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A large-scale assessment of two-way SNP interactions in breast cancer susceptibility using 46 450 cases and 42 461 controls from the breast cancer association consortium

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Part of the substantial unexplained familial aggregation of breast cancer may be due to interactions between common variants, but few studies have had adequate statistical power to detect interactions of realistic magnitude. We aimed to assess all two-way interactions in breast cancer susceptibility between 70 917 single nucleotide polymorphisms (SNPs) selected primarily based on prior evidence of a marginal effect. Thirty-eight international studies contributed data for 46 450 breast cancer cases and 42 461 controls of European origin as part of a multi-consortium project (COGS). First, SNPs were preselected based on evidence ($P < 0.01$) of a per-allele main effect, and all two-way combinations of those were evaluated by a per-allele (1 d.f.) test for interaction using logistic regression. Second, all 2.5 billion possible two-SNP combinations were evaluated using Boolean operation-based screening and testing, and SNP pairs with the strongest evidence of interaction ($P < 10^{-4}$) were selected for more careful assessment by logistic regression. Under the first approach, 3277 SNPs were preselected, but an evaluation of all possible two-SNP combinations (1 d.f.) identified no interactions at $P < 10^{-8}$. Results from the second analytic approach were consistent with those from the first ($P > 10^{-10}$). In summary, we observed little evidence of two-way SNP interactions in breast cancer susceptibility, despite the large number of SNPs with potential marginal effects considered and the very large sample size. This finding may have important implications for risk prediction, simplifying the modelling required. Further comprehensive, large-scale genome-wide interaction studies may identify novel interacting loci if the inherent logistic and computational challenges can be overcome.

INTRODUCTION

We recently identified 47 novel breast cancer susceptibility loci in a multi-centre case–control study of single nucleotide polymorphisms (SNPs) with prior evidence of association from a combined analysis of genome-wide association studies (GWAS) (1–3), raising the total number of susceptibility loci for breast cancer to >70 (4–20). Despite this and other successful efforts to identify breast cancer susceptibility variants by association, linkage and sequencing studies, a large portion of the observed familial aggregation of the disease remains unexplained (3,21–25). Part of this unexplained heritability may be due to variants that modify breast cancer risk through interaction effects (26), where (and throughout this paper) an interaction is defined in the statistical sense that the relative risks for the combined genotypes differ from those predicted by multiplying the marginal relative risks for each SNP. Few studies have evaluated genetic interactions with sufficient statistical power to reach meaningful conclusions and no SNP–SNP interactions in association with breast cancer risk have been convincingly replicated.

We aimed to assess, agnostically, two-SNP interactions for association with breast cancer risk between 70 917 common SNPs with potential marginal effects, which were selected for genotyping based primarily on a combined analysis of nine GWAS. Interactions were assessed using cases and controls of white European origin from the Breast Cancer Association Consortium (BCAC).

RESULTS

We first preselected 3277, 2788 and 1342 SNPs for the assessment of interactions in regard to risk for overall, oestrogen receptor (ER)-positive and ER-negative disease, respectively, based on statistical evidence of a marginal per-allele effect ($P < 0.01$). An evaluation of all two-SNP combinations (1 degree-of-freedom, d.f.) revealed no clear evidence of interaction in any of the three analyses (Fig. 1). For risk of breast cancer overall, of the 5.4 million interactions considered, 13 had a P -value of $< 10^{-6}$ (compared with 5.4 predicted), but none had a P -value of $< 10^{-8}$. The 13 interactions at $P < 10^{-6}$ represented eight potential interaction signals after accounting for SNPs in high linkage disequilibrium (LD) ($r^2 > 0.85$, Table 1), and three of these involved the same SNP (rs17117532) along with one of three others in modest LD ($0.58 \leq r^2 \leq 0.77$). The lowest P -value was 3.3×10^{-8} , which corresponded to a Bonferroni-corrected value of 0.16 considering all interactions evaluated. The corrected P -value was 0.058 based on the number of possible interactions between the estimated 1898 effective independent loci represented by the total 3277. In all 13 instances, the interaction effect was in the opposite direction to the main effects for the two SNPs involved, with very little LD between the two potentially interacting SNPs ($r^2 \leq 0.073$).

For risk of ER-positive breast cancer, the strongest evidence of interaction was observed for rs7603983 and rs10490346 ($P = 2.6 \times 10^{-10}$). These two SNPs are 45 kb apart and in modest LD ($r^2 = 0.65$). A re-evaluation of the cluster plot revealed poor cluster separation for rs7603983 and it was noted that data from one study in particular (Oulu Breast Cancer Study, OBCS) were overrepresented among borderline genotype determinations. When the 407 cases and 414 controls from the OBCS

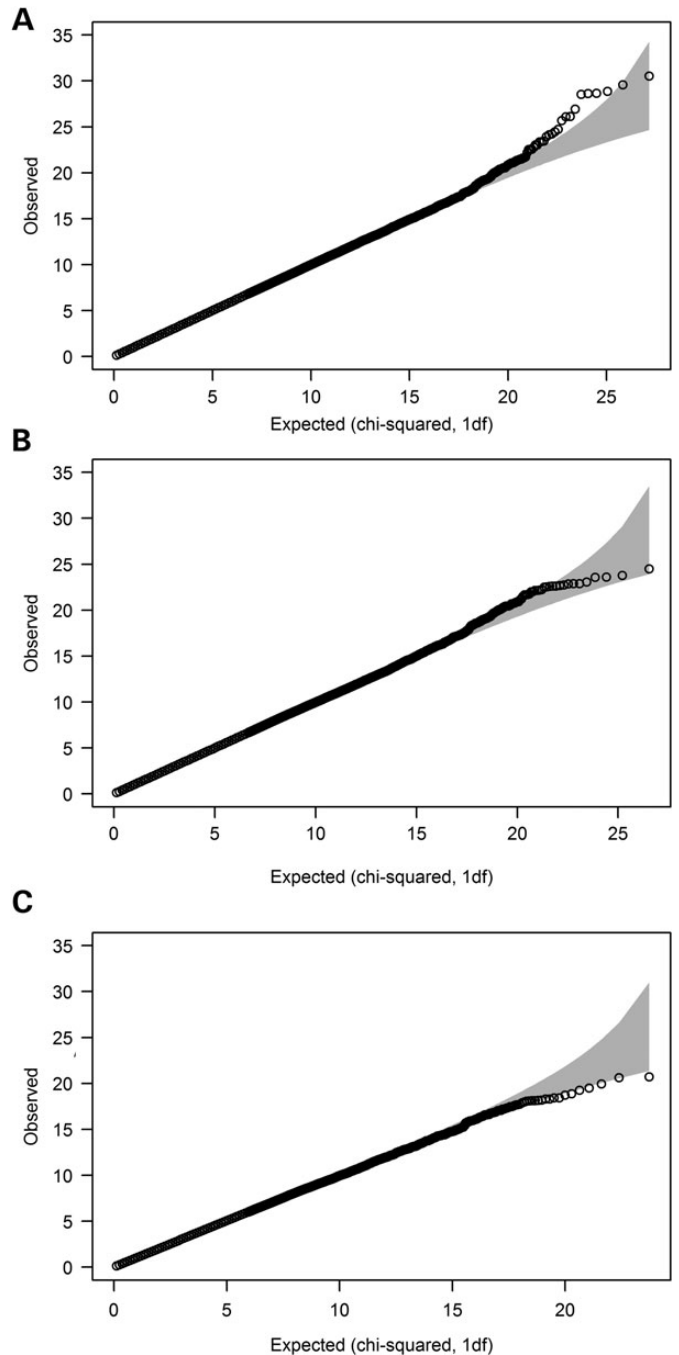


Figure 1. Q–Q plots from the first set of analyses based on the χ^2 statistics from the 1 d.f. LRT for (A) overall breast cancer, (B) oestrogen receptor (ER)-positive breast cancer and (C) ER-negative breast cancer.

(1.1% of the sample) were excluded, there was no evidence of interaction ($P = 0.58$), suggesting that the original result was an artefact of the poor clustering. For none of the other 3.9 million SNP pairs considered for ER-positive disease risk was evidence of interaction observed after correction for multiple testing ($V_{\text{effLi}} = 1647$; $P^* \geq 0.64$) and only one pair had an uncorrected P -value of $< 10^{-6}$ (Table 1). Using this first approach, no evidence of two-SNP interactions was observed in regard to the risk of ER-negative breast cancer ($V_{\text{effLi}} = 949$; $P^* \geq 0.91$).

Table 1. SNP pairs with per-allele P -values for interaction $< 10^{-6}$

	Position	MAF (r^2)	Single-SNP analysis ^a OR (95% CI), P -value	Interaction analysis ^b OR (95% CI), P -value
Overall breast cancer				
rs10822036	Chr10: 64044366	0.18	1.04 (1.01–1.06); 5.2×10^{-3}	0.88 (0.82–0.94)
rs1379805	Chr18: 18748550	0.36	1.04 (1.02–1.06); 1.4×10^{-4}	0.92 (0.87–0.97)
Interaction		(<0.01)		1.10 (1.06–1.14); 7.7×10^{-7}
Overall breast cancer				
rs132274	Chr22: 27855126	0.35	1.03 (1.01–1.05); 9.4×10^{-3}	1.19 (1.12–1.26)
rs17478824	Chr5: 87821380	0.09	1.04 (1.01–1.08); 1.1×10^{-2}	1.28 (1.18–1.41)
Interaction		(<0.01)		0.88 (0.84–0.93); 6.6×10^{-7}
Overall breast cancer				
rs1573998	Chr5: 56312250	0.20	1.06 (1.03–1.08); 2.6×10^{-6}	0.87 (0.81–0.93)
rs16886496 ^c	Chr5: 56253286	0.09	1.13 (1.09–1.17); 3.7×10^{-13}	0.88 (0.80–0.96)
Interaction		(0.073)		1.15 (1.09–1.21); 7.8×10^{-8}
Overall breast cancer				
rs17117532	Chr14: 82392717	0.10	1.05 (1.01–1.08); 5.3×10^{-3}	0.84 (0.77–0.92)
rs1936396 ^d	Chr10: 44868074	0.23	1.03 (1.01–1.06); 4.3×10^{-3}	0.87 (0.81–0.93)
Interaction		(<0.01)		1.16 (1.10–1.23); 5.4×10^{-8}
Overall breast cancer				
rs17117532	Chr14: 82392717	0.10	1.05 (1.01–1.08); 5.3×10^{-3}	0.84 (0.77–0.92)
rs2018728 ^e	Chr10: 44837353	0.34	1.03 (1.01–1.05); 5.6×10^{-3}	0.88 (0.83–0.94)
Interaction		(<0.01)		1.14 (1.09–1.20); 9.2×10^{-8}
Overall breast cancer				
rs17117532	Chr14: 82392717	0.10	1.05 (1.01–1.08); 5.3×10^{-3}	0.84 (0.77–0.91)
rs7905526 ^f	Chr10: 44837353	0.27	1.03 (1.01–1.06); 2.1×10^{-4}	0.87 (0.82–0.93)
Interaction		(<0.01)		1.15 (1.10–1.21); 3.3×10^{-8}
Overall breast cancer				
rs17355209	Chr4: 84672332	0.50	1.04 (1.02–1.06); 3.0×10^{-5}	1.19 (1.13–1.26)
rs4980025	Chr10: 80510787	0.11	0.94 (0.92–0.97); 3.0×10^{-4}	1.17 (1.07–1.29)
Interaction		(<0.01)		0.90 (0.86–0.94); 8.6×10^{-7}
Overall breast cancer				
rs7714708	Chr5: 58330771	0.36	1.03 (1.01–1.05); 4.1×10^{-3}	1.17 (1.11–1.23)
rs836808	Chr5: 80010326	0.26	0.97 (0.95–0.99); 7.5×10^{-3}	1.12 (1.06–1.19)
Interaction		(<0.01)		0.92 (0.89–0.95); 4.0×10^{-7}
ER-positive disease				
rs11604821	Chr11: 69061318	0.33	1.04 (1.02–1.07); 2.8×10^{-4}	1.22 (1.14–1.30)
rs11600497	Chr11: 68882499	0.14	0.95 (0.92–0.98); 3.0×10^{-3}	1.17 (1.07–1.27)
Interaction		(<0.01)		0.88 (0.84–0.93); 7.5×10^{-7}

MAF, minor allele frequency; r , correlation coefficient; OR, odds ratio per copy of the minor allele(s); CI, confidence interval.

^aThe single-SNP analysis modelled main effects, per copy of the minor allele, for each SNP separately (it was on this basis that SNPs were preselected for inclusion in the interaction analysis under the first analytical approach).

^bThe interaction analysis included main effects for each of the two SNPs plus an interaction effect, in all cases per copy of each minor allele.

^cSNP in LD with two SNPs, rs12655019 ($r^2 = 0.94$) and rs16886525 ($r^2 = 0.99$), for which $P = 2.1 \times 10^{-7}$ and 8.8×10^{-8} , respectively, for interaction with rs1573998. r^2 was 0.58 between rs1936396 and rs7905526, 0.77 between rs1936396 and rs7905526 and 0.74 between rs2018728 and rs7905526.

^dSNP in LD with another SNP, rs11471 ($r^2 = 0.94$), for which $P = 3.3 \times 10^{-7}$ for interaction with rs17117532. r^2 was 0.58 between rs1936396 and rs7905526, 0.77 between rs1936396 and rs7905526 and 0.74 between rs2018728 and rs7905526.

^eSNP in LD with another SNP, rs2018728 ($r^2 = 0.86$), for which $P = 9.0 \times 10^{-7}$ for interaction with rs17117532. r^2 was 0.58 between rs1936396 and rs7905526, 0.77 between rs1936396 and rs7905526 and 0.74 between rs2018728 and rs7905526.

^fSNP in LD with another SNP, rs6593456 ($r^2 = 0.98$), for which $P = 8.7 \times 10^{-8}$ for interaction with rs17117532. r^2 was 0.58 between rs1936396 and rs7905526, 0.77 between rs1936396 and rs7905526 and 0.74 between rs2018728 and rs7905526.

We evaluated the statistical power of our first approach to detect interactions between the preselected SNPs at a nominal significance level of 10^{-8} , in the absence of main effects (Table 2). We estimated that for overall breast cancer, we had >90% power to detect a per-allele interaction odds ratio (OR_{int}) as low as 1.16 between SNPs with minor allele frequency (MAF) >0.20. The corresponding minimum detectable OR_{int} were 1.29 and 1.58 for SNPs with a MAF as low as 0.10 and 0.05, respectively. For pairs of SNPs with a equal MAF of 0.20, 0.10 and 0.05, an OR_{int} of 1.16, 1.29 and 1.58, respectively, in the absence of main effects gives rise to a marginal OR of 1.08, 1.06 and 1.07, respectively, when the interactions are not accounted for. The corresponding statistical power estimates

for the detection of these marginal OR in single-SNP analysis at $P < 0.01$ (i.e. to preselect SNPs for inclusion in the assessment of interactions) were 99, 88 and 72%. The power was similar for ER-positive disease, particularly for the more common SNPs (MAF ≥ 0.10). The power was lower for ER-negative disease, although for SNPs with MAF >0.20, the power was >90% for OR_{int} as low as 1.30.

As an alternative approach, we exhaustively investigated all 2.5 billion possible two-SNP interactions using Boolean operation-based screening and testing (BOOST) and identified 278 387, 278 240 and 275 214 SNP pairs potentially associated ($P < 10^{-4}$) with risk of overall, ER-positive and ER-negative disease, respectively. These interactions were then evaluated

Table 2. Minimum interaction odds ratio detectable with 90% power at $P < 10^{-8}$ in the absence of main effects

MAF _{SNP2}	MAF _{SNP1} 0.05	0.10	0.20	0.40
Overall breast cancer				
0.05	1.58 (1.06, 56%)	1.41 (1.04, 47%)	1.30 (1.03, 48%)	1.24 (1.03, 69%)
0.10	1.41 (1.08, 85%)	1.29 (1.06, 88%)	1.21 (1.04, 78%)	1.17 (1.04, 93%)
0.20	1.30 (1.12, 99%)	1.21 (1.08, 99%)	1.16 (1.07, 99%)	1.13 (1.05, 99%)
0.40	1.24 (1.20, 99%)	1.17 (1.14, 99%)	1.13 (1.11, 99%)	1.11 (1.09, 99%)
ER-positive breast cancer				
0.05	1.69 (1.07, 55%)	1.48 (1.05, 53%)	1.35 (1.04, 60%)	1.28 (1.03, 51%)
0.10	1.48 (1.10, 89%)	1.34 (1.07, 87%)	1.25 (1.05, 83%)	1.20 (1.04, 81%)
0.20	1.35 (1.14, 99%)	1.25 (1.10, 99%)	1.19 (1.08, 99%)	1.15 (1.06, 99%)
0.40	1.28 (1.23, 99%)	1.20 (1.17, 99%)	1.15 (1.12, 99%)	1.12 (1.10, 99%)
ER-negative breast cancer				
0.05	2.17 (1.11, 52%)	1.80 (1.08, 52%)	1.57 (1.06, 53%)	1.45 (1.05, 54%)
0.10	1.80 (1.16, 88%)	1.55 (1.11, 84%)	1.40 (1.08, 82%)	1.32 (1.07, 88%)
0.20	1.57 (1.23, 99%)	1.40 (1.16, 99%)	1.30 (1.12, 99%)	1.24 (1.10, 99%)
0.40	1.45 (1.38, 99%)	1.32 (1.27, 99%)	1.24 (1.20, 99%)	1.20 (1.17, 99%)

MAF, minor allele frequency.

In parenthesis are the resulting marginal OR for SNP1 and power to detect it in the individual main-effect analysis (the latter is the probability of selecting the SNP for inclusion under the first approach at $P < 0.01$).

using logistic regression. For overall breast cancer risk, 18 SNP pairs had associated P -values of $< 10^{-10}$ from the genotype-based test for interaction (Supplementary Material, Table S1). Six of these included the SNP rs9625520, which on individual inspection of the cluster plot was determined to have failed genotyping due to merged clusters, despite having passed the previous quality-control filters based on the automatic genotype calls. All seven SNPs forming these six SNP pairs were in high LD ($0.89 \leq r^2 \leq 1.00$) and all but rs9625520 were well-clustered; we re-checked the evidence of interaction between these and observed very little (all $P[4 \text{ d.f.}] \geq 0.018$) for any of 15 other possible two-SNP combinations. A further 11 pairs in the top 18 included the SNP rs7603983 (identified using the first analytical approach) and another correlated SNP within 220 kb ($r^2 \geq 0.50$); the evidence of interaction disappeared for each of these when data from the OBCS were excluded ($P \geq 0.82$). For one of these SNP pairs, evidence of interaction was also observed in the per-allele model ($P = 8.3 \times 10^{-11}$), but not once data from the OBCS were excluded ($P = 0.71$). The remaining SNP-pair comprised two SNPs in high LD ($r^2 = 0.94$); visual inspection of cluster plots showed that one of these (rs6989466) was poorly called (merged clusters). Since the only SNP in high LD with rs6989466 was the potentially interacting one (rs2013845), the interaction could not be assessed by proxy. No other SNP pairs had interaction-associated P -values of $< 10^{-10}$ from the per-allele test for interaction (1 d.f.).

No evidence of interaction in susceptibility to ER-positive or ER-negative breast cancer was observed for any additional SNP pairs using this second approach. The seven SNP pairs with an interaction P -value of $< 10^{-10}$ for ER-positive breast cancer risk were a subset of the 18 observed for overall breast cancer risk, all involved either rs7603983 (for which evidence disappeared after exclusion of data from the OBCS), rs9625520 or rs6989466 (both of which were deemed to have been poorly called after visual inspection of cluster plots) (Supplementary Material, Table S1).

DISCUSSION

We have assessed two-way combinations of $> 70\,000$ SNPs selected because they were associated with, or potentially associated with, breast cancer risk and found little evidence of interaction, defined as departure from multiplicativity between SNP main effects. This was the case for overall breast cancer and disease subtypes defined by ER status. We considered genotype-specific interactions (4 d.f.) for all 2.5 billion SNP pairs and per-allele departure (1 d.f.) for all pairwise combinations of SNPs selected based on evidence of a marginal effect.

Few SNP-SNP interactions have been identified (27) and none associated with breast cancer risk has been convincingly replicated. That said, no large-scale systematic evaluations have been published to date. Tao *et al.* (28) used BOOST to analyse data from two GWAS of prostate cancer, totalling 3140 cases and 4273 controls, and found no convincing evidence of interactions. To our knowledge, our study of 46 450 breast cancer cases and 42 461 controls is by far the largest of its kind. The statistical power of our first analysis of selected SNPs was very high ($> 90\%$) to detect OR_{int} as low as 1.29 between common SNPs ($MAF \geq 0.10$). Also, assuming an epistasis model in which an associated risk is restricted to individuals carrying variant alleles at both loci, which would imply a weak marginal effect for each SNP, the probability that SNPs involved in interactions of this magnitude were selected was also high ($> 80\%$). It was also highly likely that more common SNPs ($MAF \geq 0.20$), involved in interactions with OR_{int} as low as 1.16, were selected. Although no SNP pair achieved $P < 10^{-8}$, we did observe an excess of pairs with an interaction $P < 10^{-6}$ (13 versus 5.4 predicted). This suggests that some of these associations could be real and may be confirmed by even larger studies, if they could be carried out. In addition, it should be noted that the current analysis was based on a set of 70 917 SNPs and was, therefore, not genome-wide. On the other hand, 70% of the SNPs were included because evidence of per-allele association at $P < 0.008$ was

observed in the combined GWAS (Stage 1) analysis of $>10\,000$ cases and $10\,000$ controls; based on the scenarios considered in Table 2, the statistical power of this Stage 1 analysis was as low as 40% (although in most cases much higher) for more common SNPs. It should also be noted that the effect sizes for untyped causal variants may be greater than those associated with the tagging SNPs (tagSNPs) on genotyping arrays, and that the relative reduction in power to detect disease association based on tag-SNPs may be greater for interaction effects than for main effects. We cannot therefore rule out that more common SNPs involved in interactions, with OR_{int} comparable to those reported above, were not selected for the iCOGS array and thus not included in our analysis. Nevertheless, our findings, based on a very large number of SNPs, are consistent with the assertion of Hill (2010) that interactions are likely to be small and very difficult to detect if the main effects are already small.

Although our study possessed lower statistical power to detect two-way interactions between less common SNPs ($MAF < 0.10$), the study sample size required to detect any such interactions may not be achievable. Zuk *et al.* (2012) concur with this, suggesting that detecting genetic interactions may require a sample size in the range of 500 000. It is also possible that interactions could exist in the absence of marginal main effects, if each SNP had associated effects that were in opposite directions, depending on the genotype of the other SNP. Such SNPs would not have been systematically selected for the iCOGS array nor preselected for the assessment of interactions using our first analytical approach. However, while mathematically possible, such qualitative interactions seem less biologically plausible.

Had we adopted a more stringent P -value criterion (than $P < 0.01$) to select SNPs based on evidence of a marginal effect using the first approach, one result in particular would have stood out. This involved two SNPs (rs1573998 and rs16886496) in low LD ($r^2 = 0.073$), both with marginal associations significant at $P < 10^{-5}$ (Table 1), located within 60 kb of each other, at an established breast cancer susceptibility region on 5q11.2 (5). Such *cis*-interactions have a somewhat different interpretation to those between SNPs located in different regions, in that they may reflect haplotype-specific risks or, equivalently, the effects of untyped rarer variants. The iCOGS array included several hundred additional variants across this region in order to inform the search for the likely causal variant(s); these fine-mapping SNPs were not included in the present analysis since they are being analysed as part of a separate, ongoing project, but it may be informative to perform a separate analysis of all possible *cis*-interactions in the future.

The results from our exhaustive assessment of all two-way interactions using BOOST were consistent with the largely null result from our first set of analyses, although the power of this second analysis is difficult to ascertain. BOOST is generally more powerful than other comparable methods, except in the presence of per-allele departure from independent SNP effects (29). The 18 SNP pairs for which strongest evidence of interaction ($P < 10^{-10}$) was observed all included one of three SNPs with poor or questionable genotype clustering and a second SNP relatively close to (≤ 178 kb) and in LD with ($r^2 \geq 0.50$) the first. All 18 were ruled out as possible interactions, either because the evidence disappeared in a sensitivity analysis excluding a very small proportion of the data from one study with particularly poor genotype clustering or

because the SNP was considered to have failed genotyping on visual inspection of the intensity cluster plots. Where possible, interactions were reassessed with other SNPs in LD with the failed SNP and no consistent results were observed. This highlights the need to check manually the cluster plots of SNPs potentially involved in interactions, since poor clustering is not always picked up in standard quality-control checks for high-throughput genotyping data.

It is possible that other types of two-way SNP–SNP interaction (not tested for in the present study) exist, as may higher order interactions. These might be discovered by other analytical approaches. Our results, particularly those from the 4 d.f. genotype-based test, suggest that the genotyping quality of SNPs involved in potential interactions should be checked to rule out artefacts resulting from poor clustering. Even so, our results are consistent with those of other large studies that have assessed two-way interactions between established susceptibility SNPs and other risk factors for breast cancer (30–33). Together, these results suggest that established risk factors for breast cancer tend to be related to disease risk such that their associated effects, expressed as ORs, can be multiplied together. This has important implications for risk prediction, simplifying the modelling required. Related to this, Aschard *et al.* (27) have shown by simulation that even if gene–gene and gene–environment interactions exist in regard to breast cancer risk, they are unlikely to improve dramatically the discrimination ability of risk-prediction models.

That we found no strong evidence of two-way SNP interactions might be surprising, given the consistent evidence of genetic epistasis in model systems (34). However, the main effects in model systems also tend to be larger than those observed for SNPs. It may be that the influence on disease risk of the biological processes that are modified by SNP interactions is too small to be detected, even using combined studies with large sample size, at least for breast cancer. This may also be related to the much more complex genetic background present in humans, perhaps together with the influence of lifestyle factors that may dilute genetic interactions. More sophisticated analyses could possibly tease out SNP interactions in breast cancer susceptibility, but this will probably require a much better understanding of how to classify SNPs in terms of their functional basis. Two-way SNP interactions have been reported for other diseases such as Psoriasis, Ankylosing Spondylitis and Behcet's disease, suggesting that such interactions can be found for complex phenotypes in humans (35–37).

The key strengths of our study are, in addition to the extremely large sample size (and resulting high statistical power), the very large set of SNPs (potentially associated with breast cancer susceptibility) considered, the uniform genotyping procedures and quality-control measures adopted and the large-scale analyses conducted to evaluate two-way interactions in a comprehensive way. A non-trivial issue for analyses of this kind is the establishment of a statistical significance threshold that adequately controls the proportion of false-positive findings. Since permutation-testing was not feasible, we dealt with the issue of non-independence of the multiple tests for interaction under our first approach by estimating the effective number of independent SNPs and used this to compute an effective number of independent interactions. No SNP pairs were robust to correction for multiple testing on this basis. However, further work is required to determine whether this approach gives a reasonable estimate for the effective

number of interaction tests. The appropriateness of the Bonferroni correction in this context could also be questioned. For the second approach, we applied a statistical significance threshold of 10^{-10} , which is almost an order of magnitude greater than the Bonferroni-corrected value based on the 2.5 billion interactions tested and can, therefore, be considered liberal.

Association analyses, including analyses of interactions, may be subject to confounding due to population structure. We aimed to correct for potential population stratification by adjusting for study and the six leading principal components. Little evidence of inflation of test statistics was observed after these adjustments in the main-effects analysis of the iCOGS data (3), suggesting that correction for population structure was adequate, and that any bias in the interaction tests was likely to be small. In any event, such confounding would be more likely to lead to false-positive associations, which we did not observe.

In conclusion, we observed little evidence of two-way SNP interactions for breast cancer susceptibility, despite the large number of SNPs (with potential marginal effects) considered

and the very large sample size. More comprehensive, large-scale genome-wide interaction studies may identify novel interacting loci if the inherent logistic and computational challenges can be overcome.

METHODS

Study subjects

A total of 38 case-control studies contributed data and DNA samples for 46 450 breast cancer cases and 42 461 controls of white European origin from 13 European countries, Canada, Australia and the USA. Details are provided in Table 3 and Supplementary Material, Table S2. The ER status of the tumour was known for 34 479 cases, 27 074 were ER-positive and 7405 were ER-negative. All study participants gave informed consent and all studies were approved by the corresponding local research ethics committees.

Table 3. Studies contributing white European cases and controls

Study	Country	Controls	Cases	ER+	ER-
Australian Breast Cancer Family Study (ABCFS) ^a	Australia	551	790	456	261
Amsterdam Breast Cancer Study (ABCS)	Netherlands	1429	1325	420	153
Bavarian Breast Cancer Cases and Controls (BBCC)	Germany	458	564	460	83
British Breast Cancer Study (BBCS)	UK	1397	1554	507	114
Breast Cancer In Galway Genetic Study (BIGGS)	Ireland	719	836	495	154
Breast Cancer Study of the University Clinic Heidelberg (BSUCH)	Germany	954	852	499	154
CECILE Breast Cancer Study (CECILE)	France	999	1019	797	144
Copenhagen General Population Study (CGPS)	Denmark	4086	2901	1919	357
Spanish National Cancer Centre Breast Cancer Study (CNIO-BCS)	Spain	876	902	242	88
California Teachers Study (CTS)	USA	71	68	0	17
ESTHER Breast Cancer Study (ESTHER)	Germany	502	478	304	98
Gene Environment Interaction and Breast Cancer in Germany (GENICA)	Germany	427	465	328	119
Helsinki Breast Cancer Study (HEBCS)	Finland	1234	1664	1295	237
Hannover-Minsk Breast Cancer Study (HMBCS)	Belarus	130	690	37	0
Karolinska Breast Cancer Study (KARBAC)	Sweden	662	722	338	63
Kuopio Breast Cancer Project (KBCP)	Finland	251	445	304	97
kConFab/Australian Ovarian Cancer Study (kConFab/AOCS)	Australia	897	613	162	59
Leuven Multidisciplinary Breast Centre (LMBC)	Belgium	1388	2671	2071	379
Mammary Carcinoma Risk Factor Investigation (MARIE)	Germany	1778	1818	1349	399
Milan Breast Cancer Study Group (MBCSG)	Italy	400	488	149	42
Mayo Clinic Breast Cancer Study (MCBCS)	USA	1931	1862	1486	295
Melbourne Collaborative Cohort Study (MCCS)	Australia	511	614	352	119
Multi-ethnic Cohort (MEC)	USA	741	731	415	87
Montreal Gene-Environment Breast Cancer Study (MTLGEBCS)	Canada	436	489	421	64
Norwegian Breast Cancer Study (NBCS)	Norway	70	22	0	22
Oulu Breast Cancer Study (OBCS)	Finland	414	507	407	100
Ontario Familial Breast Cancer Registry (OFBCR) ^b	Canada	511	1175	630	268
Leiden University Medical Center Breast Cancer Study (ORIGO)	Netherlands	327	357	211	70
NCI Polish Breast Cancer Study (PBCS)	Poland	424	519	519	0
Karolinska Mammography Project for Risk Prediction of Breast Cancer (pKARMA)	Sweden	5537	5434	3672	702
Rotterdam Breast Cancer Study (RBCS)	Netherlands	699	664	368	131
Singapore and Sweden Breast Cancer Study (SASBAC)	Sweden	1378	1163	663	144
Sheffield Breast Cancer Study (SBCS)	UK	848	843	377	105
Study of Epidemiology and Risk factors in Cancer Heredity (SEARCH)	UK	8069	9347	5160	1181
Städtisches Klinikum Karlsruhe Deutsches Krebsforschungszentrum Study (SKKDKFZS)	Germany	29	136	0	136
IHCC-Szczecin Breast Cancer Study (SZBCS)	Poland	315	365	165	60
Triple Negative Breast Cancer Consortium Study (TNBCC)	Various	542	881	0	881
UK Breakthrough Generations Study (UKBGS)	UK	470	476	96	22
Total		42 461	46 450	27 074	7405

ER+, oestrogen-receptor-positive cases; ER-, oestrogen-receptor-negative cases.

^aAustralian site of the Breast Cancer Family Registry.

^bOntario site of the Breast Cancer Family Registry.

SNP selection and genotyping

SNP selection and genotyping methods have been described previously (3). The 75 380 SNPs eligible for inclusion in the present study were selected primarily based on statistical evidence of association with breast cancer risk from a combined analysis of nine GWAS involving 10 052 cases and 12 575 controls (46% SNPs) or from analyses of disease-subtype- or population-specific GWAS (24% SNPs). Additional SNPs were selected based on other evidence of association with breast (2% SNPs) or other cancers (4% SNPs), with overall survival following breast cancer (9% SNPs) or with risk factors for breast or other hormone-related cancers (12% SNPs) or because they tagged genes in the DNA repair pathway (2% SNPs).

Genotyping was carried out at four centres using a custom-built Illumina iSelect array (iCOGS) as part of a multi-consortium project (COGS), as described previously (3). Genotypes were called using Illumina's proprietary GenCall algorithm. Two percent of samples were provided in duplicate by all studies, and 270 HapMap2 samples were genotyped at all four centres. Subjects with an overall call-rate <95% were excluded. We excluded SNPs with call-rates <95% and those for which statistical evidence of deviation from Hardy–Weinberg equilibrium was observed for controls at a significance threshold of 10^{-7} , based on a stratified 1 d.f. test in which the deviations were summed across study-based strata (38). We also excluded SNPs for which genotypes were discrepant for more than 2% of duplicate samples. A total of 72 611 of the eligible SNPs were

successfully genotyped according to these criteria; 70 917 of those had a MAF of at least 0.01 in controls and were considered for the present analysis. The cluster plots of SNPs for which evidence of interaction was observed were individually re-evaluated and, where appropriate, manually recalled.

Statistical methods

Evidence of interaction between SNP pairs in susceptibility to breast cancer was assessed using logistic regression, primarily based on a 1 d.f. likelihood ratio test (LRT) comparing two models: one model included main effects for study (categorical variable) and seven principal components (each continuous), a per-allele main effect for each of the two SNPs under consideration and an interaction parameter for the product of the latter two; the other model included main-effect parameters only. Thus, interaction was primarily considered to be per-allele departure from multiplicativity between ORs corresponding to SNP main effects. This was done for breast cancer risk overall, for risk of ER-positive breast cancer and for ER-negative breast cancer. Based on a series of pilot analyses, we estimated that, even with access to super-computing facilities with 1000 processors, an exhaustive analysis of all 2.5 billion ($\times 10^9$) two-way SNP interactions, while desirable (39), would take more than one year to complete. This was due to the number of SNP pairs to be tested, the large sample size and the inclusion

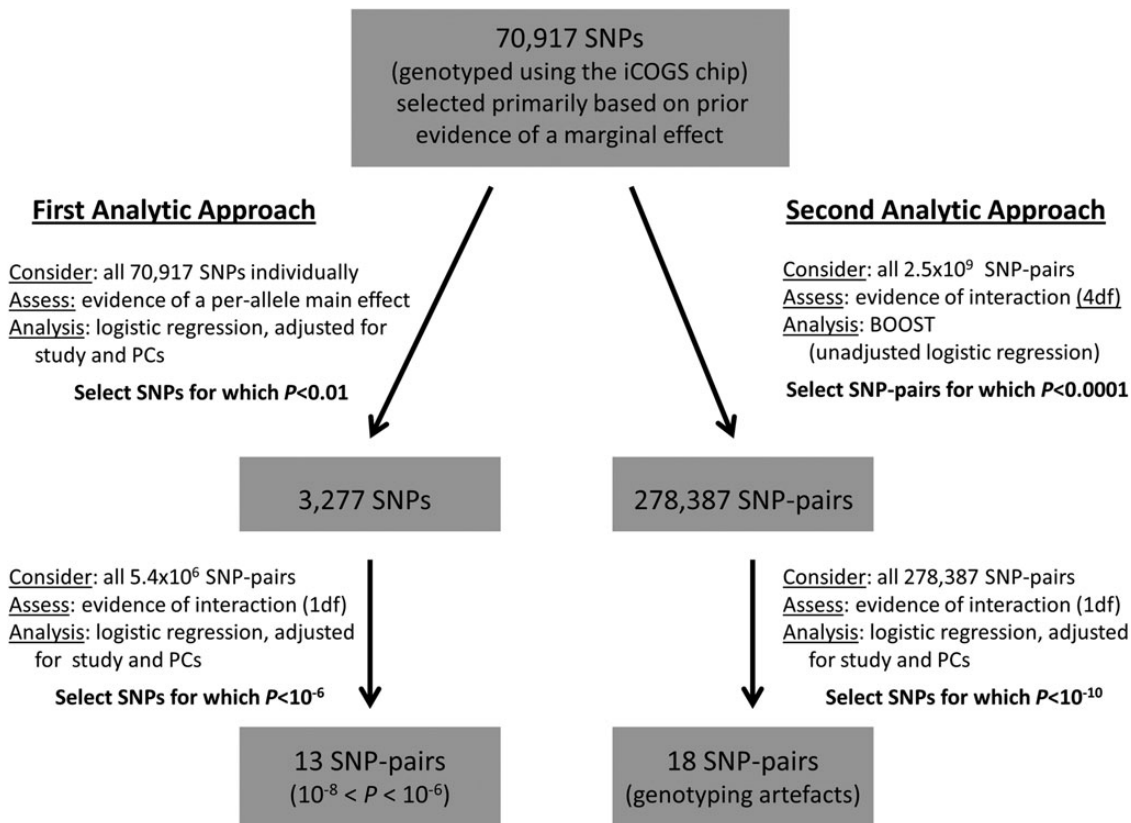


Figure 2. Schematic representation of the two strategies applied to assess pairwise interactions in susceptibility to overall breast cancer risk between the 70 917 SNPs considered.

of 37 dummy variables in the adjustment for study, the latter two factors increasing substantially the run-time per interaction.

We used two strategies to reduce the number of SNP pairs considered (Fig. 2). Under the first, we selected individual SNPs based on evidence of a marginal per-allele effect at $P < 0.01$. For each outcome (overall, ER-positive and ER-negative), SNPs were selected and then all possible two-way combinations of this reduced set of SNPs were considered in testing for evidence of interaction for that outcome. This strategy was founded on the idea that even a purely epistatic interaction (i.e. where the risk associated with one SNP is only manifest in individuals carrying a particular allele of another SNP) will give rise to apparent marginal SNP effects when the interaction is not modelled (40). Since the screening test is independent of the subsequent interaction test, substantial gain in power is obtained after correction for multiple (interaction) testing (41). Since permutation testing was not possible, given the sample size and number of interactions considered, we estimated instead the effective number of independent SNPs (V_{effLi}) using the method described by Li and Ji (42). This method was applied via the web-interface matSpDlite (<http://gump.qimr.edu.au/general/daleN/matSpDlite/>, last accessed on 20 November 2013), based on the observed correlations between SNPs, which was input as a matrix of correlation coefficients (43). The number of two-way interactions between V_{effLi} SNPs (T) was then used as a basis on which to calculate Bonferroni-corrected P -values (P^*).

Under the second strategy, we applied BOOST (29) to screen all possible SNP pairs and select a reduced set to test formally using the LRT defined above. BOOST runs the equivalent of a simplified logistic-regression-based LRT in a highly efficient way, thereby permitting all 2.5 billion possible interactions to be screened in a reasonable time-frame. However, it is based on a co-dominant, genotype-based interaction model (4 d.f.) and does not allow adjustment for covariates. We used this strategy to select SNP pairs with the strongest evidence of interaction ($P < 10^{-4}$) for more careful assessment using the less analytically efficient, but more adequate, adjusted logistic-regression models. The study- and principal-component-adjusted 4 d.f. interaction LRT test was then applied to the selected SNP pairs, as was the 1 d.f. test described above. Since BOOST requires complete data for all variants for all individuals, missing genotypes were imputed as the most common of the three genotypes for each SNP, across all subjects with available data. Imputed genotypes were not used in the final logistic-regression analysis.

Statistical power calculations were performed based on a range of MAF and OR using Quanto (<http://hydra.usc.edu/gxe/>, last accessed on 20 November 2013), and all other statistical analyses were carried out using R version 2.13.2. Q–Q plots were drawn for the first set of analyses based on the χ^2 statistics from the 1 d.f. LRT using the *qq.chisq* function (44).

SUPPLEMENTARY MATERIAL

Supplementary Material is available at *HMG* online.

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