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## ORIGINAL ARTICLE

# *Ralstonia solanacearum* lipopeptide induces chlamydospore development in fungi and facilitates bacterial entry into fungal tissues

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***Ralstonia solanacearum* is a globally distributed soil-borne plant pathogenic bacterium, which shares a broad ecological range with many plant- and soil-associated fungi. We sought to determine if *R. solanacearum* chemical communication directs symbiotic development of polymicrobial consortia. *R. solanacearum* produced a diffusible metabolite that induced conserved morphological differentiation in 34 species of fungi across three diverse taxa (Ascomycetes, Basidiomycetes and Zygomycetes). Fungi exposed to this metabolite formed chlamydospores, survival structures with thickened cell walls. Some chlamydospores internally harbored *R. solanacearum*, indicating a newly described endofungal lifestyle for this important plant pathogen. Using imaging mass spectrometry and peptidogenomics, we identified an undescribed lipopeptide, ralsolamycin, produced by an *R. solanacearum* non-ribosomal peptide synthetase-polyketide synthase hybrid. Inactivation of the hybrid non-ribosomal peptide synthetase-polyketide synthase gene, *rmyA*, abolished ralsolamycin synthesis. *R. solanacearum* mutants lacking ralsolamycin no longer induced chlamydospore development in fungal coculture and invaded fungal hyphae less well than wild-type. We propose that ralsolamycin contributes to the invasion of fungal hyphae and that the formation of chlamydospores may provide not only a specific niche for bacterial colonization but also enhanced survival for the partnering fungus.**

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## Introduction

*Ralstonia solanacearum* is the causative agent of lethal bacterial wilt and is a globally distributed soil-borne plant pathogen known to infect well over 200 different plant hosts (Hayward, 1991). The broad distribution of this bacterium in a wide variety of soil environments suggests that *R. solanacearum* likely encounters a diversity of other soil microbes, including fungi. Although the molecular interactions between *R. solanacearum* and host plants have been well studied (Genin and Denny, 2012), there has been little exploration of *R. solanacearum*'s interactions with soil cohabitants.

Soil is a heterogeneous, complex microcosm replete with inter-organismal interactions, which restructure the physical and chemical composition of the shared environment. Small molecule signals are increasingly recognized as important communication signals in microbial interactions (Tarkka *et al.*, 2009). Historically, many of these specialized metabolites (Davies, 2013) have been harnessed for pharmaceutical purposes, and there has been a resurgence of interest in natural products based on microbial crosstalk (Cragg and Newman, 2013). Genome mining of *R. solanacearum* strain GMI1000 shows that it has the potential to produce a variety of secondary metabolites. A few of these metabolites have been characterized including ralfuranone (Wackler *et al.*, 2011) and the yersinabactin-like siderophore, micacocidin (Kreutzer *et al.*, 2011). Micacocidin has moderate antimycoplasmal and antifungal effects *in vitro* because of its metal chelating properties (Kobayashi *et al.*, 2000). These and other compounds produced

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by *R. solanacearum* likely impact how it interacts with microbes in the soil.

Small molecule-mediated antibiosis has been documented in both bacterial and fungal systems (Haas and Défago, 2005; Burlinson *et al.*, 2011), but antagonistic interactions are only one potential interaction outcome. Recent advances in the understanding of bacterial–fungal interactions indicate that bacterial natural products can stimulate morphological transitions in many fungi. In particular, bacterial metabolites commonly induce fungal sporulation. For example, *Paenibacillus validus* produces trisaccharides that induce hyphal elongation and sporulation in the arbuscular mycorrhizal fungus *Glomus intraradices* (Hildebrandt *et al.*, 2006). Similarly, the fungal plant pathogen *Rhizopus microsporus* sporulates only when it is infected with its endosymbiotic bacterial partner, *Burkholderia rhizoxinica* (Lackner *et al.*, 2011a).

Other research has shown that bacterial volatile organic compounds and diffusible small molecules can induce developmental shifts in fungi aside from sporulation. For example, reciprocal volatile signaling between *R. solanacearum* and the soil fungus *Aspergillus flavus* represses *A. flavus* sporulation and inhibits production of *R. solanacearum* virulence factors (Spraker *et al.*, 2014). Fengycin A, a lipopeptide produced by *Bacillus subtilis*, causes diverse fungal taxa to form chlamydospores, thick-walled spores associated with fungal survival in harsh environmental conditions (Li *et al.*, 2012). In addition, phenazine redox metabolites produced by *Pseudomonas aeruginosa* trigger *Aspergillus* spp. to sporulate along a diffusion gradient (Zheng *et al.*, 2015). These developmental shifts may contribute to microbial niche securement in the soil by triggering hard-wired defense or survival mechanisms, however, these ecological mechanisms are largely unexplored.

Here we characterize a small molecule-mediated interaction between the common soil-borne bacterial pathogen *R. solanacearum* and a taxonomic variety of plant- and soil-associated fungi that results in fungal chlamydospore development. Using matrix-assisted laser desorption/ionization (MALDI) imaging mass spectrometry (MALDI-IMS) and tandem mass spectrometry (MS/MS) technologies, we identified a novel diffusible *R. solanacearum* lipopeptide, here named ralsolamycin. We observed a developmental shift in all fungi ranging from chlamydospore formation to growth inhibition along an increasing gradient of ralsolamycin. Inactivating *rmyA*, a gene that encodes a hybrid non-ribosomal peptide synthetase-polyketide synthase abolished ralsolamycin production. Further, the *rmyA* mutant did not induce chlamydospore formation. Remarkably, we also demonstrate that *R. solanacearum* invades the chlamydospores, a previously undescribed niche that may contribute to the noted environmental persistence of this devastating plant pathogen.

## Materials and methods

### *Media and growth conditions and coculture experiments*

All strains used in this study are listed in Supplementary Table S1. *R. solanacearum* strains were routinely grown at 30 °C on casamino acids-peptone-glucose (CPG) agar (Hendrick and Sequeira, 1984), and mucoid colonies were selected for subsequent experiments. Liquid bacterial cultures were grown overnight in CPG at 30 °C and 180 r.p.m. When appropriate, the antibiotic gentamycin (15 mg l<sup>-1</sup>) was added to the media. Overnight liquid cultures of *R. solanacearum* were pelleted by centrifugation, washed twice in equal volumes of sterilized, double-distilled water, quantified using OD<sub>600</sub> values and adjusted to ~2 × 10<sup>8</sup> cells ml<sup>-1</sup>. Then 5 µl (1 × 10<sup>6</sup> cells) of bacterial suspension was spotted at each point.

*Aspergillus* spp. were routinely grown at 30 °C on glucose minimal medium (Shimizu and Keller, 2001), and conidia were harvested by applying a 0.01% Tween 80 solution to the plate followed by agitation with a plastic cell spreader. Conidial suspensions were quantified using a hemocytometer and diluted in sterile water to 2 × 10<sup>5</sup> spores ml<sup>-1</sup>. Then 5 µl containing 1 × 10<sup>3</sup> spores was applied to a spot 2 cm away from the *R. solanacearum* spot on CPG plates. The droplets were allowed to dry in a NUAIRE biological safety cabinet (NU425-400), then plates were wrapped with parafilm once and incubated at room temperature. Owing to variable sporulation habits, all other fungi were routinely grown on potato dextrose agar and allowed to grow at room temperature. For assays, mycelial plugs were removed from actively growing cultures using the wide end of a 1 ml pipette tip and transferred to the assay plate.

For coculture experiments, *R. solanacearum* and fungi were plated as described above and incubated for 14 days at room temperature. Chlamydospore formation for all fungi was assessed as the fungal colonies made contact with the *R. solanacearum* cultures. As the colonies expanded and grew into one another, sections of the interaction zone were excised for microscopic evaluation. For IMS, experimentation colonies were grown for 3 days at room temperature before being excised and processed. For assessment of endofungal bacteria, green fluorescent protein (GFP) strains of *R. solanacearum* were similarly cocultured with *A. flavus*, but the colonies were inoculated in direct contact to allow for physical interactions.

### *Microscopy of chlamydospores*

Fungal mycelia were harvested from coculture plates by cutting 5 mm × 5 mm sections from the interaction zone between the two cultures and examined for chlamydospore formation. Cell walls were stained with calcofluor white at a final concentration of 1 mg ml<sup>-1</sup> for 1 min before microscopy. DAPI (4,6-diamidino-2-phenylindole) staining of nuclei was

carried out by submerging these sections in 1 mg ml<sup>-1</sup> DAPI solution for 30 min followed by a wash in 0.1 M phosphate-buffered saline pH 7.0. Lipid bodies were stained by submerging sections in 10 µg ml<sup>-1</sup> Nile Red, followed by a wash with 0.1 M phosphate-buffered saline. Stained samples were wet mounted and imaged on a Zeiss Axio Imager A10 microscope (Carl Zeiss, Oberkochen, Germany) equipped with a Zeiss EC Plan-NEOFLUAR 40×/1.3 Oil DIC/∞/0.17 and Zeiss EC Plan-NEOFLUAR 100×/1.3 Oil DIC/∞/0.17 objective and a series 120 X-Cite light source (EXFO). When different filters could be used, samples were stained with multiple stains to provide comparable images for the different features. Images were collected with AxioVision Release 4.7 software (Carl Zeiss).

For counts of chlamydospores of all fungi, interaction zones from coculture with GMI1000 and the *rmyA* mutant were collected and stained with calcofluor as described above. Three separate images of hyphae collected from these interaction zone were captured and chlamydospores per high-powered field were enumerated manually using the cell-counter plug-in of the Fiji software package (Schindelin *et al.*, 2012).

Time-lapse microscopy experiments were carried out on a Nikon Eclipse Ti inverted fluorescent microscope (Nikon Instruments, Melville, NY, USA) with a PlanFluor 10× objective attached to a Nikon DS-Qi1Mc CCD camera (Nikon Instruments). Four filters for four different excitation/emission wavelengths were available: ex-350 nm/em-400 nm; ex-492 nm/em-517 nm; ex-572 nm/em-600 nm; ex-647 nm/em-700 nm. Images were collected with NIS-Elements AR version 4.30.01 software (Nikon Instruments).

Confocal laser-scanning microscopy experiments were prepared as above but performed on a Zeiss 510 Meta microscope (Carl Zeiss). To verify endohyphal localization of bacteria, 0.5 µm Z-sections were taken across multiple planes of view. An argon ion laser was used to excite GFP-labeled cells and linear unmixing calculations were carried out in the Zeiss LSM software to reduce fungal autofluorescence. A helium laser was used to excite the FM4-64-labeled membranes. All confocal images were processed in the open source Fiji software package and all overlays of multiple fluorescence channels are of single confocal Z-planes.

#### Plasmid construction and genetic manipulation

The primers and plasmids used in this work are listed in Supplementary Table S2. PCR amplification was carried out on a C1000 Thermal Cycler from Bio-Rad (Hercules, CA, USA). For creation of the *rmyA*-targeted disruption cassette, flanking regions of the targeted promoter and fatty acyl-AMP ligase (FAAL) domain were amplified from GMI1000 genomic DNA using primers RSp0641\_5'F and RSp0641\_5'R for the upstream sequence and RSp0641\_3'F and RSp0641\_3'R for the downstream sequence, each

amplifying ~1-kb fragments. Both RSp0641\_5'R and RSp0641\_3'F were designed with overlapping sequences to the 0.8-kb gentamycin resistance cassette, *Gmr*, which was amplified from the plasmid pUCgm using the primer set, GentR\_F and GentR\_R. These PCR products were purified with a QIAquick gel extraction kit (Qiagen, Valencia, CA, USA) and quantified before being fused using standard double joint PCR protocols (Yu *et al.*, 2004). The fusion product was blunt-end ligated into the Zero Blunt PCR cloning kit (Invitrogen, Carlsbad, CA, USA) to create pJES12.3. *R. solanacearum* strain GMI1000 was transformed with pJES12.3 by electroporation and plated on gentamycin media to select for appropriate transformants. Colony PCR using RSp0641\_5'F and RSp0641\_3'R were used to screen for loss of the targeted *rmyA* promoter and FAAL domain. All PCR steps were carried out with Pfu Ultra II DNA Polymerases (Agilent, Santa Clara, CA, USA).

The GMI1000 strain constitutively expressing GFP was made by using natural transformation to move the Tn5::gfp (Liu *et al.*, 2001) insertion from genomic DNA of strain K60GFP (Yao and Allen, 2006) into the GMI1000 chromosome and transformants were selected on tetracycline plates at a concentration of 15 mg ml<sup>-1</sup> and confirmed with fluorescence microscopy. Subsequently, the  $\Delta rmyA$  strain constitutively expressing GFP was made by using natural transformation to move the Tn5::gfp insertion from the GMI1000-GFP strain. These strains were selected for tetracycline and gentamycin resistance, confirmed with fluorescence microscopy, and verified phenotypically to not induce chlamydospores.

#### Imaging mass spectrometry

Liquid bacterial cultures were grown overnight in CPG at 30 °C and 180 r.p.m. Overnight liquid cultures of *R. solanacearum* were pelleted by centrifugation, washed 2× in equal volumes of sterilized, double-distilled water, quantified using OD<sub>600</sub> values and adjusted to 2 × 10<sup>8</sup> cells ml<sup>-1</sup>. Then 5 µl (~1 × 10<sup>6</sup> cells) of this cell suspension was spotted at each point. GMI1000 or K60 (5 µl) were spotted next to *A. flavus* (1 µl) on yeast extract/malt extract ISP2 agar (10 ml) in 100 × 25 mm Petri dishes (Yang *et al.*, 2012). All organisms were inoculated on separate Petri dishes as controls. Samples were incubated at room temperature for 72 h. The region of interest and controls were excised from the agar and transferred to the same MALDI MSP 96 anchor plate (Bruker Daltonics, Billerica, MA, USA). A photograph was taken and the aerial hyphae of *A. flavus* were removed gently with a cotton swab dampened with acetonitrile (Moree *et al.*, 2012). Following the removal of the aerial hyphae, another photograph was taken and Universal MALDI matrix (Sigma-Aldrich, St Louis, MO, USA) was applied using a 53 µm stainless steel sieve (Hogentogler & Co., INC., Columbia, MD, USA). The plate was then transferred to an oven and desiccated at 37 °C overnight.

Following adherence to the MALDI plate, another photographic image was taken and the sample was subjected to MALDI-TOF mass spectrometry (Microflex from Bruker Daltonics) for IMS acquisition. Data were acquired in positive reflectron mode, with a 500  $\mu\text{m}$  laser interval in the XY and a mass range of 250–2500 Da. The resulting data were analyzed using FlexImaging software v. 3.0 (Bruker Daltonik GmbH, Bremen, Germany).

#### Extraction and semi-purification

*R. solanacearum* 24-h liquid cultures were filter sterilized using a 0.2  $\mu\text{m}$  pore-size filter and applied to actively growing hyphae of *A. flavus* to determine if direct bacterial contact was necessary for the formation of the chlamyospore-like structures. Within <12 h, a large number of mature hyphal cells differentiated into chlamyospores, indicating that direct physical contact between the microbes was unnecessary for chlamyospore development suggesting diffusible metabolite was responsible for chlamyospore development.

*R. solanacearum* GMI1000, K60 and *A. flavus* and their interaction zones were excised from thin agar (1  $\times$  1 cm) and transferred to separate glass scintillation vials. Acetonitrile (1 ml) was added to the samples and sonicated for 10 min. These samples were partitioned against both 50:50 ACN:H<sub>2</sub>O and butanol and the resulting mixture was centrifuged at 10 000 r.p.m. for 1 min. The soluble organic fraction was removed and dried *in vacuo*. Samples were then re-suspended in 100  $\mu\text{l}$  of MeOH for further analysis.

Bulk extraction of the metabolite was done from liquid culture by growing GMI1000 for 4 days in 1 liter of CPG with 100 g Amberlite XAD-16 resin (Sigma-Aldrich). The resin was separated from the culture broth via vacuum filtration and rinsed with 500 ml double-distilled H<sub>2</sub>O to remove excess salts and cellular debris. The adsorbed compounds were eluted in 1:1 MeOH/DCM and organic fractions were dried *in vacuo*. Subsequently, these fractions were subjected to solid phase extraction using a Waters Sep-Pak C<sub>18</sub> cartridge (5 g) eluting with a 20% step gradient of MeOH/H<sub>2</sub>O, ranging from 20 to 100% MeOH.

#### MS/MS data acquisition

Samples were directly infused into the mass spectrometer using a Triversa nanomate-electrospray ionization source (Advion Biosystems, Ithaca, NY, USA) coupled to a 6.42 T Thermo LTQ-FT-ICR mass spectrometer (Thermo-Electron Corporation, San Jose, CA, USA). For nanomate, samples were diluted in 50:50 MeOH:H<sub>2</sub>O with 0.1% formic acid and then directly infused using a back pressure of 0.35–0.5 psi and a spray voltage of 1.3–1.45 kV. FT-MS and ion trap MS/MS spectra were acquired using Tune Plus software version 1.0 and Xcalibur software version 1.4 SR1 (Thermo-Electron Corporation). The instrument was tuned on *m/z* 816, the 15+ charge state of cytochrome C.

The instrument scan cycle consisted of one 10-min segment, during which a profile FT scan with a resolution of 25 000 was cycled with four data-dependent scans in the ion trap. MS/MS data were collected in a data-dependent manner, during which profile mode FT-MS scans (150–1600 *m/z*) cycled with four MS/MS scans in the ion trap. The four most abundant peaks were fragmented (2 *m/z* isolation width, a normalized collision energy of 35%, and an activation time of 30 ms) and then added to an exclusion list for 600 s.

#### Network analyses

The resulting spectra were converted to mzXML format using msconvert in the ProteoWizard software package (<http://proteowizard.sourceforge.net/tools.shtml>). The data were then subjected to molecular networking (Watrous *et al.*, 2012). The data were then clustered with MS-Cluster with a parent mass tolerance of 1.0 Da and a MS/MS fragment ion tolerance of 0.3 Da to create consensus spectra. Further, consensus spectra that contained <1 spectrum were discarded. A network was then created where edges were filtered to have a cosine score above 0.65 and >6 matched peaks. Further edges between two nodes were kept in the network if and only if each of the nodes appeared in each other's respective top 10 most similar nodes.

#### Phylogenetic analysis of bacteria

Bacterial 16S DNA sequences were obtained from GenBank and aligned using Muscle in the MEGA6 software package (Tamura *et al.*, 2013) using default parameters. The evolutionary history was inferred using the maximum parsimony method. Tree 1 out of 2 most parsimonious trees (length = 1648) is shown. The consistency index is (0.480386), the retention index is (0.730577) and the composite index is 0.372381 (0.350959) for all sites and parsimony-informative sites (in parentheses). The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches (Felsenstein, 1985). The maximum parsimony tree was obtained using the Subtree-Pruning-Regrafting algorithm (Nei and Kumar, 2000) with search level 1 in which the initial trees were obtained by the random addition of sequences (10 replicates). The analysis involved 29 nucleotide sequences. All positions containing gaps and missing data were eliminated. There were a total of 1211 positions in the final data set.

## Results

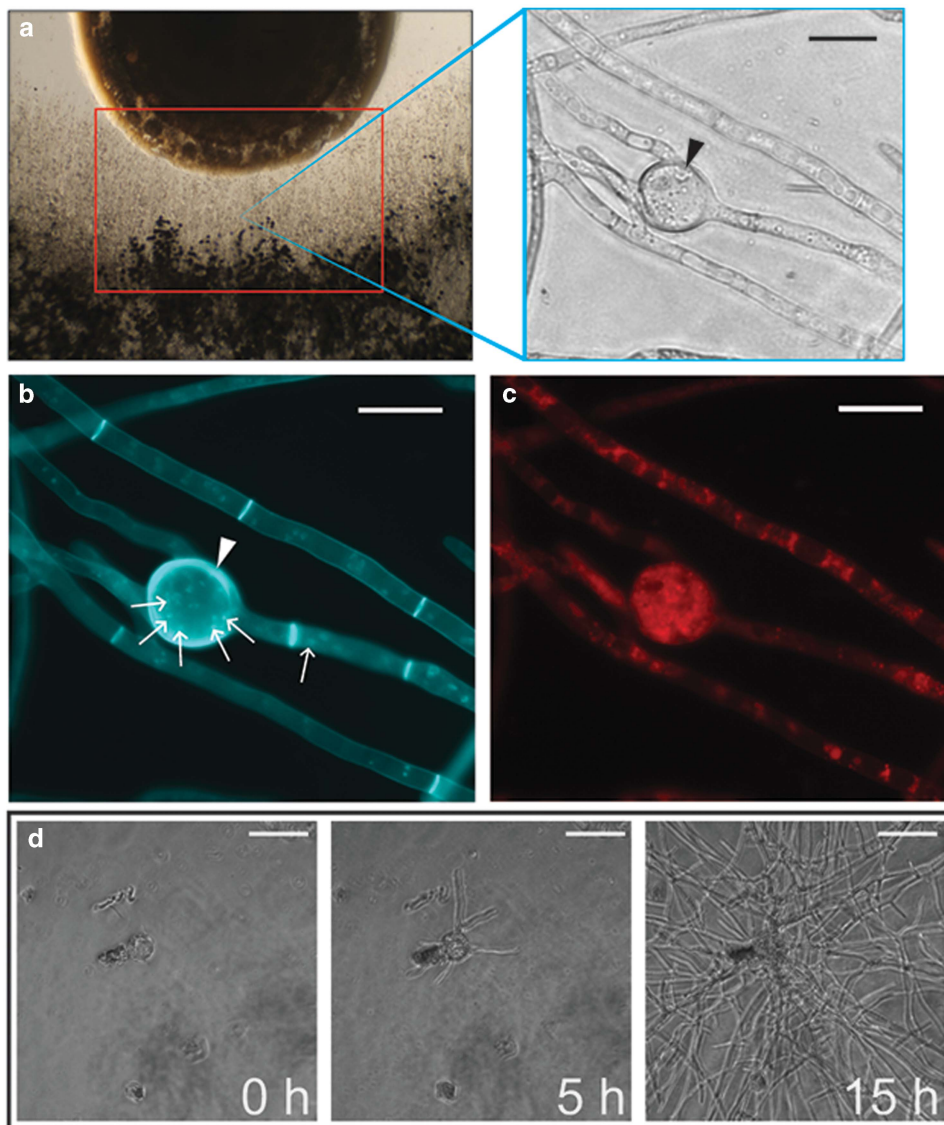
### *A diffusible compound produced by R. solanacearum induces chlamyospore development across diverse fungal taxa*

*R. solanacearum* inhabits a variety of soils and infects over 200 plant hosts. Soils and plants are both

associated with many different fungal species, so we hypothesized that *R. solanacearum* utilizes chemical signals to mediate interactions with a diversity of fungi. Using the characterized and sequenced *R. solanacearum* strain GMI1000, we examined the interaction of cocultures of the bacterium and 34 species of fungi: 30 Ascomycota species, 1 Basidiomycota species (*Rhizoctonia solani*) and 3 Zygomycota species (*Phycomyces blakesleeanus*, *Mucor bacilliformis* and *M. hiemalis*). We found that a distinct zone of fungal inhibition formed between 2 and 14 days of coculture. In addition, all fungi

formed distinct hyphal swellings near (~1.5 cm) the *R. solanacearum* colony, referred to hereafter as the 'interaction zone' (Figure 1a).

The hyphal swellings resembled chlamydospores, thick-walled spores associated with certain fungal genera and thought to aid in fungal survival under harsh environments (Couteaudier and Alabouvette, 1990b; Li *et al.*, 2012). The rates of chlamydospore formation varied considerably between species, but all of the fungi tested formed chlamydospores within 14 days of coculture with *R. solanacearum*. Several fungi only formed chlamydospores after being in



**Figure 1** Examination of the interaction zone of the *R. solanacearum*/fungal coculture revealed fungal hypertrophy resembling chlamydospores produced by other fungi (a) *left*—coculture set-up of bacteria (top) and fungi (bottom). Red box indicates the 'interaction zone' where many intercalated chlamydospore-like structures (*right*—black arrows) developed in response to coculture with *R. solanacearum* strain GMI1000. Control images of non-chlamydospore-inducing bacterial cocultures can be seen in Supplementary Figure S1. Histological analysis of chlamydospores shows that they are (b) polynucleate (small white arrows), have thickened cell walls (white arrowheads) and (c) accumulate lipid bodies. (d) Time-lapse microscopy of isolated chlamydospores placed in fresh media shows these structures are independently viable, and they can germinate to form a new fungal colony. Complete time-lapse video can be viewed in Supplementary Video S1.

direct contact with the *R. solanacearum* colony for multiple days: the necrotrophic plant pathogens *Botrytis cinerea* and *Sclerotinia sclerotiorum*, the coprophilous fungus *Sordaria fimicola*, one *Fusarium* species (*verticillioides*) and the Basidiomycete *Rhizoctonia solani*.

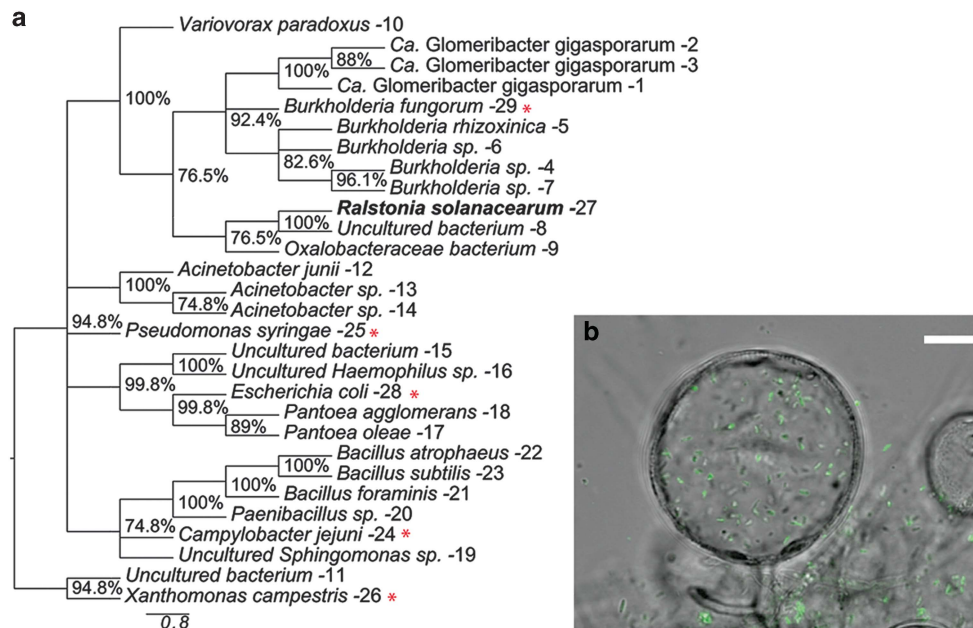
To study the hypertrophic structures in detail, the interaction zones of *A. flavus* were excised and characterized. We chose *A. flavus* because volatile-mediated interactions between *A. flavus* and *R. solanacearum* do not result in chlamydospore formation (Spraker et al., 2014) and also because the genus is generally not known to produce chlamydospores. Micromanipulation and fluorescent histological microscopy confirmed that the hyphal swellings were thick walled, chlamydospores. Hoescht staining showed that most chlamydospores harbored multiple nuclei (Figure 1b) although some lacked nuclei. A fungal cell wall-specific stain, calcofluor white, demonstrated that chlamydospore cell walls were thicker than normal hyphae cell walls (Figure 1b), and Nile Red staining showed chlamydospores accumulated lipid bodies (Figure 1c). Together, the histological evidence suggested that *A. flavus* chlamydospores could be viable structures. To test this hypothesis, chlamydospores were isolated via micromanipulation, washed and placed in fresh media and monitored for germination. After 5 h, the chlamydospores germinated with multiple germ tubes (Figure 1d, Supplementary Video S1).

#### Endofungal lifestyle of *R. solanacearum*

Many bacteria live as endofungal symbionts in a variety of fungal hosts. *Ralstonia* spp. are nested within a larger clade of identified endofungal bacteria (Hoffman and Arnold, 2010; Figure 2a, Supplementary Table S3). In order to test the hypothesis that *R. solanacearum* was capable of invading fungal hyphae, a constitutive GFP-producing *R. solanacearum* strain was cocultured with *A. flavus* for 72 h. After harvesting the resulting mycelial mat and thoroughly washing the hyphae to remove extracellular bacteria, samples were imaged with a fluorescent microscope. Many bacterial cells remained associated with the fungal hyphae, and bacterial aggregates were often associated with chlamydospores, many appearing to be within the chlamydospores. Confocal laser-scanning microscopy confirmed that a substantial number of bacteria were localized to the intracellular space of some chlamydospores (Figure 2b, Supplementary Video S2). Bacteria were rarely found within the non-differentiated hyphae. These data supported our hypothesis that *R. solanacearum* can invade fungal hyphae with specificity to the chlamydospores.

#### Identification of the chlamydospore-inducing compound

To identify *Ralstonia*-specific metabolites that induced chlamydospore formation, we cocultured *A. flavus* with a phylogenetically diverse set of



**Figure 2** *R. solanacearum* is nested among other known endofungal bacteria and can be found within fungal cells. (a) Phylogenetic analysis of endofungal bacteria based on maximum parsimony analysis of 16S ribosomal RNA gene sequences. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) is shown next to the branches. Branches with <70% support values were collapsed. GenBank accession numbers are listed in Supplementary Table S3. Well-known bacterial species not characterized as endofungal symbionts were inserted for reference and are marked with a red asterisk (\*). (b) Confocal laser-scanning microscopy image of GFP-labeled GMI1000 colonizing *A. flavus* chlamydospore. Scale bar (white) represents 10  $\mu$ m. Z-stack scan can be viewed in Supplementary Video S2.

*R. solanacearum* strains to identify a non-chlamydospore-producing isolate. Under these conditions, strain GMI1000 was the only isolate to induce substantial chlamydospore formation in *A. flavus* (Supplementary Figure S1). We selected the non-chlamydospore-inducing *R. solanacearum* strain, K60 for comparative metabolite studies because like GMI1000, the K60 genome was sequenced and publicly available. GMI1000 and K60 interaction zones were excised at 72 h. At this time point, chlamydospores were plentiful but the cultures were not in physical contact. The zones were coated with universal MALDI matrix and subjected to MALDI-TOF IMS (Yang *et al.*, 2012) to explore the spatial distribution of secreted metabolites and to identify differential signals between these isolates. In total, 22 ions were detected in the bacterial fungal interactions. We attributed production of most metabolites to the bacterium or the fungus based on their spatial distributions with respect to the optical image and their presence in individual cultures.

Isolate GMI1000 produced high amounts of four ions that K60 did not produce. Three of these ions clearly diffused beyond the margins of the bacterial colony and reached the margins of the fungal colony. These ions were different ionized forms (protonated, sodiated and potassiated species) of one compound with the exact mass of 1291.7142 (Figure 3a, Supplementary Figure S2). The diffusible nature of this compound suggested it was the chlamydospore-inducing metabolite. This compound was further characterized by tandem mass spectrometry (MS/MS) coupled with molecular networking analysis. The molecular network analyses showed that the fragmentation pattern of this compound was unique to GMI1000 (Figure 3b, Supplementary Figure S3).

#### Mapping compound to gene cluster and assessing bacterial colonization

Using tandem mass spectrometry, an automated peptidogenomics analysis (Kersten *et al.*, 2011) and the program Pep2Path (Medema *et al.*, 2014), we identified characteristic partial peptide fragmentation patterns associated with the signal of interest and identified one fragment exhibiting the following residue sequence tag: Gly-Thr-Ser-Ser-Gly-Phe-Ala (Figure 3b). Although this sequence tag does not explain the entire mass of the metabolite, it does strongly indicate that a non-ribosomal peptide synthase (NRPS) was involved in the synthesis of this compound. Using the *in silico* analysis capabilities of Pep2Path, in combination with AntiSMASH (Medema *et al.*, 2011), we examined the *R. solanacearum* GMI1000 and K60 genomes to identify putative secondary metabolic gene clusters where a major criterion was to find NRPS clusters unique to the GMI1000 genome that matched the partial sequence tag described above.

One such gene cluster contained two large genes encoding a non-ribosomal peptide synthetase-polyketide synthase hybrid (RSp0641) and a NRPS (RSp0642). Several lines of evidence suggested this gene cluster produced the chlamydospore-inducing metabolite: these genes were absent from the K60 genome, and the domain architecture of both RSp0641 and RSp0642 predicted a product best matched to the 1291.7 *m/z* of the signal identified in the IMS studies. In addition, the NRPS adenylation domains were predictive of the amino-acid residues detected in our MS/MS studies (Supplementary Table S4). We termed this new compound ralsolamycin and named RSp0641 and RSp0642 ralsolamycin synthase genes *rmyA* and *rmyB*, respectively (Figure 4a). The protein products of *rmyA-rmyB* appear to a large secondary metabolite megasynthase containing both PKS and NRPS domains (Figure 4b).

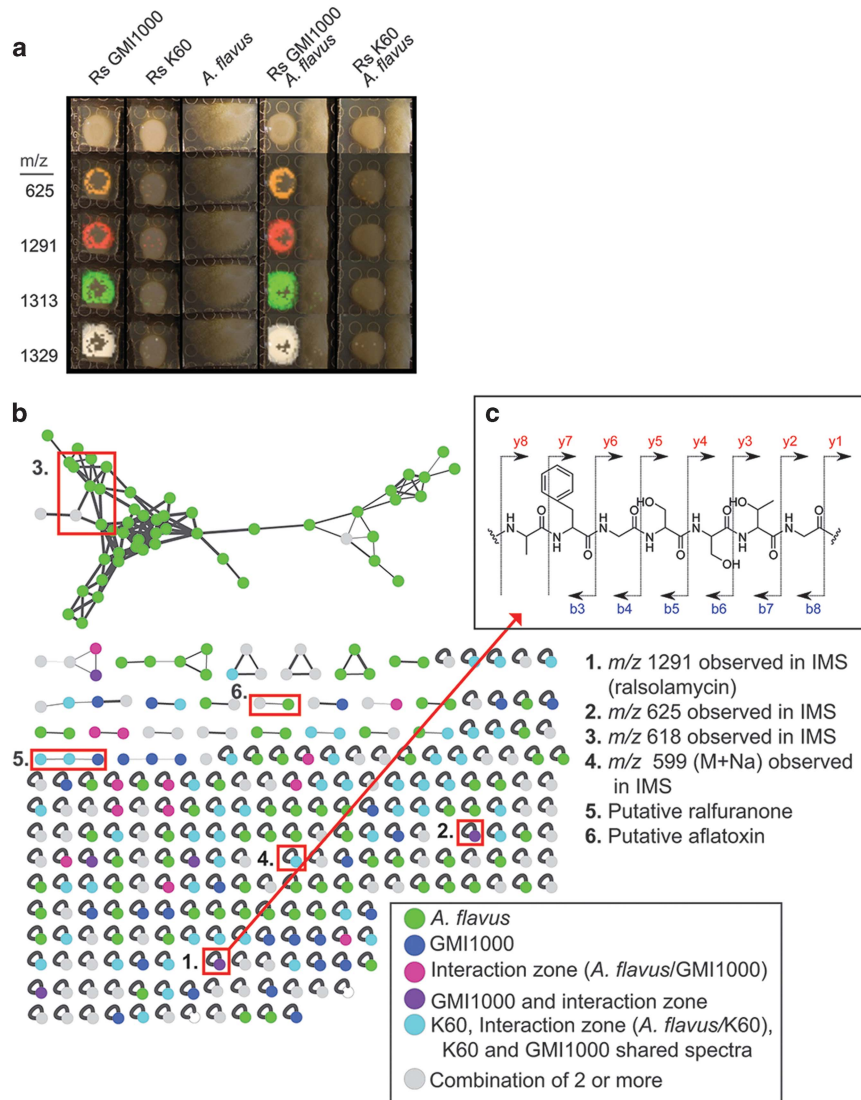
*In silico* analysis using a PKS-NRPS predictor (Bachmann and Ravel, 2009) and domain-specific BLAST searches, suggested that RmyA contains an initiating FAAL domain at its N-terminus followed by a keto-synthase domain, and two acetyl ornithine aminotransferase domains coupled to luciferase-like monooxygenases. The remainder of the *rmyA* gene product contains four canonical NRPS modules containing adenylation (A), peptide carrier protein (PCP) and condensation (C) domains. The predicted peptide product of *rmyA* terminates with a PCP domain. Conventional knowledge of PCP domains suggests that the product would remain tethered to RmyA. Similar analysis of RmyB showed that it initiates with a C domain, suggesting that RmyB functionally associates with the C-terminal PCP domain of RmyA. Further, *rmyB* encodes five NRPS modules, ending with a predicted thioesterase (TE) domain. We predict the TE domain cyclizes and releases the final product. The FAAL and PKS domains of RmyAB suggest the product is a lipopeptide (a fatty acid-like moiety attached to an amino-acid chain). Supplementary Table S4 lists all predicted domain substrates and their closest identified homologs.

We disrupted the putative promoter and FAAL domain of *rmyA* (Liu *et al.*, 2013). The *rmyA* mutant had significantly reduced chlamydospore-inducing activity ( $P < 0.05$ ) in 28 of the 34 assayed fungal cocultures (Table 1), including *A. flavus* (Figure 4c), and lost all ralsolamycin IMS signals (Figure 4d). Furthermore, addition of semi-purified ralsolamycin to *A. flavus* triggers chlamydospore formation (Supplementary Figure S4). These data show that *rmyA* contributes to ralsolamycin biosynthesis and that ralsolamycin induces fungal chlamydospore formation.

#### Ralsolamycin contributes to bacterial invasion of fungal cells

As we saw an abundance of *R. solanacearum* cells internalized in the chlamydospores but not in





**Figure 3** Analysis of differential IMS data sets, MS-MS networks indicate a single lipopeptide produced by *R. solanacearum* strain GMI1000 is responsible for initiating chlamydo-spore formation. **(a)** IMS analysis of strains GMI1000 and K60 showing only differential signals from this data set. Complete data set is in Supplementary Figure S4. **(b)** Network analyses of microbial metabolites detected in the MS-MS studies. The network is composed of nodes representing ions associated with the microbial colonies. Nodes are connected by edges, which represent the relatedness of the fragmentation patterns of each spectrum. Nodes only associated with *A. flavus* colonies are green. Nodes only associated with only GMI1000 are dark blue. Nodes associated with the ‘interaction zone’ (*A. flavus*/GMI1000) are labeled purple. Nodes associated with any of the K60 culture conditions or that are shared between k60 and GMI1000 are colored in light blue. Nodes common to two or more of the previous node classes are labeled in gray. Nodes of interest from the IMS study (1–4) and those that are known products of the microbes (5–6) are highlighted with red boxes and listed on the right of the figure. **(c)** MS/MS spectra of extracted metabolites shows a unique fragmentation pattern of G-T-S-S-G-F-A for the compound with a  $m/z$  of 1291(top right).

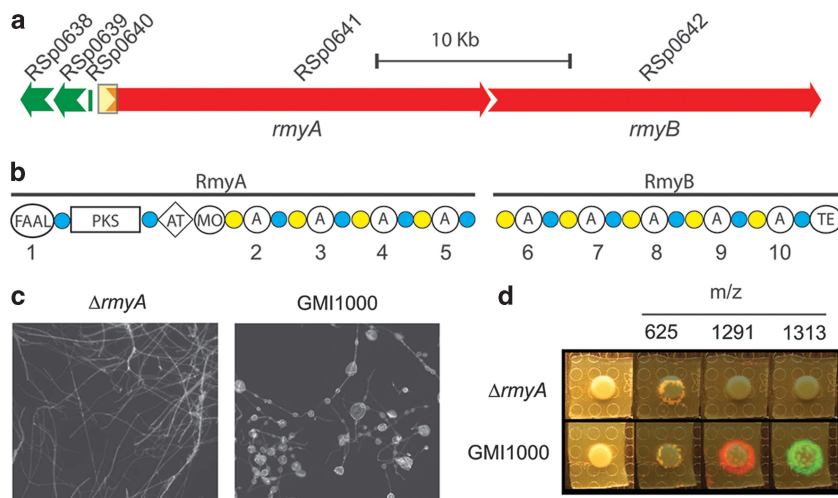
undifferentiated hyphae, we hypothesized that ralsolamycin is necessary for invasion of fungal tissues. To test this hypothesis, GFP-producing GMI1000 wild-type and *rmyA* mutant were cocultured with *A. flavus*. Samples were examined with confocal microscopy using amphiphilic styryl dye FM4-64 as a membrane counterstain. The  $\Delta rmyA$  bacteria rarely invaded the hyphae. In contrast, the wild-type cells were abundant in the chlamydo-spores (Figure 5a). These findings suggest that ralsolamycin

enhances *R. solanacearum*’s invasion of fungal tissues with specificity to chlamydo-spores.

## Discussion

### *Fungal chlamydo-spore development in response to ralsolamycin*

Many fungi produce chlamydo-spores under adverse conditions. However, because not all fungi are



**Figure 4** (a) Graphical representation of the predicted biosynthetic gene cluster as predicted by comparative genomics. Yellow box indicates the region (putative promoter sequence and FAAL encoding sequence) targeted for creation of the *rmyA* mutant using a gentamycin resistance cassette. Genes and their closest protein homologs are listed in Supplementary Table S6. (b) Predicted domain architecture of the hybrid PKS megasynthases RmyA and RmyB. FAAL, fatty acid acyl ligase domain; PKS, polyketides synthase domain; AT, aminotransferase domain; MO, monooxygenase domain; A, adenylation domain; TE, thioesterase domain; blue circles, phosphopantetheine attachment site; yellow circles, condensation domains. Adenylation domains are numbered for reference to their predicted substrates listed in Supplementary Table S4. (c) *rmyA* mutant does not induce fungal chlamydospores. (d) *rmyA* mutant does not produce the ralsolamycin signals ( $m/z$  of 1291 and 1313) but does produce the other GMI1000-specific compound ( $m/z$  625).

known to produce chlamydospores, traditional taxonomic studies use these structures as a trait to delineate evolutionary relationships. Our histological data suggest that chlamydospores formed in response to ralsolamycin are similar to those described in the literature (van Eck, 1978). We provide the first evidence of chlamydospore formation in many of these fungi including all of the *Aspergillus* species tested. Our results also indicate that these structures may be more common across the kingdom Fungi than is currently recognized.

The finding that all tested *Aspergillus* species can produce chlamydospores was unexpected as only *A. parasiticus* chlamydospores have been putatively described (Abou-Gabal and Fagerland, 1981). These data suggest a novel mode of persistence for this well-studied genus. Although most of the assayed fungi formed abundant chlamydospores in response to ralsolamycin, a subset formed relatively few chlamydospores (Table 1). We have not ruled out that these taxa may form chlamydospores more abundantly under other conditions. Perhaps differences in plasma membrane composition or signaling cascades among these fungi temper the conserved chlamydospore formation response to ralsolamycin. Alternatively, these fungi may respond to specific cues shown to affect chlamydospore formation in other fungi including nutrients (Barran et al., 1977; Ohara and Tsuge, 2004), light (Kües et al., 1998), osmolarity (Eisman et al., 2006), pH (Regúlez et al., 1980), air (Barran et al., 1977), temperature (Ohara and Tsuge, 2004) and other interkingdom signals (Li et al., 2005; Staib and Morschhäuser, 2005;

Li et al., 2012). Regardless of quantity, the finding that all of the fungi developed chlamydospores when exposed to ralsolamycin supports a conserved differentiation process in the kingdom Fungi in response to this lipopeptide.

#### Endofungal lifestyle of *R. solanacearum*

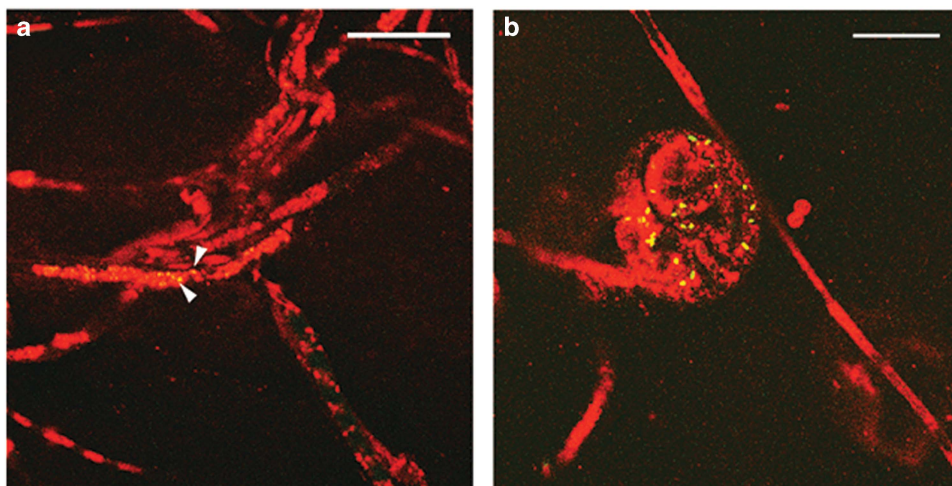
Our data support the hypothesis that *R. solanacearum* colonizes fungal cells, and that formation of the chlamydospores facilitates bacterial entry. Recently, diverse fungal lineages have been shown to harbor endofungal bacteria, most of which are Gram-negative taxa (Hoffman and Arnold, 2010). Of these related genera, *Burkholderia rhizoxinica* and other closely related species have been particularly well studied for their endofungal lifestyle and their role in the production of bioactive secondary metabolites (Partida-Martinez et al., 2007; Lackner et al., 2009; Lackner et al., 2011a, b). Our data analyzing  $\Delta rmyA$  and GMI1000 suggest that although ralsolamycin production is not essential for bacterial invasion of fungal hyphae, it does contribute significantly to this process and/or that chlamydospores may provide a more suitable site for bacterial growth upon invasion.

Many bacterial lipopeptides such as iturin (Maget-Dana and Peypoux, 2003), fengycin (Patel et al., 2011), lichenysin (Madslie et al., 2013) and syringomycin (Hutchison et al., 1995) form pores and permeabilize membranes. We presume that ralsolamycin-mediated membrane restructuring contributes to the initiation of chlamydospore formation

**Table 1** Chlamydospore counts per high-power field (magnification: x200) from interactions with *R. solanacearum* strains GMI1000 and  $\Delta rmyA$

| Fungal species                                 | GMI1000 |       | $\Delta rmyA$ |      | P-value  |
|--|---------|-------|---------------|------|----------|
|  | Mean    | s.d.  | Mean          | s.d. |          |
| <i>Alternaria alternata</i> <sup>a</sup>       | 28.00   | 8.49  | 0.33          | 0.47 | 1.00E-02 |
| <i>Alternaria solani</i>                       | 88.00   | 47.77 | 12.67         | 5.25 | 9.09E-02 |
| <i>Aspergillus aculeatus</i> <sup>a</sup>      | 80.00   | 17.57 | 6.33          | 2.87 | 4.25E-03 |
| <i>Aspergillus brasiliensis</i> <sup>a</sup>   | 136.00  | 24.39 | 85.33         | 8.26 | 4.97E-02 |
| <i>Aspergillus carbonarius</i> <sup>a</sup>    | 36.00   | 11.52 | 4.33          | 1.70 | 1.84E-02 |
| <i>Aspergillus clavatus</i> <sup>a</sup>       | 97.33   | 22.60 | 0.00          | 0.00 | 3.68E-03 |
| <i>Aspergillus flavus</i> <sup>a</sup>         | 105.00  | 17.91 | 0.00          | 0.00 | 1.15E-03 |
| <i>Aspergillus fumigatus</i> <sup>a</sup>      | 11.33   | 0.47  | 0.00          | 0.00 | 4.46E-06 |
| <i>Aspergillus nidulans</i> <sup>a</sup>       | 42.33   | 2.05  | 0.00          | 0.00 | 8.26E-06 |
| <i>Aspergillus niger</i> <sup>a</sup>          | 9.67    | 2.87  | 0.33          | 0.47 | 1.05E-02 |
| <i>Aspergillus oryzae</i> <sup>a</sup>         | 48.33   | 4.64  | 1.00          | 0.82 | 1.43E-04 |
| <i>Aspergillus terreus</i> <sup>a</sup>        | 207.00  | 62.98 | 5.33          | 1.70 | 1.06E-02 |
| <i>Aspergillus tubingensis</i> <sup>a</sup>    | 32.33   | 11.47 | 7.67          | 0.94 | 3.87E-02 |
| <i>Aspergillus zonatus</i> <sup>a</sup>        | 126.00  | 58.76 | 0.00          | 0.00 | 3.87E-02 |
| <i>Botrytis cinerea</i>                        | 2.33    | 1.25  | 0.00          | 0.00 | 5.72E-02 |
| <i>Colletotrichum graminicola</i> <sup>a</sup> | 58.67   | 18.26 | 16.67         | 5.31 | 3.54E-02 |
| <i>Fusarium fujikuroi</i> <sup>a</sup>         | 31.33   | 3.09  | 1.33          | 0.47 | 1.71E-04 |
| <i>Fusarium graminearum</i> <sup>a</sup>       | 180.67  | 39.85 | 0.67          | 0.94 | 3.09E-03 |
| <i>Fusarium oxysporum</i> <sup>a</sup>         | 47.33   | 4.78  | 1.00          | 1.41 | 1.94E-04 |
| <i>Fusarium solani</i> <sup>a</sup>            | 21.67   | 8.73  | 0.00          | 0.00 | 2.47E-02 |
| <i>Fusarium sporotrichioides</i> <sup>a</sup>  | 40.33   | 13.89 | 12.00         | 3.74 | 4.95E-02 |
| <i>Fusarium verticillioides</i> <sup>a</sup>   | 16.00   | 7.07  | 0.00          | 0.00 | 3.29E-02 |
| <i>Morchella esculenta</i> <sup>a</sup>        | 21.00   | 4.55  | 0.00          | 0.00 | 2.84E-03 |
| <i>Mucor bacilliformis</i>                     | 58.00   | 30.01 | 17.67         | 2.87 | 1.31E-01 |
| <i>Mucor hiemalis</i> <sup>a</sup>             | 26.00   | 3.56  | 0.67          | 0.47 | 5.67E-04 |
| <i>Neosartorya fischeri</i> <sup>a</sup>       | 51.00   | 15.77 | 0.67          | 0.94 | 1.08E-02 |
| <i>Penicillium italicum</i> <sup>a</sup>       | 102.00  | 32.26 | 1.67          | 1.25 | 1.17E-02 |
| <i>Phycomyces blakesleeianus</i> <sup>a</sup>  | 6.67    | 1.70  | 0.00          | 0.00 | 5.17E-03 |
| <i>Rhizoctonia solani</i>                      | 27.00   | 11.86 | 7.33          | 8.99 | 1.35E-01 |
| <i>Sclerotinia sclerotiorum</i> <sup>a</sup>   | 84.00   | 11.86 | 5.67          | 3.09 | 8.30E-04 |
| <i>Sordaria fimicola</i> <sup>a</sup>          | 102.33  | 6.65  | 22.00         | 4.55 | 1.47E-04 |
| <i>Trichoderma hazarianum</i>                  | 64.00   | 38.74 | 0.00          | 0.00 | 7.97E-02 |
| <i>Trichothecium roseum</i>                    | 24.33   | 9.88  | 7.33          | 3.30 | 8.22E-02 |
| <i>Verticillium albo-atrum</i> <sup>a</sup>    | 173.00  | 37.98 | 10.00         | 0.82 | 3.73E-03 |

<sup>a</sup>Indicates strains with significantly greater ( $P < 0.05$ ) chlamydospore counts in coculture with GMI1000 relative to strain  $\Delta rmyA$ . Mean, s.d. and P-values are listed to show variations between strains.



**Figure 5** Confocal laser-scanning microscopy shows that (a) the *rmyA* disruption does not completely inhibit bacterial invasion of fungal hyphae (arrowheads indicate GFP-expressing bacteria) but (b) much larger populations of wild-type bacteria can be found in chlamydospores. White bar indicates 10  $\mu\text{m}$ .

and may facilitate the passage of *R. solanacearum* into fungal cells. *R. solanacearum* possesses other factors that could contribute to fungal endosymbioses such as chitinolytic enzymes, known to be important in fungal cell wall reorganization (Adams, 2004), and genes for all of the major Gram-negative protein secretion systems (Genin and Boucher, 2004). The *R. solanacearum* type III secretion machinery shows significant conservation of gene order and primary sequence to that of *B. rhizoxinica*, a system that has been shown to be important for maintaining a stable endofungal symbiosis of *B. rhizoxinica* with the fungus *R. microsporus* (Lackner *et al.*, 2011a) and possibly a similar mechanism contributes to *R. solanacearum* invasion of fungal cells.

*R. solanacearum* is known to persist in soils for remarkably long periods of time relative to other bacterial plant pathogens (Álvarez *et al.*, 2010). The traits underlying environmental persistence are poorly studied, but external factors such as non-host reservoir plants, permissive soil temperatures and high soil moisture content may contribute to persistence. Our findings suggest that *R. solanacearum* colonization within chlamydospores is a novel persistence mechanism for bacterial survival in the absence of host plants. The large numbers of bacteria within colonized chlamydospores indicates that they multiply within chlamydospores, which may contribute to increasing inoculum load once plants are introduced. The chlamydospores contain substantial amounts of lipid droplets, which could contribute to bacterial replication in these specialized structures. Currently, little is known regarding nutritional interactions between bacterial/fungal symbioses (Burlinson *et al.*, 2011) and efforts toward elucidating the nutritional interactions and survival outcomes of the *R. solanacearum*/chlamydospore complex are currently underway in our lab.

As homologs of the ralsolamycin biosynthetic cluster are present in a subset of the *R. solanacearum* strains sequenced to date, it is intriguing to consider that the invasion of chlamydospores may be important to these strains. We did not find any phylogenetic or geographic trends that suggest that ralsolamycin biosynthesis is restricted to a single phylotype, geographic locale or host plant (Supplementary Table S5); however, as more isolates are sequenced a pattern of occurrence may be uncovered. In addition, CMR15 homologs of RmyA and RmyB showed 94% and 93% identity although this strain did not induce chlamydospore formation. These data suggest that either this cluster makes a different metabolite; that the genes do not produce functional proteins; or that the cluster may be differentially regulated under the conditions tested here. Continued research into these strain differences as well as increasing genomic information will help elucidate the impact of ralsolamycin biosynthesis on *R. solanacearum* biology.

### Ralsolamycin identification, characterization and biosynthetic genes

The combination of spatial data from our IMS experiments and the MS/MS network analysis indicates a single compound, ralsolamycin, is largely responsible for the bacterial induction of fungal chlamydospore formation. Multiple *in silico* analyses tools indicate that ralsolamycin is produced by the megasynthase enzyme encoded by the genes *rmyA-rmyB*. These two backbone genes, *rmyA* (a PKS-NRPS hybrid) and *rmyB* (a NRPS), are the two largest ORFs in the GMI1000 genome, approximately 20.7 and 17.9 kb, respectively. The bipartite genomes of strains GMI1000 and K60 were analyzed via AntiSMASH (Medema *et al.*, 2011) to look for canonical backbone secondary metabolite genes. This analysis identified 19 putative secondary metabolic clusters in GMI1000. Of these, only three were predicted to contain putative NRPSs: the micacocidin-producing RSc1806-1811 (Kreutzer *et al.*, 2011), the ralfuranone-producing RSp1419-1434 (Wackler *et al.*, 2011) and the ralsolamycin-producing RSp0638-0642 cluster characterized here. Identification of the 1290 Da metabolite by IMS as the key metabolite was supported by the Pep2Path program, which can link NRPS-products to their biosynthetic genes based on predictions of adenylation substrate recognition domains (Medema *et al.*, 2014). This program mapped the partial amino-acid sequence tag G-T-S-S-G back to the hybrid non-ribosomal peptide synthetase-polyketide synthase backbone gene RSp0641-0642, which encodes 10 predicted adenylation domains in total.

The domain architecture of RmyA is very similar to that responsible for the production of the antifungal lipopeptide mycosubtilin, MycA, produced by *Bacillus subtilis* (Duitman *et al.*, 1999). The presence of the N-terminal FAAL domain followed by a keto-synthase and aminotransferase domain is a known motif in bacterial lipopeptide biosynthesis (Du *et al.*, 2001). These domains cooperate to convert carboxylic acid moieties into activated  $\beta$ -amino acids, which are utilized by subsequent NRPS modules. The remaining modules in RmyA-B appear to have a canonical NRPS adenylation, peptide carrier, condensation domain structure suggesting normal non-ribosomal peptide biosynthesis beyond the initial FAAL domain. We are currently working toward complete characterization of this compound to verify its amino-acid composition.

Although it appears that RmyA and RmyB constitute a single operon, examination of this region suggests that there may be up to five genes involved in its biosynthesis and transport including *rmyA*, *rmyB* and three additional genes: RSp0638, RSp0639 and RSp0640 (Figure 4a) encoding a transmembrane cyclic peptide transporter, a metallo-beta-lactamase and an MbtH-like protein, respectively (Figure 5b, Supplementary Table S6). Comparative analysis of the available *R. solanacearum* strain genomes

showed that these five genes are located within a copper resistance cluster common to most *R. solanacearum* strains. Although we have not verified the functions of RSp0638-40, we predict that they participate in the production and transport of ralsolamycin as they are all common components of secondary metabolic clusters.

## Conclusions

We have demonstrated that the bacterial plant pathogen *R. solanacearum* strain GMI1000 induces chlamydo-spore formation in a wide range of fungal taxa, many of which were not known to produce chlamydo-spores. We used IMS to rapidly identify potential mediators of this diffusible interaction phenotype. Tandem mass spectrometry coupled with *in silico* peptidogenomics tools and molecular genetics identified a putative *R. solanacearum* lipopeptide, ralsolamycin and the PKS-NRPS responsible for its production.

We provide the first evidence that *R. solanacearum* can induce and invade specialized fungal cells, chlamydo-spores, which may have significant implications for environmental persistence of *R. solanacearum*. Chlamydo-spores enhance persistence of soil-borne fungi and related organisms (Couteaudier and Alabouvette, 1990a; Crone et al., 2013) and endosymbiotic mutualisms provide survival benefits (Nikoh et al., 2014; O'Connor et al., 2014). *R. solanacearum* has an unusually long survival time in soil, even when host plants are removed (Álvarez et al., 2010). Our data suggest ralsolamycin-induced chlamydo-spores contributes to bacterial invasion of fungal cells, but ralsolamycin is not the sole factor contributing to this process. We speculate that endofungal colonization benefits *R. solanacearum* survival in soil in the absence of plant–host tissue, and *R. solanacearum* co-opts the accumulated lipids in chlamydo-spores. Further research in our lab aims to determine if this symbiotic phenotype impacts the survival of *R. solanacearum* and if there is any impact on the fungi it interacts with.

## Conflict of Interest

The authors declare no conflict of interest.

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## Author contributions

The majority of the experiments were designed and executed by JES under the guidance of NPK. The IMS and chemical characterization of ralsolamycin was done by LMS under the guidance of PCD. *R. solanacearum* transformations were done by JES and TML.

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