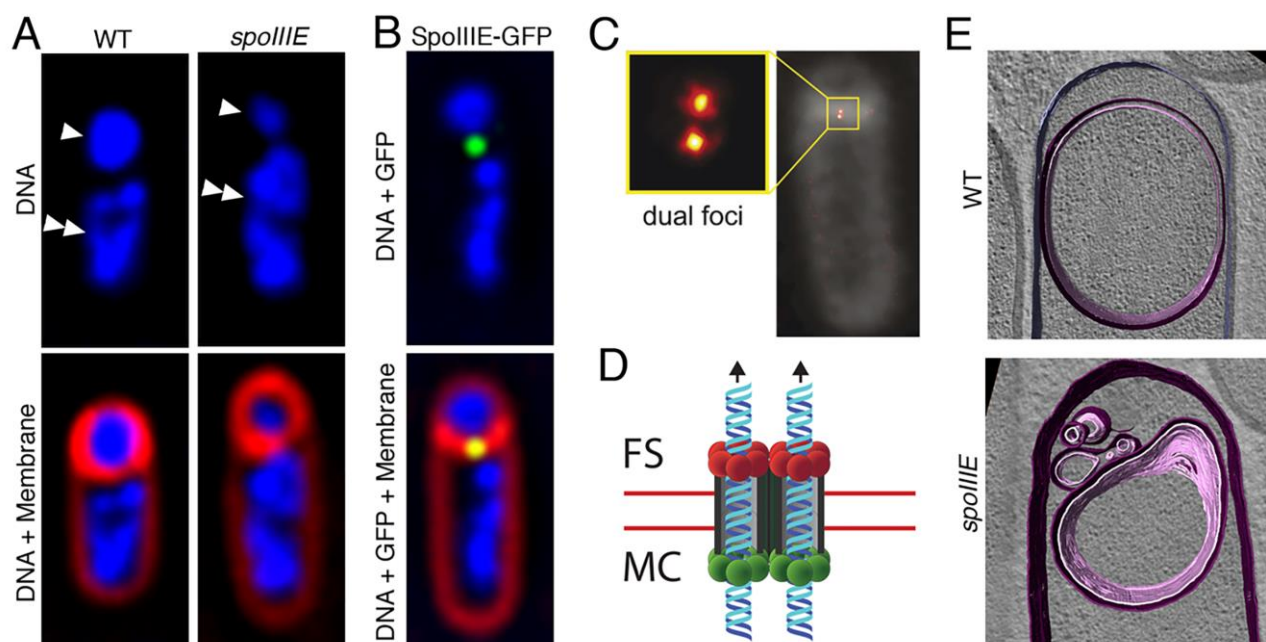


FIGURE 7: Progress in our understanding of SpoIIIE-mediated chromosome translocation. (A) Fluorescence microscopy images of a wild-type (WT, left) and of a *spoIIIE* mutant (*spoIIIE*, right) strain of *B. subtilis*. The upper panels show the DNA stained with DAPI (blue), and the lower panels the overlay of DAPI-stained DNA and the membranes stained with FM 4-64 (red). In the upper panel, forespores are indicated by single arrowheads, and mother cells by double arrowheads. Wild-type forespores contain a complete chromosome. However, forespores of *spoIIIE* mutant strains contain only ~30% of a chromosome, and the rest remains trapped in the mother cell. **(B)** Fluorescence microscopy image of a sporulating cell producing a SpoIIIE-GFP fusion during chromosome translocation. The upper panel shows the DNA stained with DAPI (blue) and the GFP signal, and the lower panel shows in addition the membranes stained with FM 4-64 (red). SpoIIIE forms a focus (green) at septal midpoint, where the chromosome is trapped. **(C)** SpoIIIE visualized by super-resolution microscopy (PALM) in living cells with thicker polar septa. SpoIIIE forms two foci (dual foci), which are separated by a distance equivalent to the septal thickness, indicating that one cluster is present at one side of the septum and the other at the opposite side. Reproduced from [101]. **(D)** Model for the organization and function of the SpoIIIE translocation complex at the septal midpoint. SpoIIIE forms two side-by-side channels spanning both septal membranes (red lines), thereby allowing the simultaneous transport of both arms of the chromosome from the mother cell (MC) to the forespore (FS). Translocation is powered by SpoIIIE motor domains at the mother-cell side of the septum (green circles), which are activated to export the chromosome to the forespore. Forespore motor domains (red circles) remain inactive. **(E)** Cryo-electron microscopy of a wild-type sporulating cell (top), and of a *spoIIIE* mutant (bottom). Membranes are annotated as follows: forespore membrane, pink; mother-cell membrane, purple. Chromosome translocation is required to maintain the shape of the forespore. In the absence of chromosome translocation, the forespore appears deflated. Reproduced with permission from [100].

the forespore during engulfment [132–139], the synthesis of the peptidoglycan cortex [140], and the assembly of the proteinaceous coat [20, 141], to name a few.

HISTORICAL CONTINGENCIES

Sporulation in *B. subtilis* is one of the most well-understood developmental processes, but there are still gaps in our understanding, which are due in part to the historical contingencies of sporulation research. Much of the sporulation research performed so far has focused on understanding the role of different developmental genes in spore formation. The definition of *spo* genes—those that, when mutated, impair the ability to form spores without affecting vegetative growth—has likely biased our understanding of sporulation, as it necessarily excluded all the essential genes and non-essential genes that are also required for optimal vegetative growth. It has therefore remained unclear whether essential housekeeping proteins involved in central metabolism, redox reactions, protein

folding, and translational regulation play a significant role in assembling the spore, and if so, if they are required in a specific cell or during a specific stage of sporulation. Recent high-throughput studies have identified vegetative proteins that likely play a role in sporulation [142, 143]. Genetic tools to interfere with the function of specific proteins in a precise, cell- and developmental stage-specific manner during spore formation have also recently been developed (Fig. 6C and D) [113], and these present a promising new approach for understanding the role played by these and other vegetative proteins during endospore formation. Although many facets of sporulation remain unclear, new technologies and innovative approaches will continue to advance our knowledge of this remarkable process.

A more general aspect of endospore formation that remains poorly studied is how the process functions in different species. Endospore formation occurs in a broad range of bacterial species that belong to an ancient and exceptionally diverse bacterial phylum, the Firmicutes

[144]. However, most mechanistic studies on sporulation over the previous 50 years have been done in *B. subtilis*, as this organism is particularly amenable to genetic studies. Placing the focus on a single species has provided us with extraordinarily detailed descriptions of sporulation but, at the same time, has prevented us from fully appreciating the diversity of endospore formation. In recent years, *Clostridioides difficile*—formerly known as *Clostridium difficile*—has become a model for endospore formation in anaerobic bacteria. Studies in this organism reveal that, although the general scheme of endospore formation is similar to that of *B. subtilis*, there are mechanistic differences that affect every sporulation stage, from initiation to germination [145–151]. In addition, while *B. subtilis* and *C. difficile* produce one spore per sporangium, there are some species that can produce two [152, 153], and others that can produce multiple spores [154–156], opening the intriguing possibility that the sporulation pathway was co-opted as a reproductive strategy in some species, or that sporulation evolved from what was originally a reproductive strategy [157–160]. Clearly, this is a rich and interesting area of future research that will require the development of new culturing techniques and genetic tools to manipulate non-model bacteria. Understanding the diversity

of endospore formation may allow us to address key questions regarding the ecological roles and the evolutionary origin of this fascinating developmental process.

ACKNOWLEDGMENTS

We thank Jan Böhning for assistance with translation of the Koch and Cohn publications. This work was supported by ERC starting grant 853323.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

COPYRIGHT

© 2020 Riley *et al.* This is an open-access article released under the terms of the Creative Commons Attribution (CC BY) license, which allows the unrestricted use, distribution, and reproduction in any medium, provided the original author and source are acknowledged.

Please cite this article as: Eammon P. Riley, Corinna Schwarz, Alan I. Derman and Javier Lopez-Garrido (2020). Milestones in *Bacillus subtilis* sporulation research. **Microbial Cell** 8(1): 1-16. doi: 10.15698/mic2021.01.739

REFERENCES

- Cano R, and Borucki M (1995). Revival and identification of bacterial spores in 25- to 40-million-year-old Dominican amber. **Science** 268(5213): 1060–1064. doi: 10.1126/science.7538699
- Crichton M (1990). Jurassic Park, 1st ed. **Alfred A. Knopf**.
- D'Amelio E, Gentile B, Lista F, and D'Amelio R (2015). Historical evolution of human anthrax from occupational disease to potentially global threat as bioweapon. **Environ Int** 85: 133–146. doi: 10.1016/j.envint.2015.09.009
- Ehrenberg CG (1838). Die Infusionsthierchen als vollkommene Organismen. **Verlag Von Leopold Voss, Leipzig**.
- Cohn F (1876). Untersuchungen über Bakterien IV. Beiträge zur Biologie der Bazillen. **Beitrage zur Biol der Pflanz** 249–276.
- Koch R (1876). Untersuchungen über Bakterien V. Die Aetiologie der Milzbrand-Krankheit, begründet auf die Entwicklungsgeschichte des Bacillus Anthracis. **Beitrage zur Biol der Pflanz** 277–310.
- Tyndall J (1877). On heat as a germicide when discontinuously applied. **Proc R Soc** 25: 569–570.
- Knaysi G (1938). Cytology of bacteria. **Bot Rev** 4: 83–112.
- Lewis IM (1941). The cytology of bacteria. **Bact Rev** 5: 181–230.
- Ernst P (1889). Über Kern- und Sporenbildung in Bakterien. **Z Hyg Infect** 5: 428–486.
- Peters WL (1889). Die Organismen des Sauerteigs und ihre Bedeutung für die Brotgährung. **Bot Ztg** 47: 437–439.
- Young E, and Fitz-James P (1959). Chemical and morphological studies of bacterial spore formation i . The formation of spores in *Bacillus cereus*. **J Biophysical Biochem Cytol** 6(3): 467–482. doi: 10.1083/jcb.6.3.467
- Fitz-James P (1960). Participation of the cytoplasmic membrane in the growth and spore formation of *Bacilli*. **J Biophysical Biochem Cytol** 8(9): 507–528. doi: 10.1083/jcb.8.2.507
- Fitz-james PC (1962). Morphology of spore development in *Clostridium pectinovorum*. **J Bacteriol** 84: 104–114. doi: 10.1128/JB.84.1.104-114.1962
- Kawata T, Inoue T, and Takagi A (1963). Electron microscopy of spore formation and germination in *Bacillus subtilis*. **Jpn J Microbiol** 7(1): 23–41. doi: 10.1111/j.1348-0421.1963.tb00926.x
- Ohye DF, and Murrell DP (1962). Formation and structure of the spore of *Bacillus coagulans*. **J Cell Biol** 14: 111–123. doi: 10.1083/jcb.14.1.111
- Ryter A (1965). Étude morphologique de la sporulation de *Bacillus subtilis*. **Ann Inst Pasteur** 108: 40–60. PMID: 14289982
- Ryter A, Schaeffer P, and Ionesco H (1966). Classification cytologique, par leur stade de blocage, des mutants de sporulation de *Bacillus subtilis* Marburg. **Ann Inst Pasteur** 110(3): 305–315. PMID: 4955547
- Piggot PJ, and Cooté JG (1976). Genetic aspects of bacterial endospore formation. **Bacteriol Rev** 40(4): 908–62. 12736. doi: 10.1128/membr.40.4.908-962.1976
- McKenney PT, Driks A, and Eichenberger P (2013). The *Bacillus subtilis* endospore: assembly and functions of the multilayered coat. **Nat Rev Microbiol** 11(1): 33–44. doi: 10.1038/nrmicro2921
- Spizizen J (1958). Transformation of biochemically deficient strains of *Bacillus subtilis* by deoxyribonucleate. **Proc Natl Acad Sci** 44(10): 1072–1078. doi: 10.1073/pnas.44.10.1072
- Takahashi I (1961). Genetic transduction in *Bacillus subtilis*. **Biochem Biophys Res Commun** 5(3): 171–175. doi: 10.1016/0006-291x(61)90104-8
- Takahashi I (1963). Transducing phages for *Bacillus subtilis*. **J Gen Microbiol** 31: 211–217. doi: 10.1099/00221287-31-2-211
- Thorne CB (1962). Transduction in *Bacillus subtilis*. **J Bacteriol** 83(1): 106–111. doi: 10.1128/jb.83.1.106-111.1962

25. Taylor MJ and Thorne CB (1963). Transduction of *Bacillus licheniformis* and *Bacillus subtilis* by each of two phages. **J Bacteriol** 86:452-461. doi: 10.1128/JB.86.3.452-461.1963
26. Chamberland C, and Roux E (1883). Sur l'atténuation de la virulence de la bactérie charbonneuse, sous l'influence des substances antiseptiques. **Comptes Rendus l'Académie des Sci** 96: 1088–1092.
27. Schaeffer P, and Ionesco H (1960). Contribution à l'étude génétique de la sporogenèse bactérienne. **Comptes rendus Hebdomadaires des Séances l'Académie des Sci** 251: 3125–3127.
28. Takahashi I (1965). Transduction of sporogenesis in *Bacillus subtilis*. **J Bacteriol**. 89(2): 294–298. doi: 10.1128/jb.89.2.294-298.1965
29. Donovan W, Zheng L, Sandman K, and Losick R (1987). Genes encoding spore coat polypeptides from *Bacillus subtilis*. **J Mol Biol** 196(1): 1–10. doi: 10.1016/0022-2836(87)90506-7
30. Hullo M-F, Moszer I, Danchin A, and Martin-verstraete I (2001). CotA of *Bacillus subtilis* is a copper-dependent laccase. **J Bacteriol** 183(18): 5426–5430. doi: 10.1128/JB.183.18.5426
31. Schaeffer P, Ionesco H, and Jacob F (1959). Sur le déterminisme génétique de la sporulation bactérienne. **Comptes rendus Hebdomadaires des Séances l'Académie des Sci** 249: 577–578.
32. Michel J, Cami B, and Schaeffer P (1970). Genetics of sporulation in *Bacillus subtilis* Marburg. **Simp Bact Spores** 13–24.
33. Piggot PJ (1973). Mapping of asporogenous mutations of *Bacillus subtilis*: a minimum estimate of the number of sporulation operons. **J Bacteriol** 114(3): 1241–1253. 4197271. doi: 10.1128/jb.114.3.1241-1253.1973
34. Rogolsky M (1969). Chromosomal regions which control sporulation in *Bacillus subtilis*. **Can J Microbiol** 15: 787–790. doi: 10.1139/m69-137
35. Trowsdale J, and Smith DA (1975). Isolation, characterization, and mapping of *Bacillus subtilis* 168 germination mutants. **J Bacteriol** 123(1): 83–95. 806583. doi: 10.1128/JB.123.1.83-95.1975
36. Vary JC (1975). Properties of temperature-sensitive germination mutants of *Bacillus megaterium*. **J Bacteriol** 121(1): 197–203. doi: 10.1128/JB.121.1.197-203.1975
37. Moir A, Lafferty E, and Smith DA (1979). Genetic analysis of spore germination mutants of *Bacillus subtilis* 168: the correlation of phenotype with map location. **J Gen Microbiol** 111(1): 165–180. doi: 10.1099/00221287-111-1-165
38. Moir A, and Smith DA (1990). The genetics of bacterial spore germination. **Annu Rev Microbiol** 44: 531–53. doi: 10.1146/annurev.mi.44.100190.002531
39. Wax R, Freese E, and Cashel M (1967). Separation of two functional roles of L-alanine in the initiation of *Bacillus subtilis* spore germination. **J Bacteriol** 94(3): 522–529. doi: 10.1128/JB.94.3.522-529.1967
40. Foerster HF (1971). Gamma-aminobutyric acid as a required germinant for mutant spores of *Bacillus megaterium*. **J Bacteriol** 108(2): 817–823. doi: 10.1128/jb.108.2.817-823.1971
41. Sammons RL, Slynn GM, and Smith DA (1987). Genetical and molecular studies on *gerM*, a new developmental locus of *Bacillus subtilis*. **J Gen Microbiol** 133: 3299–3312. doi: 10.1099/00221287-133-12-3299
42. Rodrigues CDA, Ramírez-Guadiana FH, Meeske AJ, Wang X, and Rudner DZ (2016). GerM is required to assemble the basal platform of the SpoIIA–SpoIIQ transenvelope complex during sporulation in *Bacillus subtilis*. **Mol Microbiol** 102(2): 260–273. doi: 10.1111/mmi.13457
43. Curiel-quesada E, Setlow B, and Setlow P (1983). Cloning of the gene for C protein, a low molecular weight spore-specific protein from *Bacillus megaterium*. **Proc Natl Acad Sci U S A** 80: 3250–3254. doi: 10.1073/pnas.80.11.3250
44. Connors MJ, Mason JM, and Setlow P (1986). Cloning and nucleotide sequencing of genes for three small, acid-soluble proteins from *Bacillus subtilis* spores. **J Bacteriol** 166(2): 417–425. doi: 10.1128/jb.166.2.417-425.1986
45. Setlow P (1988). Small, acid-soluble spore proteins of *Bacillus* species: structure, synthesis, genetics, function, and degradation. **Annu Rev Microbiol** 42: 319–338. doi: 10.1146/annurev.mi.42.100188.001535
46. Zheng LB, Donovan WP, Fitz-James PC, and Losick R (1988). Gene encoding a morphogenic protein required in the assembly of the outer coat of the *Bacillus subtilis* endospore. **Genes Dev** 2(8): 1047–54. doi: 10.1101/gad.2.8.1047
47. Connors MJ, and Setlow P (1985). Cloning of a small, acid-soluble spore protein gene from *Bacillus subtilis* and determination of its complete nucleotide sequence. **J Bacteriol** 161(1): 333–339. doi: 10.1128/jb.161.1.333-339.1985
48. Mason JM, and Setlow P (1986). Essential role of small, acid-soluble spore proteins in resistance of *Bacillus subtilis* spores to UV light. **J Bacteriol** 167(1): 174–8. doi: 10.1128/jb.167.1.174-178.1986
49. Hackett RH, and Setlow P (1988). Properties of spores of *Bacillus subtilis* strains which lack the major small, acid-soluble protein. **J Bacteriol** 170(3): 1403–1404. doi: 10.1128/jb.170.3.1403-1404.1988
50. Sandman K, Losick R, and Youngman P (1987). Genetic analysis of *Bacillus subtilis* spo mutations generated by Tn917-mediated insertional mutagenesis. **Genetics** 117(4): 603–617. doi: 10.1128/JB.01343-06
51. Youngman PJ, Perkins JB, and Losick R (1983). Genetic transposition and insertional mutagenesis in *Bacillus subtilis* with *Streptococcus faecalis* transposon Tn917. **Proc Natl Acad Sci U S A** 80(8): 2305–9. doi: 10.1073/pnas.80.8.2305
52. Youngman P (1990). Use of transposons and integrational vectors for mutagenesis and construction of gene fusions in *Bacillus* species. In: Harwood CR, Cutting SM, editors *Mol. Biol. Methods Bacillus*, First. **John Wiley & Sons, Chichester, New York, Brisbane, Toronto, Singapore**; pp 221–266.
53. Zeigler DR (2002). Integration Vectors for gram-positive bacteria. **Bacillus Genet Stock Cent Cat** 4: 1–58.
54. Bron S (1990). Plasmids. In: Harwood CR, Cutting SM, editors *Mol. Biol. Methods Bacillus*, First. **John Wiley & Sons, Chichester, New York, Brisbane, Toronto, Singapore**; pp 75–174.
55. Errington J (1990). Gene cloning techniques. In: Harwood CR, Cutting SM, editors *Mol. Biol. Methods Bacillus*, First. **John Wiley & Sons, Chichester, New York, Brisbane, Toronto, Singapore**; pp 175–220.
56. Errington J, and Mandelstam J (1986). Use of a *lacZ* gene fusion to determine the dependence pattern and the spore compartment expression of sporulation operon *spoVA* in *spo* mutants of *Bacillus subtilis*. **J Gen Microbiol** 132(1986): 2977–2985. doi: 10.1099/00221287-132-11-2977
57. Panzer S, Losick R, Sun D, and Setlow P (1989). Evidence for an additional temporal class of gene expression in the forespore compartment of sporulating *Bacillus subtilis*. **J Bacteriol** 171(1): 561–564. doi: 10.1128/jb.171.1.561-564.1989
58. Mason JM, Hackett RH, and Setlow P (1988). Regulation of expression of genes coding for small, acid-soluble proteins of *Bacillus subtilis* spores: studies using *lacZ* gene fusions. **J Bacteriol** 170(1):

239–244. doi: 10.1128/jb.170.1.239-244.1988

59. De Lencastre H, and Piggot PJ (1979). Identification of different sites of expression for *spo* loci by transformation of *Bacillus subtilis*. **J Gen Microbiol** 114(2): 377–389. doi: 10.1099/00221287-114-2-377

60. Illing N, Young M, and Errington J (1990). Use of integrational plasmid excision to identify cellular localization of gene expression during sporulation in *Bacillus subtilis*. **J Bacteriol** 172(12): 6937–6941. doi: 10.1128/jb.172.12.6937-6941.1990

61. Burgess RR, Travers AA, Dunn JJ, and Bautz EKF (1969). Factor stimulating transcription by RNA polymerase. **Nature** 221(5175): 43–46. doi: 10.1038/221043a0

62. Travers AA, and Burgess RR (1969). Cyclic Re-use of the RNA polymerase sigma factor. **Nature** 222(5193): 537–540. doi: 10.1038/222537a0

63. Pero J, Tjian R, Nelson J, and Losick R (1975). *In vitro* transcription of a late class of phage SP01 genes. **Nature** 257: 248–251. doi: 10.1038/257248a0

64. Haldenwang WG, and Losick R (1979). A modified RNA polymerase transcribes a cloned gene under sporulation control in *Bacillus subtilis*. **Nature** 282: 256–260. doi: 10.1038/282256a0

65. Losick R, Youngman P, and Piggot PJ (1986). Genetics of endospore formation in *Bacillus subtilis*. **Annu Rev Genet.** 20: 625–669. doi: 10.1146/annurev.genet.20.1.625

66. Losick R, and Youngman PJ (1984). Endospore formation in *Bacillus*. In: Losick R, Shapiro L, editors *Microb. Dev. Cold Spring Harbor Lab, Cold Spring Harbor, NY*, pp 63–88.

67. Haldenwang WG, Lang N, and Losick R (1981). A sporulation-induced sigma-like regulatory protein from *B. subtilis*. **Cell** 23(2): 615–624. doi: 10.1016/0092-8674(81)90157-4

68. Stragier P, Bouvier J, Bonamy C, and Szulmajster J (1984). A developmental gene product of *Bacillus subtilis* homologous to the sigma factor of *Escherichia coli*. **Nature** 312: 376–378. doi: 10.1038/312376a0

69. Trempy JE, Bonamy C, Szulmajster J, and Haldenwang WG (1985). *Bacillus subtilis* σ factor σ^{29} is the product of the sporulation-essential gene *spoIIG*. **Proc Natl Acad Sci U S A** 82(12): 4189–92. doi: 10.1073/pnas.82.12.4189

70. Errington J, Fort P, and Mandelstam J (1985). Duplicated sporulation genes in bacteria. Implications for simple developmental systems. **FEBS Lett** 188(2): 184–188. doi: 10.1016/0014-5793(85)80368-9

71. Stragier P (1986). Comment on “Duplicated sporulation genes in bacteria” by J. Errington, P. Fort and J. Mandelstam (FEBS Letters 188 (1985) 184-188). **FEBS Lett** 195(1–2): 9–11. doi: 10.1016/0014-5793(86)80119-3

72. Sun D, Stragier P, and Setlow P (1989). Identification of a new σ -factor involved in compartmentalized gene expression during sporulation of *Bacillus subtilis*. **Genes Dev** 3: 141–149. doi: 10.1101/gad.3.2.141

73. Karmazyn-Campelli C, Bonamy C, Savelli B, and Stragier P (1989). Tandem genes encoding σ -factors for consecutive steps of development in *Bacillus subtilis*. **Genes Dev** 3(2): 150–157. doi: 10.1101/gad.3.2.150

74. Masuda ES, Anaguchi H, Yamada K, and Kobayashi Y (1988). Two developmental genes encoding σ factor homologs are arranged in tandem in *Bacillus subtilis*. **Proc Natl Acad Sci U S A** 85(20): 7637–41. doi: 10.1073/pnas.85.20.7637

75. Kroos L, Kunkel B, and Losick R (1989). Switch protein alters specificity of RNA polymerase containing a compartment-specific

sigma factor. **Science** 243(4890): 526–529. doi: 10.1126/science.2492118

76. Errington J (2003). Regulation of endospore formation in *Bacillus subtilis*. **Nat Rev Microbiol** 1(2): 117–26. doi: 10.1038/nrmicro750

77. Hilbert DW, and Piggot PJ (2004). Compartmentalization of gene expression during *Bacillus subtilis* spore formation. **Microbiol Mol Biol Rev** 68(2): 234–262. doi: 10.1128/MMBR.68.2.234

78. Tan IS, and Ramamurthi KS (2014). Spore formation in *Bacillus subtilis*. **Environ Microbiol Rep** 6(3): 212–225. doi: 10.1111/1758-2229.12130

79. Higgins D, and Dworkin J (2012). Recent progress in *Bacillus subtilis* sporulation. **FEMS Microbiol Rev** 36(1): 131–48. doi: 10.1111/j.1574-6976.2011.00310.x

80. Eichenberger P, Fujita M, Jensen ST, Conlon EM, Rudner DZ, Wang ST, Ferguson C, Haga K, Sato T, Liu JS, and Losick R (2004). The program of gene transcription for a single differentiating cell type during sporulation in *Bacillus subtilis*. **PLoS Biol** 2(10): e328. doi: 10.1371/journal.pbio.0020328

81. Wang ST, Setlow B, Conlon EM, Lyon JL, Imamura D, Sato T, Setlow P, Losick R, and Eichenberger P (2006). The forespore line of gene expression in *Bacillus subtilis*. **J Mol Biol** 358(1): 16–37. doi: 10.1016/j.jmb.2006.01.059

82. Eichenberger P (2012). Genomics and cellular biology of endospore formation. In: Graumann PL, editor *Bacillus Cell. Mol. Biol., Second. Caister Academic Press, Norfolk, UK*; pp 319–350.

83. Zhu B, and Stülke J (2018). SubtiWiki in 2018: from genes and proteins to functional network annotation of the model organism *Bacillus subtilis*. **Nucleic Acids Res** 46: D743–D748. doi: 10.1093/nar/gkx908

84. Karow LM, Glaser P, and Piggot PJ (1995). Identification of a gene, *spoIIr*, which links the activation of σ^E to the transcriptional activity of σ^E during sporulation in *Bacillus subtilis*. **Proc Natl Acad Sci USA** 92(6): 2012–2016. doi: 10.1073/pnas.92.6.2012

85. Londoño-Vallejo J a, Fréhel C, and Stragier P (1997). SpoIIQ, a forespore-expressed gene required for engulfment in *Bacillus subtilis*. **Mol Microbiol** 24(1): 29–39. doi: 10.1046/j.1365-2958.1997.3181680.x

86. Clarke S, Lopez-Diaz I, and Mandelstam J (1986). Use of *lacZ* gene fusions to determine the dependence pattern of the sporulation gene *spoIID* in *spo* mutants of *Bacillus subtilis*. **J Gen Microbiol** 132(1 986): 2987–2994. doi: 10.1099/00221287-132-11-2987

87. Rong S, Rosenkrantz MS, and Sonenshein AL (1986). Transcriptional control of the *Bacillus subtilis* spoIID gene. **J Bacteriol** 165(3): 771–9. doi: 10.1128/jb.165.3.771-779.1986

88. Roels S, Driks A, and Losick R (1992). Characterization of *spoIIA*, a sporulation gene involved in coat morphogenesis in *Bacillus subtilis*. **J Bacteriol** 174(2): 575–85. doi: 10.1128/jb.174.2.586-594.1992

89. Nicholson WL, Sun DX, Setlow B, and Setlow P (1989). Promoter specificity of sigma G-containing RNA polymerase from sporulating cells of *Bacillus subtilis*: identification of a group of forespore-specific promoters. **J Bacteriol** 171(5): 2708–2718. doi: 10.1128/jb.171.5.2708-2718.1989

90. Cutting S, Panzer S, and Losick R (1989). Regulatory studies on the promoter for a gene governing synthesis and assembly of the spore coat in *Bacillus subtilis*. **J Mol Biol** 207(2): 393–404. doi: 10.1016/0022-2836(89)90262-3

91. Zheng L, and Losick R (1990). Cascade regulation of spore coat gene expression in *Bacillus subtilis*. **J Mol Biol** 212: 645–660. doi: 10.1016/0022-2836(90)90227-d

92. Sun D, Fajardo-Cavazos P, Sussman MD, Tovar-Rojo F, Cabrera-Martinez RM, and Setlow P (1991). Effect of chromosome location of *Bacillus subtilis* forespore genes on their *spo* gene dependence and transcription by Eo^f: identification of features of good Eo^f-dependent promoters. **J Bacteriol** 173(24): 7867–74. doi: 10.1128/jb.173.24.7867-7874.1991
93. Li Z, and Piggot PJ (2001). Development of a two-part transcription probe to determine the completeness of temporal and spatial compartmentalization of gene expression during bacterial development. **Proc Natl Acad Sci U S A** 98(22): 12538–43. doi: 10.1073/pnas.221454798
94. Wu LJ, and Errington J (1998). Use of asymmetric cell division and *spoIIIE* mutants to probe chromosome orientation and organization in *Bacillus subtilis*. **Mol Microbiol** 27(4): 777–786. doi: 10.1046/j.1365-2958.1998.00724.x
95. Sharp MD, and Pogliano K (2002). Role of cell-specific SpoIIIE assembly in polarity of DNA transfer. **Science**. 295(5552): 137–9. doi: 10.1126/science.1066274.
96. Becker EC, and Pogliano K (2007). Cell-specific SpoIIIE assembly and DNA translocation polarity are dictated by chromosome orientation. **Mol Microbiol** 66(5): 1066–79. doi: 10.1111/j.1365-2958.2007.05992.x
97. Ptacin JL, Nollmann M, Becker EC, Cozzarelli NR, Pogliano K, and Bustamante C (2008). Sequence-directed DNA export guides chromosome translocation during sporulation in *Bacillus subtilis*. **Nat Struct Mol Biol** 15(5): 485–93. doi: 10.1038/nsmb.1412
98. Burton BM, Marquis K a, Sullivan NL, Rapoport T a, and Rudner DZ (2007). The ATPase SpoIIIE transports DNA across fused septal membranes during sporulation in *Bacillus subtilis*. **Cell** 131(7): 1301–12. doi: 10.1016/j.cell.2007.11.009
99. Marquis KA, Burton BM, Nollmann M, Ptacin JL, Bustamante C, Ben-Yehuda S, and Rudner DZ (2008). SpoIIIE strips proteins off the DNA during chromosome translocation. **Genes Dev** 22(13): 1786–95. doi: 10.1101/gad.1684008
100. Lopez-Garrido J, Ojic N, Khanna K, Wagner FR, Villa E, Endres RG, and Pogliano K (2018). Chromosome translocation inflates *Bacillus* forespores and impacts cellular morphology. **Cell** 172(4). doi: 10.1016/j.cell.2018.01.027
101. Shin JY, Lopez-Garrido J, Lee S-HH, Diaz-Celis C, Fleming T, Bustamante C, and Pogliano K (2015). Visualization and functional dissection of coaxial paired SpoIIIE channels across the sporulation septum. **Elife** 4: e06474. doi: 10.7554/eLife.06474
102. Blaylock B, Jiang X, Rubio A, Moran CP, and Pogliano K (2004). Zipper-like interaction between proteins in adjacent daughter cells mediates protein localization. **Genes Dev** 18(23): 2916–2928. doi: 10.1101/gad.1252704
103. Doan T, Morlot C, Meisner J, Serrano M, Henriques AO, Moran CP, and Rudner DZ (2009). Novel secretion apparatus maintains spore integrity and developmental gene expression in *Bacillus subtilis*. **PLoS Genet** 5(7): e1000566. doi: 10.1371/journal.pgen.1000566
104. Kellner EM, Decatur A, and Moran CP (1996). Two-stage regulation of an anti-sigma factor determines developmental fate during bacterial endospore formation. **Mol Microbiol** 21(5): 913–924. doi: 10.1046/j.1365-2958.1996.461408.x
105. Sun Y, Sharp MD, and Pogliano K (2000). A dispensable role for forespore-specific gene expression in engulfment of the forespore during sporulation of *Bacillus subtilis*. **J Bacteriol** 182(10): 2919–2927. doi: 10.1128/JB.182.10.2919-2927.2000
106. Camp AH, and Losick R (2008). A novel pathway of intercellular signalling in *Bacillus subtilis* involves a protein with similarity to a component of type III secretion channels. **Mol Microbiol** 69(2): 402–17. doi: 10.1111/j.1365-2958.2008.06289.x
107. Meisner J, Maehigashi T, Andre I, Dunham CM, and Moran CP (2012). Structure of the basal components of a bacterial transporter. **Proc Natl Acad Sci** 109(14): 5446–5451. doi: 10.1073/pnas.1120113109
108. Levdivkov VM, Blagova EV, McFeat A, Fogg MJ, Wilson KS, and Wilkinson AJ (2012). Structure of components of an intercellular channel complex in sporulating *Bacillus subtilis*. **Proc Natl Acad Sci U S A** 109(14): 5441–5. doi: 10.1073/pnas.1120087109
109. Rodrigues CDA, Henry X, Neumann E, Kurauskas V, Bellard L, Fichou Y, Schanda P, Schoehn G, Rudner DZ, and Morlot C (2016). A ring-shaped conduit connects the mother cell and forespore during sporulation in *Bacillus subtilis*. **Proc Natl Acad Sci U S A** 113(41): 11585–11590. doi: 10.1073/pnas.1609604113
110. Zeytuni N, Hong C, Flanagan KA, Worrall LJ, Theiltges KM, Vuckovic M, Huang RK, Massoni SC, Camp AH, Yu Z, and Strynadka NC (2017). Near-atomic resolution cryoEM structures of the 30-fold homooligomeric SpoIIAG channel essential to spore formation in *Bacillus subtilis*. **Proc Natl Acad Sci USA** 114(34): E7073–E7081. doi: 10.1073/pnas.1704310114
111. Meisner J, Wang X, Serrano M, Henriques AO, and Moran CP (2008). A channel connecting the mother cell and forespore during bacterial endospore formation. **Proc Natl Acad Sci U S A** 105(39): 15100–5. doi: 10.1073/pnas.0806301105
112. Camp AH, and Losick R (2009). A feeding tube model for activation of a cell-specific transcription factor during sporulation in *Bacillus subtilis*. **Genes Dev** 23(8): 1014–24. doi: 10.1101/gad.1781709
113. Riley EP, Trinquier A, Reilly ML, Durchon M, Perera VR, Pogliano K, and Lopez-Garrido J (2018). Spatiotemporally regulated proteolysis to dissect the role of vegetative proteins during *Bacillus subtilis* sporulation: cell-specific requirement of σ^B and σ^A . **Mol Microbiol** 108(1): 45–62. doi: 10.1111/mmi.13916
114. Griffith KL, and Grossman AD (2008). Inducible protein degradation in *Bacillus subtilis* using heterologous peptide tags and adaptor proteins to target substrates to the protease ClpXP. **Mol Microbiol** 70(4): 1012–25. doi: 10.1111/j.1365-2958.2008.06467.x
115. Riley EP, Lopez-Garrido J, Sugie J, Liu RB and Pogliano K (2020). Metabolic differentiation and intercellular nurturing underpin bacterial endospore formation. *Science Advances*. *In press*
116. Wu LJ, and Errington J (1994). *Bacillus subtilis* SpoIIIE protein required for DNA segregation during asymmetric cell division. **Science** 264(5158): 572–575. doi: 10.1126/science.8160014
117. Wu LJ, and Errington J (1997). Septal localization of the SpoIIIE chromosome partitioning protein in *Bacillus subtilis*. **EMBO J** 16(8): 2161–9. doi: 10.1093/emboj/16.8.2161
118. Bath J, Wu LJ, Errington J, and Wang JC (2000). Role of *Bacillus subtilis* SpoIIIE in DNA transport across the mother cell-forespore division septum. **Science** 290(5493): 995–7. doi: 10.1126/science.290.5493.995
119. Cattoni DI, Thakur S, Godefroy C, Le Gall A, Lai-Kee-Him J, Milhiet P-E, Bron P, and Nöllmann M (2014). Structure and DNA-binding properties of the *Bacillus subtilis* SpoIIIE DNA translocase revealed by single-molecule and electron microscopies. **Nucleic Acids Res** 42(4): 2624–36. doi: 10.1093/nar/gkt1231
120. Cattoni DI, Chara O, Godefroy C, Margeat E, Trigueros S, Milhiet P-E, and Nöllmann M (2013). SpoIIIE mechanism of directional translocation involves target search coupled to sequence-dependent motor stimulation. **EMBO Rep** 14(5): 473–9. doi: 10.1038/embor.2013.39
121. Liu N, Chistol G, and Bustamante C (2015). Two-subunit DNA

- escort mechanism and inactive subunit bypass in an ultra-fast ring ATPase. *Elife* 4: 1–20. doi: 10.7554/eLife.09224.001
122. Liu N, Chistol G, Cui Y, and Bustamante C (2018). Mechanochemical coupling and bi-phasic force-velocity dependence in the ultra-fast ring ATPase SpoIIIE. *Elife* 7: e32354. doi: 10.7554/eLife.32354
123. Sharp MD, and Pogliano K (1999). An *in vivo* membrane fusion assay implicates SpoIIIE in the final stages of engulfment during *Bacillus subtilis* sporulation. *Proc Natl Acad Sci U S A* 96(25): 14553–8. doi: 10.1073/pnas.96.25.14553
124. Fleming TC, Shin JY, Lee SH, Becker E, Huang KC, Bustamante C, and Pogliano K (2010). Dynamic SpoIIIE assembly mediates septal membrane fission during *Bacillus subtilis* sporulation. *Genes Dev* 24: 1160–1172. doi: 10.1101/gad.1925210
125. Fiche J-B, Cattoni DI, Diekmann N, Langerak JM, Clerte C, Royer C a, Margeat E, Doan T, and Nöllmann M (2013). Recruitment, assembly, and molecular architecture of the SpoIIIE DNA pump revealed by superresolution microscopy. *PLoS Biol* 11(5): e1001557. doi: 10.1371/journal.pbio.1001557
126. Ben-Yehuda S, Rudner DZ, and Losick R (2003). Assembly of the SpoIIIE DNA translocase depends on chromosome trapping in *Bacillus subtilis*. *Curr Biol* 13(24): 2196–2200. doi: 10.1016/j.cub.2003.12.001
127. Besprozvannaya M, Pivorunas VL, Feldman Z, and Burton BM (2013). SpoIIIE protein achieves directional DNA translocation through allosteric regulation of ATPase activity by an accessory domain. *J Biol Chem* 288(40): 28962–28974. doi: 10.1074/jbc.M113.484055
128. Chara O, Borges A, Milhiet PE, Nöllmann M, and Cattoni DI (2018). Sequence-dependent catalytic regulation of the SpoIIIE motor activity ensures directionality of DNA translocation. *Sci Rep*. 8(1): 1–10. doi: 10.1038/s41598-018-23400-8.
129. Barák I, Muchová K, and Labajová N (2019). Asymmetric cell division during *Bacillus subtilis* sporulation. *Future Microbiol* 14(4): 353–363. doi: 10.2217/fmb-2018-0338
130. Errington J, and Wu LJ (2017). Cell cycle machinery in *Bacillus subtilis*. In: Löwe J, Amos LA, editors *Prokaryotic Cytoskeleton*. Springer International Publishing; pp 67–101.
131. Khanna K, Lopez-Garrido J, Sugie J, Pogliano K, and Villa E (2020). Asymmetric localization of the cell division machinery during *Bacillus subtilis* sporulation. *bioRxiv*. doi: 10.1101/2020.07.22.216184
132. Abanes-De Mello A, Sun YL, Aung S, and Pogliano K (2002). A cytoskeleton-like role for the bacterial cell wall during engulfment of the *Bacillus subtilis* forespore. *Genes Dev* 16: 3253–3264. doi: 10.1101/gad.1039902
133. Broder DH, and Pogliano K (2006). Forespore engulfment mediated by a ratchet-like mechanism. *Cell* 126(5): 917–28. doi: 10.1016/j.cell.2006.06.053
134. Morlot C, Uehara T, Marquis K a, Bernhardt TG, and Rudner DZ (2010). A highly coordinated cell wall degradation machine governs spore morphogenesis in *Bacillus subtilis*. *Genes Dev* 24(4): 411–22. doi: 10.1101/gad.1878110
135. Ojkic N, López-Garrido J, Pogliano K, and Endres RG (2016). Cell-wall remodeling drives engulfment during *Bacillus subtilis* sporulation. *Elife* 5: e18657. doi: 10.7554/eLife.18657
136. Chastanet A, and Losick R (2007). Engulfment during sporulation in *Bacillus subtilis* is governed by a multi-protein complex containing tandemly acting autolysins. *Mol Microbiol* 64(1): 139–52. doi: 10.1111/j.1365-2958.2007.05652.x
137. Ojkic N, López-Garrido J, Pogliano K, and Endres RG (2014). Bistable forespore engulfment in *Bacillus subtilis* by a zipper mechanism in absence of the cell wall. *PLoS Comput Biol* 10(10): e1003912. doi: 10.1371/journal.pcbi.1003912
138. Khanna K, Lopez-Garrido J, Zhao Z, Watanabe R, Yuan Y, Sugie J, Pogliano K, and Villa E (2019). The molecular architecture of engulfment during *Bacillus subtilis* sporulation. *Elife* 8: 1–22. doi: 10.7554/eLife.45257
139. Khanna K, Lopez-garrido J, and Pogliano K (2020). Shaping an endospore: architectural transformations during *Bacillus subtilis* sporulation. *Annu Rev Microbiol* 74: 361–386. doi: 10.1146/annurev-micro-022520-074650
140. Popham DL, and Bernhards CB (2015). Spore peptidoglycan. *Microbiol Spectr* 3(6): 1–21. doi: 10.1128/microbiolspec.TBS-0005-2012
141. Driks A, and Eichenberger P (2016). The spore coat. *ASMscience* 4(2): 1–19. doi: 10.1128/microbiolspec.TBS
142. Meeske AJ, Rodrigues CDA, Brady J, Lim HC, Bernhardt TG, and Rudner DZ (2016). High-throughput genetic screens identify a large and diverse collection of new sporulation genes in *Bacillus subtilis*. *PLoS Biol* 14(11): e1002341. doi: 10.1371/journal.pbio.1002341
143. Koo BM, Kritikos G, Farelli JD, Todor H, Tong K, Kimsey H, Wapinski I, Galardini M, Cabal A, Peters JM, Hachmann AB, Rudner DZ, Allen KN, Typas A, and Gross CA (2017). Construction and analysis of two genome-scale deletion libraries for *Bacillus subtilis*. *Cell Syst* 4(3): 291-305.e7. doi: 10.1016/j.cels.2016.12.013
144. Galperin MY (2013). Genome diversity of spore-forming Firmicutes. *Microbiol Spectr* 1(2): TBS-0015-2012. doi: 10.1128/microbiolspec.tbs-0015-2012
145. Al-Hinai MA, Jones SW, and Papoutsakis ET (2015). The *Clostridium* sporulation programs: diversity and preservation of endospore differentiation. *Microbiol Mol Biol Rev* 79(1): 19–37. doi: 10.1128/MMBR.00025-14
146. Fimlaid KA, Bond JP, Schutz KC, Putnam EE, Leung JM, Lawley TD, and Shen A (2013). Global analysis of the sporulation pathway of *Clostridium difficile*. *PLoS Genet* 9(8). doi: 10.1371/journal.pgen.1003660
147. Ribis JW, Fimlaid KA, and Shen A (2018). Differential requirements for conserved peptidoglycan remodeling enzymes during *Clostridioides difficile* spore formation. *Mol Microbiol* (617): 0–2. doi: 10.1111/mmi.14090
148. Ribis JW, Ravichandran P, Putnam EE, Pishdadian K, and Shen A (2017). The conserved spore coat protein SpoVM is largely dispensable in *Clostridium difficile* spore formation. *mSphere* 2(5): e00315-17. doi: 10.1128/mSphere.00315-17
149. Fimlaid KA, and Shen A (2015). Diverse mechanisms regulate sporulation sigma factor activity in the Firmicutes. *Curr Opin Microbiol* 24: 88–95. doi: 10.1016/j.mib.2015.01.006
150. Dembek M, Kelly A, Barwinska-Sendra A, Tarrant E, Stanley WA, Vollmer D, Biboy J, Gray J, Vollmer W, and Salgado PS (2018). Peptidoglycan degradation machinery in *Clostridium difficile* forespore engulfment. *Mol Microbiol* 0–2. doi: 10.1111/mmi.14091
151. Serrano M, Crawshaw AD, Dembek M, Monteiro JM, Pereira FC, Pinho MG, Fairweather NF, Salgado PS, and Henriques AO (2016). The SpoIIQ-SpoIIAH complex of *Clostridium difficile* controls forespore engulfment and late stages of gene expression and spore morphogenesis. *Mol Microbiol* 100(1): 204–228. doi: 10.1111/mmi.13311
152. Smith LDS (1970). *Clostridium oceanicum*, sp. n., a sporeforming anaerobe isolated from marine sediments. *J Bacteriol* 103(3): 811–813. doi: 10.1128/jb.103.3.811-813.1970
153. Chapman GB, Herk AS van, and Eguía JM (1992). The occurrence of disporous *Bacillus thuringiensis* cells. *Antonie Van Leeuwenhoek*

61(4): 265–268. doi: 10.1007/BF00713934

154. Duda VI, Lebedinsky A V., Mushegjan MS, and Mitjushina LL (1987). A new anaerobic bacterium, forming up to five endospores per cell -*Anaerobacter polyendosporus* gen. et spec. nov. **Arch Microbiol** 148(2): 121–127. doi: 10.1007/BF00425359

155. Kunstýř I, Schiel R, Kaup FJ, Uhr G, and Kirchoff H (1988). Giant Gram-negative noncultivable endospore-forming bacteria in rodent intestines. **Naturwissenschaften** 75(10): 525–527. doi: 10.1007/BF00361293

156. Clements KD, Sutton DC, and Choat JH (1989). Occurrence and characteristics of unusual protistan symbionts from surgeonfishes (*Acanthuridae*) of the Great Barrier Reef, Australia. **Mar Biol** 102(3): 403–412. doi: 10.1007/BF00428493

157. Angert ER, Hutchison EA, and Miller DA (2014). Sporulation in bacteria: beyond the standard model. **Bact Spore from Mol to Syst** 87–102. doi: 10.1128/microbiolspec.TBS-0013-2012

158. Angert ER, and Losick RM (1998). Propagation by sporulation in the guinea pig symbiont *Metabacterium polyspora*. **Proc Natl Acad Sci U S A** 95(17): 10218–10223. doi: 10.1073/pnas.95.17.10218

159. Angert ER, Brooks AE, and Pace NR (1996). Phylogenetic analysis of *Metabacterium polyspora*: Clues to the evolutionary origin of daughter cell production in *Epulopiscium* species, the largest bacteria. **J Bacteriol** 178(5): 1451–1456. doi: 10.1128/jb.178.5.1451-1456.1996

160. Angert ER, and Clements KD (2004). Initiation of intracellular offspring in *Epulopiscium*. **Mol Microbiol** 51(3): 827–835. doi: 10.1046/j.1365-2958.2003.03869.x