# Obesity, insulin resistance and comorbidities – Mechanisms of association

Obesidade, resistência à insulina e comorbidades -Mecanismos de associação

Ana Valeria B. Castro<sup>1</sup>, Cathryn M. Kolka<sup>2</sup>, Stella P. Kim<sup>2</sup>, Richard N. Bergman<sup>2</sup>

#### **ABSTRACT**

Overall excess of fat, usually defined by the body mass index, is associated with metabolic (e.g. glucose intolerance, type 2 diabetes mellitus (T2DM), dyslipidemia) and non-metabolic disorders (e.g. neoplasias, polycystic ovary syndrome, non-alcoholic fat liver disease, glomerulopathy, bone fragility etc.). However, more than its total amount, the distribution of adipose tissue throughout the body is a better predictor of the risk to the development of those disorders. Fat accumulation in the abdominal area and in non-adipose tissue (ectopic fat), for example, is associated with increased risk to develop metabolic and non-metabolic derangements. On the other hand, observations suggest that individuals who present peripheral adiposity, characterized by large hip and thigh circumferences, have better glucose tolerance, reduced incidence of T2DM and of metabolic syndrome. Insulin resistance (IR) is one of the main culprits in the association between obesity, particularly visceral, and metabolic as well as non-metabolic diseases. In this review we will highlight the current pathophysiological and molecular mechanisms possibly involved in the link between increased VAT, ectopic fat, IR and comorbidities. We will also provide some insights in the identification of these abnormalities. Arq Bras Endocrinol Metab. 2014;58(6):600-9

#### **Keywords**

Obesity; fat distribution; visceral fat; ectopic fat; insulin resistance; metabolic syndrome

#### **RESUMO**

Excesso de gordura, geralmente definido pelo índice de massa corporal, está associado a distúrbios metabólicos (p. ex., intolerância à glicose, diabetes melito tipo 2 (DM2), dislipidemia) e não metabólicos (p. ex., neoplasias, síndrome dos ovários policísticos, esteatose hepática não alcoólica, glomerulopatia, fragilidade óssea etc.). No entanto, mais do que sua quantidade total, a forma da distribuição corporal de tecido adiposo constitui-se em um melhor indicador de risco para o desenvolvimento de tais doenças. O acúmulo de gordura na região abdominal e em tecido não adiposo (gordura ectópica), por exemplo, está associado ao aumento de risco para distúrbios metabólicos e não metabólicos. Por outro lado, observações sugerem que os indivíduos que apresentam adiposidade periférica, caracterizada por aumento das circunferências dos quadris e da coxas, têm melhor tolerância à glicose, redução das incidências de DM2 e da síndrome metabólica. Uma das alterações subjacentes na relação entre a obesidade, particularmente a visceral, e os distúrbios citados é a resistência à insulina. Nesta revisão, enfatizaremos os mecanismos fisiopatológicos e moleculares possivelmente implicados na ligação entre o aumento das gorduras visceral e ectópica, IR e comorbidades. Também mencionaremos os métodos diagnósticos mais frequentemente usados na identificação dessas anormalidades. Arg Bras Endocrinol Metab. 2014;58(6):600-9

#### Descritores

Obesidade; distribuição de gordura; gordura visceral; gordura ectópica; resistência à insulina; síndrome metabólica

Departamento de Clínica
 Médica, Divisão de Endocrinologia
 e Metabolismo, FMRP-USP,
 Ribeirão Preto, SP, Brazil
 Diabetes and Obesity Research

Institute, Cedars Sinai Medical

Center, Los Angeles, CA, USA

Correspondence to:

Ana Valeria B. Castro
Centro Universitário Barão de Mauá,
Departamento de Clínica Médica,
Divisão de Endocrinologia e
Metabolismo,
Faculdade de Medicina,
Universidade de São Paulo,
Ribeirão Preto, SP, Brazil
dra.anavaleria@gmail.com

Received on Jan/13/2014 Accepted on Mar/30/2014

DOI: 10.1590/0004-2730000003223

#### INTRODUCTION

verall excess of fat, usually defined by the body mass index (BMI), has long been recognized as a risk factor for metabolic related diseases, such as cardiovascular diseases (CVD), type 2 diabetes mellitus (T2DM), bone fragility as well as non-metabolic derangements such as non-alcoholic fat liver disease, neoplasias, polycystic ovary syndrome, glomerulopathy, among others (1-6). However, several observations underscored that, more than the excess of fat itself, the distribution of fat, especially in the central regions of the body (also referred to as visceral, omental or intraabdominal fat) plays an important role in these associations (2,7-9). Conversely, peripheral adiposity, characterized by large hips and thighs circumferences, has been associated with a better metabolic profile (10,11).

The advance in imaging methods allowed not solely to confirm those previous anthropometric-based observations but to attempt discriminating which component of the abdominal fat were more predictive of the aforementioned comorbidities (12-15). Those studies showed that although subcutaneous adipose tissue (SAT) is an important determinant of metabolic risk, visceral adipose tissue (VAT) could be a stronger player (14,16).

The link between obesity and the development of IR has been well documented. In African-Americans, for example, it has been observed that both increased abdominal SAT and VAT are strongly correlated with insulin resistance (16). Other studies, however, have shown that individuals with central obesity have higher incidence of insulin resistance than those with subcutaneous obesity (14,15). In fact, some authors believe that insulin resistance (IR) is the main culprit in the association between visceral obesity and metabolic as well as non-metabolic diseases (17).

Moreover, it has also been shown that the presence of fat in ectopic sites such as liver, muscle, pancreas, kidney etc. either alone or in association with increased visceral fat, was also an independent determinant of the development of IR and associated comorbidities (3,6).

Factors that contribute to the preferential accumulation of fat in certain body regions as well to the development of insulin resistance involve modifiable (physical activity levels, nutritional surplus, hormonal status) and non-modifiable factors (age, gender, genetic predisposition and ethnic background) (1,12).

The exact role that adipose tissue, and particularities in its distribution, may play in the pathogenesis of

IR has not yet been fully elucidated. In this review, we will highlight the current mechanisms believed to be involved in the link between increased VAT, ectopic fat, IR and associated comorbidities. We will also provide some insights in the identification of these abnormalities.

# ADIPOSE TISSUE DISTRIBUTION ABNORMALITIES – CLINICAL RELEVANCE

Clinical observations show that disregarding the BMI, either normal-weight or obese individuals may present a healthy or an unhealthy metabolic profile. Accordingly to both BMI and metabolic profile, individuals may be classified in the following phenotypes: 1) lean and healthy, 2) lean and unhealthy (also known as thin outside, fat inside or metabolically obese but normal weight), 3) obese and unhealthy and 4) obese and healthy (also coined as metabolically healthy obese or insulin-sensitive obese) (9,18-20). Some of the main features that distinguish metabolically healthy or unhealthy phenotypes include increased VAT, ectopic fat deposition and insulin resistance in the latter phenotype, which impose on them a higher risk for metabolic and non-metabolic comorbidities than their metabolically healthy counterparts (21).

Lipodystrophic patients who present partial or total loss of SAT (acquired or inherited) and more VAT and ectopic fat accumulation are also more insulin-resistant and more prone to metabolic and non-metabolic comorbidities (22,23). On the other hand, other group comprising patients with predominant subcutaneous fat accumulation, frequently extensive and deforming (multiple symmetric lipomatosis, lipedema and Dercum's disease) usually present a metabolically healthier profile (21,24,25). These clinical situations corroborate the possibility of a protective role of accumulation of fat in SAT for a healthier metabolic profile, in oppose to the accumulation of fat in VAT or in ectopic sites.

# ADIPOSE TISSUE – AN OVERVIEW

There are two main types of adipose tissue in our body, white adipose tissue (WAT) and brown adipose tissue (BAT) that may coexist throughout the adipose tissue sites (1,26-29) (Figure 1). Besides its fat storage role, WAT is considered the largest endocrine in the body (30) and exerts autocrine, paracrine and endocrine functions (e.g. brain, muscle, liver, vessels, kidney, bone etc.) (4,30). BAT is essential to dissipate energy through the regulation of thermogenesis in response to food

intake and cold, sympathetic activation, hormones such as irisin, released by the muscle etc. (31,32).

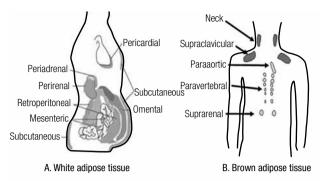


Figure 1. Classification of fat and distribution of white adipose tissue (WAT) and brown adipose tissue (BAT) – (A) WAT is classified in subcutaneous adipose tissue (SAT) and internal adipose tissue. SAT is further subdivided in superficial and deep adipose tissue. Internal adipose tissue is comprised by intrathoracic (*e.g.* pericardial) and visceral adipose tissue (VAT). The latter is further compartmentalized in intraperitoneal fat (greater omentum and mesenteric) and extraperitoneal fat (pre and retroperitoneal). (B) BAT is found along vessels (aorta, carotid, coronary arteries etc.), neck, interscapular and supraclavicular regions, axilla, abdominal wall, inguinal fossa and muscle (not shown). (A – adapted from Cook A. and Cowan C., Adipose (2009)- doi/10.3824/stembook.1.40.1, B – adapted from Awada R, Parimisetty A, Lefebvre d'Hellencourt C (2013)- doi/10.5772/5367).

Adipose tissue has two important features: capacity to expand and plasticity. Expandability reflects the ability of adipose tissue to store lipid, either by the increase of adipocyte size (hypertrophy) and/or by the differentiation or adipogenesis (hyperplasia) of pre-existing adipocytes. As obesity progresses, hypertrophy occurs and reaches a plateau (as observed in massively obese patients). Then, the presence of large adipocytes may recruit new cells triggering adipocytes hyperplasia. One of the characteristic of VAT is its lower proliferation rate and differentiation capacity which leads to growth mainly by hypertrophy, which renders impaired adipocyte functions. On the other hand, SAT (mainly lower body) grows mainly by hyperplasia; however when adipogenesis is limited, hypertrophic dysfunctional adipocytes will take place. Whenever the capacity to expand of both compartments is surpassed, lipid spills over to ectopic sites (non-adipose tissues) (1,33).

In relation to its plasticity, it has been suggested that human WAT is also able to transdifferentiate into brown adipose tissue and vice-versa accordingly to the condition to which the fat is exposed (28,31). For instance, during cold exposure, some WAT may be transformed into brown adipose tissue in order to increase heat production; on the other hand, exposure to an obesogenic diet, BAT is transdifferentiated into WAT enabling greater energy storage (31).

#### **INSULIN RESISTANCE – AN OVERVIEW**

Detailed description of insulin action is out of the scope of this review and has been reviewed elsewhere (34,35).

Insulin resistance is characterized by the impairment of insulin action. Classically, the insulin-resistant state is defined by the impairment of glucose uptake in muscle and the increment of endogenous glucose production by the liver resulting in hyperglycemia, both in fasting and postprandial states. However, in a broader sense, the insulin resistant state is also characterized by the impairment of insulin action on lipid metabolism (e.g. increment of lipolysis in adipocytes) or on protein metabolism (e.g. impairment of protein synthesis in muscle, predisposing to sarcopenia). Also, IR affects the function of other organs such as vessels (leading to vasoconstriction/hypertension); brain (resulting in increased caloric intake); pancreas (decreased in beta-cell mass and in glucose sensing); bone (possibly decreasing bone mass and strength) among others effects (4,33).

Notably, the cellular mitogenic effect of insulin is preserved in insulin resistant states leading to cell growth (e.g. acantosis nigricans, predisposition to cancer). Moreover, the insulin may bind to other receptors such as the IGF-1 receptor in the cartilage (promoting acromegaloid features), or in the theca cell in the ovary (leading to excessive androgen production and secretion, hirsutism and polycystic ovary syndrome) (1,4,36,37).

The mechanisms involved in the etiopathogenesis of insulin resistance related to obesity encompass pre-receptor, receptor and postreceptor defects characterized, for example, by decreased access of insulin to muscle secondary to FFA excess (pre-receptor), insulin receptor downregulation secondary to hyperinsulinemia (receptor) and inhibition of the intracellular cascades by several adiposity-related factors (e.g. increased FFA, impaired adipokines and/or cytokines secretion) (post receptor) (36). All these aspects are discussed further and are depicted in figure 2.

# CENTRAL OBESITY, INSULIN RESISTANCE AND COMORBIDITIES – MECHANISMS OF ASSOCIATION

Pathophysiological mechanisms (Figure 2)

It is uncertain how obesity results in insulin resistance; however, several factors have been implicated as playing a pivotal role, as described in the following sections and depicted in figure 2.

Figure 2. Summary of the pathophysiological mechanisms associated with the development of insulin resistance associated with obesity and comorbidities - Hormonal status (e.g. menopause), aging, gender, genetic susceptibility and ethnic background interact with lifestyle factors to predispose to the increase of VAT/ectopic fat and the development of IR. Energy surplus secondary to nutritional factors (e.g., high energy food intake) associated with low physical activity levels lead to an increase in SAT/VAT. When the capacity of these tissues to expand becomes saturated (obesity) or limited (lipodystrophy), lipids spill over to non-adipose tissue sites (ectopic fat deposition). Fat growth by hypertrophy generates dysfunctional adipocytes that are more resistant to insulin's antilipolytic effect and present impaired secretion of cytokines/adipokines (e.g. decreased adiponectin, increased TNFalpha and IL-6). Consequently, FFA and cytokines are released into the circulation. The surplus of FFA to the cells is oxidized, stored (lipids droplets) or metabolized into toxic derivatives (DAG and ceramides). These toxic derivatives lead to insulin resistance, impair cell function (lipotoxicity) or lead to apoptosis (lipoapoptosis). In the pancreas these toxic effects lead to decreased number and impaired capacity of β-cells to secrete insulin, predisposing to the development of type 2 diabetes mellitus; in the liver it leads to non-alcoholic steatohepatitis and subsequently to cirrhosis; in the muscle, to sarcopenia; in the kidney to glomerulopathy etc. Cell dysfunction and death elicit macrophage infiltration, and local and systemic inflammation. In addition, the secretion of inflammatory molecules into the circulation also impairs intracellular insulin signaling. The consequent insulin resistance increases endogenous glucose production by the liver and decreases glucose utilization by peripheral tissues (e.g. muscle). Consequently, glycemia rises and promotes increase in insulin secretion by the pancreas. In addition, hepatic insulin clearance is impaired contributing to hyperinsulinemia, which promotes down regulation of insulin receptor. Among several effects, hyperinsulinemia promotes cell growth (e.g. acanthosis nigricans, neoplasias) and leads to endothelial dysfunction (increased vasoconstriction).

SAT: subcutaneous adipose tissue; VAT: visceral adipose tissue; BAT: brown adipose tissue; FFA: free fatty acids; mo-macrophage; TG: triglycerides; SNS: sympathetic nervous system; DM: type 2 diabetes mellitus; DAG: diacylglycerol; NAFLD: non-alcoholic fatty liver disease; PCOS: polycystic ovary syndrome; HBP: high blood pressure.

# Dysfunctional adipose tissue

Adipocytes and adipose tissue are key players in the pathogenesis of insulin resistance associated with obesity. Hypertrophic dysfunctional adipocytes, mainly encountered in VAT and upper body SAT, are highly lipolytic resulting in enhanced free fatty acids (FFA) release as well as in impaired secretion of adipokines (increased leptin and resistin, decreased adiponectin etc.) into the circulation.

Two main hypotheses, not mutually exclusive, explain how elevated FFA may be associated with central obesity and IR: the portal hypothesis and the spillover (or ectopic fat) hypothesis. Accordingly to the portal theory an increase in central abdominal fat tissue leads to an elevation of the delivery of FFA to the liver through Accordingly to the "spillover hypothesis", in face of positive energy balance, a reduced or limited ability of adipose tissue to expand (especially the peripheral subcutaneous compartment) would lead to a spillover of FFA to the visceral fat compartment and to non-adipose tissues (e.g. liver, muscle, pancreas, kidney, bone) (4,6,40). Consequent to the limited ability of non-adipose tissue to oxidize and/or to store FFA, ectopic accumulation of FFA and/or its metabolic active derivatives would lead to IR as well as cell lipotoxicity and apoptosis compromising the function of the involved organs (40).

Moreover, hypertrophic adipocytes may lead to local hypoxia driving endoplasmic reticulum (ER) stress, adipocyte death and macrophage infiltration. The latter event increases the secretion of inflammatory cytokines, such TNF- $\alpha$  and interleukin (IL)-6 and monocytes chemoattractant protein (MCP)-1 which lead to local and systemic low grade inflammation (1,12) and the consequent impairment of insulin signaling.

### Decreased insulin clearance/hyperinsulinemia

Insulin blood levels result from a balance between insulin production by the pancreas and its clearance mainly by the liver as well other sites (kidney, muscle and adipose tissue) (41,42). Insulin clearance occurs by the uptake and the degradation of insulin in many tissues. In the liver, insulin uptake is mainly receptor-mediated. Both uptake and degradation are regulated by many factors, including increased FFA that inhibits those processes (36).

In the obesity state, hyperinsulinemia ensues both due to increased production of insulin induced by the excess of fatty-acid and glucose as well as by the decrease of insulin clearance by the liver (and possibly kidney and other sites) (36). Hyperinsulinemic states, promote downregulation of insulin receptors leading to a decrease of insulin removal from circulation. Hyperinsulinemia also promotes insulin resistance due to a negative feedback loop that inhibits IRS-1/2 (36).

Decreased insulin clearance is also mediated by impairments of CEACAM1 expression, a key regulator of hepatic insulin clearance that is ubiquitously present

in the body. In diet-induced obese dogs, it has been demonstrated that prospective changes in CEACAM1 tracks alterations in insulin clearance (43).

Recently, the results from a large cohort study in dogs, emphasized the importance of insulin clearance as the primary determinant of insulin sensitivity under baseline, overnight-fasted conditions (41). The authors proposed that insulin clearance impairment could be the primary defect leading to insulin resistance.

# Abnormalities in the hypothalamus-pituitaryadrenal-fat axis

Similarities between metabolic derangements and insulin resistance associated with glucocorticoid excess states and obesity are well known. However, the mechanisms involved in these similarities are still a matter of debate (1,44).

Excess of cortisol, as seen in Cushing syndrome and chronic stress, is associated with the development of abdominal obesity, insulin resistance and metabolic disorders. Conversely, abdominally obese subjects may also present abnormalities of hypothalamus-pituitary-adrenal axis such as abnormal diurnal variation of cortisol, lack of meal response to cortisol etc. In addition, it has been also shown that visceral fat is more sensitive to the action of cortisol (44).

On the other hand, besides the effect of the systemic cortisol on the adipocytes, it has been shown that visceral fat also produces cortisol. The paracrine action of cortisol in the adipocyte may interfere with several aspect of adipose tissue by increasing adipogenesis, altering adipocyte metabolism (e.g. increasing or decreasing lipolysis), impairing adipokines secretion besides promoting IR (1,44).

# Sympathetic nervous system (SNS) overdrive

High levels of circulating catecholamines lead to insulin resistance (45). One of the outcomes of increased adrenergic outflow is increased lipolysis which promotes excessive influx of FFA to the cells. Obesity, especially visceral, is associated with increased SNS activity.

Activation of SNS has also been implicated in the transdifferentiation of WAT into BAT. Absence of catecholaminergic receptors has been associated with decreased BAT activity and development of obesity (31).

Moreover, the SNS controls the intensity of the immune response. Reciprocally, the immune system controls the thermogenic effect of SNS via cytokines (46).

# Decreased brown or beige adipocytes

Activity of BAT is negatively correlated with age and BMI. In human, it has been shown that cold-induced activity of BAT is reduced with age, and that *ucp1* expression is decreased in the subcutaneous WAT in obese and diabetic patients (27,29). It has been shown that the decrease of BAT is associated with insulin resistance and hyperglycemia in older mice. However, the role of BAT in the pathogenesis of IR in humans is still speculative.

# **Molecular mechanisms (Figure 3)**

The following molecular mechanisms, summarized in figure 3, have been implicated in the development of IR associated with obesity:

# Lipotoxicity/lipoapoptosis

Lipotoxicity is a morphological and functional impairment of non-adipose tissue caused by toxic reactions secondary to intracellular accumulation of lipids and their derivatives that may lead to cell death (lipoapoptosis) (47).

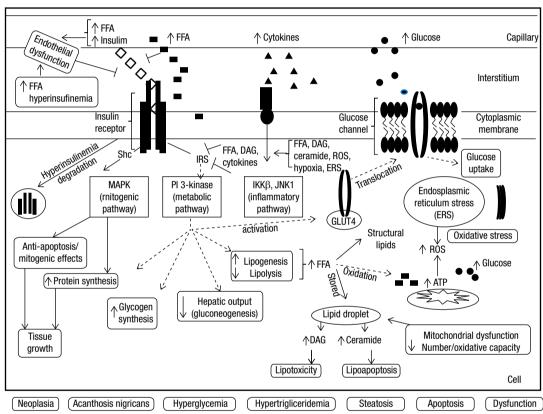


Figure 3. Summary of the main putative molecular mechanisms involved in the development of insulin resistance associated with obesity in a hypothetical cell (e.g. hepatocyte, myocyte, adipocyte) – Insulin resistance associated with obesity, especially central, occurs due to pre-receptor, receptor and/or post-receptor impairments, mainly secondary to elevated FFA, hyperinsulinemia and increased cytokines. Insulin access to the interstitial space (pre-receptor impairment) may be induced by the excess of FFA and their metabolites as well by endothelial dysfunction secondary to increased circulating insulin. Hyperinsulinemia, secondary to the decrease of FFA-induced insulin clearance and the increase of insulin secretion, causes downregulation of insulin receptors (receptor impairment). In addition, insulin receptor downstream signaling (post-receptor impairment) is inhibited by FFA and cytokines. Increased intracellular FFA also contribute to excessive production of ATP, oxidative stress and mitochondrial dysfunction, production of reactive oxidative species, endoplasmic reticulum stress and lipid storage and accumulation of non-oxidative toxic derivatives (dyacylgycerol and ceramide). The aforementioned factors also activate inflammation pathways. Independently of the upstream or downstream level of the insulin receptor impairment, insulin resistance occurs by the inhibition of the phosphorylation of the insulin receptor substrates (IRS-1 or 2) and the subsequent inhibition of PI3K pathway, responsible for metabolic effects. Consequently to this inhibition occurs 1) decrease in the activation of GLUT-4 which impairs glucose uptake; 2) increase of glucose production by the liver (either by inhibiting glucogenesis and/or promoting glycogenolysis) and 3) increase of de novo lipogenesis, storage of lipid (lipid droplets) and toxic derivatives. On the other hand, the insulin receptor substrate, Shc, is spared from inhibition by FFA or cytokines and is stimulated by hyperinsulinemia. Consequently, MAPK pathway (mitogenic) is activated leading to anti-apoptotic and proliferation effects, culminating with tissue growth. Metabolic and non-metabolic consequences of IR and ectopic accumulation of fat are: hyperglycemia, hypertriglyceridemia, acanthosis nigricans, neoplasias, steatosis, cell growth or apoptosis or cell dysfunction.

FFA: free fatty acids; IRS: insulin receptor substrates; MAPK: mitogen-activated protein kinase; PI3K: phosphoinositide 3-kinase; DAG: dyacylgycerol; ROS: reactive oxidative species; ERS: endoplasmic reticulum stress, AN: acanthosis nigricans.

FFA per se and the intracellular accumulation of some species of DAG leads to the impairment of insulin action directly by inhibition of downstream pathways of insulin signaling (47,49). Accumulation of ceramide, a pro-apoptotic lipid, leads to cell death. Accordingly to the respective tissue, ceramide may ultimately contribute to sarcopenia, heart failure,  $\beta$ -cell mass decline, cirrhosis, glomerulopathy etc. (6,47).

Although the ectopic deposition of FFA within the tissue or the cell (lipid droplets) is of major importance when considering the links between obesity, dyslipidemia, insulin resistance and comorbidities, some authors suggest that the ectopic lipid droplet accumulation itself is not harmful to the cell, but it is rather a marker of excessive fuel availability (47). They argue that the toxic FFA-derivative accumulation, originated from non-oxidative metabolism of excessive intracellular lipids, may actually be the actual culprit linking fat distribution abnormalities, insulin resistance and associated disorders.

# Endoplasmic reticulum stress (ERS)

In the presence of several stimuli, such as excess of lipids and glucose (as well as nutrient deprivation) and hypoxia, the ability of endoplasmic reticulum (ER) to fold proteins becomes impaired. This results in the accumulation of misfolded or unfolded proteins that elicits ERS. The aim of ERS is to stop translation of the misfolded protein and activate molecular processes to fold them, preventing any further ER stress. When this aim is not achieved ERS elicits reactive oxygen species (ROS) accumulation, inflammatory response, cellular metabolic dysfunction and apoptosis. In target tissues for example, ERS may culminate with insulin and leptin resistance and decreased insulin secretion (49).

# **Decreased fat oxidation capacity**

Adipose tissue is a key player in controlling oxidative capacity of other organs, such as muscle, through the production and secretion of adipokines, such as leptin, adiponectin and TNF $\alpha$  (47,49). It has been suggested that subjects that have an increased muscle capacity to oxidize FFA are less susceptible to obesity. On the other hand, it has been shown that reduction of the capacity of skeletal muscle to oxidize FFA is associated with insulin resistance (50).

Sarcopenia associated with aging and lack of exercise is one factor leading to decreased FFA oxidative capacity and fat accumulation (18,50). On the other hand, the increment of oxidative capacity by physical activity or pharmacological intervention (metformin, glitazones, leptin) also improves insulin sensitivity (42,51).

# Mitochondrial dysfunction

Human and experimental studies have suggested that mitochondrial dysfunction is involved in the pathogenesis of insulin resistance and ectopic accumulation of fat (26). However, the direction of the relationship is still unclear; it may be cause or consequence of insulin resistance (6,36).

Excessive supply of substrate to the cell may overwhelm the capacity of the mitochondria to oxidize nutrients, diverting the substrate to *de novo* lipogenesis and predisposing to the production of ROS (26). The surplus of lipids to liver, muscle, and brown fat over activates mitochondrial function to enhance energy disposal, promoting the production of high levels of ATP. As a protective mechanism against excessive ATP levels, intracellular energy accumulation ultimately leads to the inhibition of insulin-induced glucose uptake response in those tissues (49). A reduction of mitochondrial capacity of fat oxidation may also accelerate deposition of ectopic fat and their deleterious metabolites ultimately resulting in inhibition of insulin signaling (26,36,52).

#### **Oxidative stress**

Reactive oxygen species are required for normal cell function. Normally, the production of ROS is quenched by endogenous antioxidant mechanisms, present in the peroxisomes, preventing its deleterious consequences (49,52).

However, mitochondrial dysfunction, inflammation and glycation, among other mechanisms, lead to the accumulation of ROS that overcomes the endogenous antioxidant capacity. As a result of ROS accumulation, increased lipid peroxidation and protein misfolding (leading to endoplasmic reticulum stress) occur, ultimately leading to DNA and cellular damage and/or metabolic dysfunction (1).

# FAT DISTRIBUTION ASSESSMENT

It has become evident that assessments of total, regional and ectopic adiposity are important for predicting cardiometabolic risk, for identifying the fat distribution phenotype as well as for obesity treatment follow up. Several publications have discussed in details the methods used to assess body composition (13,53,54). Here we briefly cite current and most commonly used methodology to assess fat distribution.

Clinical methods such as BMI and body adiposity index (BAI) make possible to estimate total body fat but not regional adiposity (55); waist circumference (WC), waist to hip ratio (WHR) and sagittal abdominal diameter and bioimpedance may predict or estimate abdominal fat, but do not distinguish between SAT and VAT (1,12). Some authors propose that the "hypertriglyceridemic waist" that combines WC and trygliceridemia values could also predict visceral obesity. They showed that, in large studies, patients presenting values of both parameters above the cut-off references were more prone to metabolic risk and coronary disease (1).

Computadorized tomography and MRI are currently the recommended methods to directly assess SAT, VAT and ectopic fat deposition. Magnetic resonance spectroscopy is the gold standard to evaluate ectopic fat deposition and positron emission computerized tomography (PET/CT) is currently the gold standard to evaluate BAT. The validation to assess BAT by MRI is also under way (13). However, there is still no consensus of optimal cut-offs of fat measurements in those sites for predicting cardiometabolic risk.

Dual energy X-ray absorptiometry (DXA) and plethysmography precisely assess both total and regional adiposity, and indirectly estimate visceral fat. Ultrasound is suitable to estimate SAT and VAT, however due to the variability and subjectivity of the measurements of intraabdominal adiposity it is not currently recommended for VAT estimation (13,54).

# **INSULIN SENSITIVITY ASSESSMENT**

Taking into account that insulin resistance is an important link between adiposity and comorbidities, its assessment becomes useful.

Severe insulin resistance, as seen in lipodystrophic syndromes, may be recognized in clinical grounds by phenotypical features at birth such as hypertrichosis, severe acanthosis nigricans, pseudoacromegalia or elf-like facies. However, the clinical recognition of milder insulin resistance may be more challenging and may require quantitative assessment (36).

Currently, the majority of the quantitative methods to assess insulin sensitivity are restricted to research settings. The ones considered reference techniques (euglycemic hyperinsulinemic clamp-EGC and frequent-sampling intravenous glucose tolerance test) pose important limitations to their use in clinical practice for they are costly, time consuming and/or technically demanding. Moreover, the gold standard method to assess IR, the EGC, is not fully "physiological" (56).

More feasible, cost-effective and convenient alternatives to the reference methods have been proposed and, potentially could be used in routine clinical research (56). They are fasting or OGTT-derived indices from insulin alone or along with glucose measurements (e.g. HOMA-IR, QUICKI etc.), fasting levels of SHBG, IGFBP-1, ratio leptin:adiponectin among others (57,58). However, some limitations of the aforementioned indices are biological and analytical variability, interference of medications and dependency on glucose tolerance degree levels. Moreover, they lack validated cut-offs values for different populations and conditions. Therefore, their application is not currently recommended in clinical practice.

Some authors proposed decision rules, validated by EGC, to define individuals with insulin resistance. They showed that BMI, WC, HOMA-IR, fasting plasma insulin and LDL cholesterol are strong predictors of IR. Using three different combinations of those clinical and laboratory measurements they were able to identify insulin-resistant individuals (mainly Caucasians and normal-weight subjects) (59). However, the use of these decision rules in other clinical settings is still unknown.

Detailed description of methods to assess insulin sensitivity has been extensively described elsewhere (56,60).

# CONCLUSION

Body fat distribution, more than the total amount of fat, is relevant to assess the risk of developing metabolic and non-metabolic morbidities associated with insulin resistant individuals.

Considerable advancements in understanding the links binding the obesity, fat distribution abnormalities, insulin resistance and comorbidities have been achieved. Nonetheless, determination of validated, feasible and cost-effective tools to assess fat distribution and insulin sensitivity as well as safe and efficient targeted pharmacological interventions tackling the putative mechanisms involve in this link are still warranted.

Disclosure: no potential conflict of interest relevant to this article was reported.

# REFERENCES

- Tchernof A, Despres JP. Pathophysiology of human visceral obesity: an update. Physiol Rev. 2013;93(1):359-404.
- Vazquez G, Duval S, Jacobs DR Jr, Silventoinen K. Comparison of body mass index, waist circumference, and waist/hip ratio in predicting incident diabetes: a meta-analysis. Epidemiol Rev. 2007;29:115-28
- Fabbrini E, Sullivan S, Klein S. Obesity and nonalcoholic fatty liver disease: biochemical, metabolic, and clinical implications. Hepatology. 2010;51(2):679-89.
- 4. Kawai M, de Paula FJ, Rosen CJ. New insights into osteoporosis: the bone-fat connection. J Intern Med. 2012;272(4):317-29.
- Després JP. Body fat distribution and risk of cardiovascular disease: an update. Circulation. 2012;126(10):1301-13.
- Guebre-Egziabher F, Alix PM, Koppe L, Pelletier CC, Kalbacher E, Fouque D, et al. Ectopic lipid accumulation: a potential cause for metabolic disturbances and a contributor to the alteration of kidney function. Biochimie. 2013;95(11):1971-9.
- Vague J. The degree of masculine differentiation of obesities: a factor determining predisposition to diabetes, atherosclerosis, gout, and uric calculous disease. 1956. Nutrition. 1999;15(1):89-90; discussion 1.
- Ohlson LO, Larsson B, Svardsudd K, Welin L, Eriksson H, Wilhelmsen L, et al. The influence of body fat distribution on the incidence of diabetes mellitus. 13.5 years of follow-up of the participants in the study of men born in 1913. Diabetes. 1985;34(10):1055-8.
- Peppa M, Koliaki C, Papaefstathiou A, Garoflos E, Katsilambros N, Raptis SA, et al. Body composition determinants of metabolic phenotypes of obesity in nonobese and obese postmenopausal women. Obesity (Silver Spring). 2013;21(9):1807-14.
- Snijder MB, Visser M, Dekker JM, Goodpaster BH, Harris TB, Kritchevsky SB, et al. Low subcutaneous thigh fat is a risk factor for unfavourable glucose and lipid levels, independently of high abdominal fat. The Health ABC Study. Diabetologia. 2005;48(2):301-8.
- Zhang X, Hu EA, Wu H, Malik V, Sun Q. Associations of leg fat accumulation with adiposity-related biological factors and risk of metabolic syndrome. Obesity (Silver Spring). 2013;21(4):824-30.
- Wajchenberg BL. Subcutaneous and visceral adipose tissue: their relation to the metabolic syndrome. Endocr Rev. 2000;21(6):697-738.
- Machann J, Horstmann A, Born M, Hesse S, Hirsch FW. Diagnostic imaging in obesity. Best Pract Res Clin Endocrinol Metab. 2013;27(2):261-77.
- HayashiT, Boyko EJ, McNeely MJ, Leonetti DL, Kahn SE, Fujimoto WY. Visceral adiposity, not abdominal subcutaneous fat area, is associated with an increase in future insulin resistance in Japanese Americans. Diabetes. 2008;57(5):1269-75.

- Preis SR, Massaro JM, Robins SJ, Hoffmann U, Vasan RS, Irlbeck T, et al. Abdominal subcutaneous and visceral adipose tissue and insulin resistance in the Framingham heart study. Obesity (Silver Spring). 2010;18(11):2191-8.
- Tulloch-Reid MK, Hanson RL, Sebring NG, Reynolds JC, Premkumar A, Genovese DJ, et al. Both subcutaneous and visceral adipose tissue correlate highly with insulin resistance in african americans. Obes Res. 2004;12(8):1352-9.
- Gallagher EJ, Leroith D, Karnieli E. Insulin resistance in obesity as the underlying cause for the metabolic syndrome. Mt Sinai J Med. 2010;77(5):511-23.
- Prado CM, Wells JC, Smith SR, Stephan BC, Siervo M. Sarcopenic obesity: a critical appraisal of the current evidence. Clin Nutr. 2012;31(5):583-601.
- Jennings CL, Lambert EV, Collins M, Joffe Y, Levitt NS, Goedecke JH. Determinants of insulin-resistant phenotypes in normal-weight and obese Black African women. Obesity (Silver Spring). 2008;16(7):1602-9.
- Succurro E, Marini MA, Frontoni S, Hribal ML, Andreozzi F, Lauro R, et al. Insulin secretion in metabolically obese, but normal weight, and in metabolically healthy but obese individuals. Obesity (Silver Spring). 2008;16(8):1881-6.
- Messier V, Karelis AD, Prud'homme D, Primeau V, Brochu M, Rabasa-Lhoret R. Identifying metabolically healthy but obese individuals in sedentary postmenopausal women. Obesity (Silver Spring). 2010;18(5):911-7.
- Dinges WL, Chen D, Snell PG, Weatherall PT, Peterson DM, Garg A. Regional body fat distribution in HIV-infected patients with lipodystrophy. J Investig Med. 2005;53(1):15-25.
- 23. Garg A. Clinical review#: lipodystrophies: genetic and acquired body fat disorders. J Clin Endocrinol Metab. 2011;96(11):3313-25.
- Chen K, Xie Y, Hu P, Zhao S, Mo Z. Multiple symmetric lipomatosis: substantial subcutaneous adipose tissue accumulation did not induce glucose and lipid metabolism dysfunction. Ann Nutr Metab. 2010:57(1):68-73.
- Herbst KL. Rare adipose disorders (RADs) masquerading as obesity. Acta Pharmacol Sin. 2012;33(2):155-72.
- Medina-Gómez G. Mitochondria and endocrine function of adipose tissue. Best Pract Res Clin Endocrinol Metab. 2012;26(6):791-804.
- Wu J, Cohen P, Spiegelman BM. Adaptive thermogenesis in adipocytes: is beige the new brown? Genes Dev. 2013;27(3):234-50.
- 28. Gesta S, Tseng YH, Kahn CR. Developmental origin of fat: tracking obesity to its source. Cell. 2007;131(2):242-56.
- Sacks H, Symonds ME. Anatomical locations of human brown adipose tissue: functional relevance and implications in obesity and type 2 diabetes. Diabetes. 2013;62(6):1783-90.
- Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. J Clin Endocrinol Metab. 2004;89(6):2548-56.
- Gil A, Olza J, Gil-Campos M, Gomez-Llorente C, Aguilera CM. Is adipose tissue metabolically different at different sites? Int J Pediatr Obes. 2011;6 Suppl 1:13-20.
- 32. Cinti S. Between brown and white: novel aspects of adipocyte differentiation. Ann Med. 2011;43(2):104-15.
- Jo J, Gavrilova O, Pack S, Jou W, Mullen S, Sumner AE, et al. Hypertrophy and/or hyperplasia: dynamics of adipose tissue growth. PLoS Comput Biol. 2009;5(3):e1000324.
- 34. Zecchin HG, Carvalheira JBC, Saad MJA. Mecanismos moleculares de resitência à insulina na síndrome metabólica. Rev Soc Cardiol Estado de São Paulo. 2004;14(4):574-89.
- Taniguchi CM, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. Nat Rev Mol Cell Biol. 2006;7(2):85-96.
- Stears A, O'Rahilly S, Semple RK, Savage DB. Metabolic insights from extreme human insulin resistance phenotypes. Best Pract Res Clin Endocrinol Metab. 2012;26(2):145-57.

- Dunaif A. Insulin resistance and the polycystic ovary syndrome: mechanism and implications for pathogenesis. Endocr Rev. 1997;18(6):28.
- Björntorp P. "Portal" adipose tissue as a generator of risk factors for cardiovascular disease and diabetes. Arteriosclerosis. 1990;10(4):493-6.
- 39. Item F, Konrad D. Visceral fat and metabolic inflammation: the portal theory revisited. Obes Rev. 2012;13 Suppl 2:30-9.
- Virtue S, Vidal-Puig A. Adipose tissue expandability, lipotoxicity and the metabolic syndrome--an allostatic perspective. Biochim Biophys Acta. 2010;1801(3):338-49.
- Ader M, Stefanovski D, Kim SP, Richey JM, Ionut V, Catalano KJ, et al. Variable hepatic insulin clearance with attendant insulinemia is the primary determinant of insulin sensitivity in the normal dog. Obesity (Silver Spring). 2013 Sep 30. PubMed PMID: 24123967. Epub 2013/10/15. Eng.
- 42. Home PD, Pacini G. Hepatic dysfunction and insulin insensitivity in type 2 diabetes mellitus: a critical target for insulin-sensitizing agents. Diabetes Obes Metab. 2008;10(9):699-718.
- Kabir M, Catalano KJ, Ananthnarayan S, Kim SP, Van Citters GW, Dea MK, et al. Molecular evidence supporting the portal theory: a causative link between visceral adiposity and hepatic insulin resistance. Am J Physiol Endocrinol Metab. 2005;288(2):E454-61.
- Lee MJ, Pramyothin P, Karastergiou K, Fried SK. Deconstructing the roles of glucocorticoids in adipose tissue biology and the development of central obesity. Biochim Biophys Acta. Biochim Biophys Acta. 2014;1842(3):473-81.
- Lambert GW, Straznicky NE, Lambert EA, Dixon JB, Schlaich MP. Sympathetic nervous activation in obesity and the metabolic syndrome--causes, consequences and therapeutic implications. Pharmacol Ther. 2010;126(2):159-72.
- 46. Garg A. Adipose tissue dysfunction in obesity and lipodystrophy. Clin Cornerstone. 2006;8 Suppl 4:S7-S13.
- Slawik M, Vidal-Puig AJ. Lipotoxicity, overnutrition and energy metabolism in aging. Ageing Res Rev. 2006;5(2):144-64.

- Morelli M, Gaggini M, Daniele G, Marraccini P, Sicari R, Gastaldelli A. Ectopic fat: the true culprit linking obesity and cardiovascular disease? Thromb Haemost. 2013;110(4):651-60.
- Ye J. Mechanisms of insulin resistance in obesity. Front Med. 2013;7(1):14-24.
- Sakuma K, Yamaguchi A. Sarcopenic obesity and endocrinal adaptation with age. Int J Endocrinol. 2013;2013:204164.
- Roberts CK, Hevener AL, Barnard RJ. Metabolic syndrome and insulin resistance: underlying causes and modification by exercise training. Compr Physiol. 2013;3(1):1-58.
- Bremer AA, Mietus-Snyder M, Lustig RH. Toward a unifying hypothesis of metabolic syndrome. Pediatrics. 2012;129(3):557-70.
- Shuster A, Patlas M, Pinthus JH, Mourtzakis M. The clinical importance of visceral adiposity: a critical review of methods for visceral adipose tissue analysis. Br J Radiol. 2012;85(1009):1-10.
- Thomas EL, Parkinson JR, Frost GS, Goldstone AP, Dore CJ, McCarthy JP, et al. The missing risk: MRI and MRS phenotyping of abdominal adiposity and ectopic fat. Obesity (Silver Spring). 2012;20(1):76-87.
- Bergman RN. A better index of body adiposity. Obesity (Silver Spring, Md). 2012;20(6):1135. PubMed PMID: 22627975. Epub 2012/05/26. eng.
- Borai A, Livingstone C, Kaddam I, Ferns G. Selection of the appropriate method for the assessment of insulin resistance. BMC Med Res Methodol. 2011;11:158.
- Monzillo LU, Hamdy O. Evaluation of insulin sensitivity in clinical practice and in research settings. Nutr Rev. 2003;61(12):397-412.
- Finucane FM, Luan J, Wareham NJ, Sharp SJ, O'Rahilly S, Balkau B, et al. Correlation of the leptin:adiponectin ratio with measures of insulin resistance in non-diabetic individuals. Diabetologia. 2009;52(11):2345-9.
- Stern SE, Williams K, Ferrannini E, DeFronzo RA, Bogardus C, Stern MP. Identification of individuals with insulin resistance using routine clinical measurements. Diabetes. 2005;54(2):333-9.
- Geloneze B, Tambascia MA. [Laboratorial evaluation and diagnosis of insulin resistance]. Arq Bras Endocrinol Metabol. 2006;50(2):208-15.