

IVUS-based imaging modalities for tissue characterization: similarities and differences

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Abstract Gray-scale intravascular ultrasound (IVUS) is the modality that has been established as the golden standard for in vivo imaging of the vessel wall of the coronary arteries. The use of IVUS in clinical practice is an important diagnostic tool used for quantitative assessment of coronary artery disease. This has made IVUS the de-facto invasive imaging method to evaluate new interventional therapies such as new stent designs and for atherosclerosis progression-regression studies. However, the gray-scale representation of the coronary vessel wall and plaque morphology in combination with the limited resolution of the current IVUS catheters makes it difficult, if not impossible, to identify qualitatively (e.g. visually) the plaque morphology similar as that of histopathology, the golden standard to characterize and quantify coronary plaque tissue components. Meanwhile, this limitation has been partially overcome by new innovative IVUS-based post-processing methods such as: virtual histology IVUS (VH-IVUS, Volcano Therapeutics, Rancho Cordova, CA, USA), iMAP-IVUS (Bostoc Scientific, Santa Clara, CA, USA), Integrated Backscatter IVUS

(IB-IVUS) and Automated Differential Echogenicity (ADE).

Keywords Intravascular ultrasound · Radiofrequency data analysis · Atherosclerosis · Tissue characterization

Introduction

Atherogenesis is the leading cause of cardiovascular mortality and morbidity in the developed world. The imaging of coronary atherosclerosis and more in particular the high risk atheromatous plaque has made an explosive progress the last decade with the advent of innovative techniques that focus on the high-resolution visualization of the coronary vascular wall. Gray-scale intravascular ultrasound (IVUS) is the modality that has been established as the golden standard for in vivo imaging of the vessel wall of the coronary arteries [1, 2]. The use of IVUS is an important diagnostic tool used for quantitative assessment of coronary artery disease. This has made IVUS the de-facto invasive imaging method of choice to evaluate new interventional therapies such as new stent designs [3–5] and for atherosclerosis progression-regression studies [6–14]. However, the gray-scale representation of the coronary vessel wall and plaque morphology in combination with the limited

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resolution of the current IVUS catheters makes it difficult, if not impossible, to identify qualitatively (e.g. visually) the plaque morphology similar as that of histopathology, the golden standard to characterize and quantify coronary plaque tissue components. Meanwhile, this limitation has been partially overcome by new innovative IVUS-based post-processing methods such as: virtual histology IVUS [15, 16] (VH-IVUS, Volcano Therapeutics, Rancho Cordova, CA, USA), iMAP-IVUS [17] (Bostoc Scientific, Santa Clara, CA, USA), Integrated Backscatter IVUS [18] (IB-IVUS) and Automated Differential Echogenicity [19] (ADE).

Intravascular ultrasound (IVUS) principles

The size of current IVUS catheters are ranging from 2.6 to 3.5 French (0.87–1.17 mm) and are inserted into the coronary arteries through 6-French guiding catheters. The principle of IVUS imaging is based on the oscillatory movement (expansion and contraction) of a piezoelectric transducer (crystal) in order to produce sound waves when electrically excited. There are two major different transducer designs [2]: (1) the mechanical single element rotating device and (2) the electronic phased array. The mechanical rotating element device uses a single piezoelectric transducer that rotates with 1,800 rotations per minute, while the electronic phased array device uses multiple stationary placed piezoelectric transducers which are sequentially activated. The generated sound waves by the transducers propagates through the different tissues and is reflected according to the acoustic properties of the tissue it travels through [20].

Gray-scale IVUS based atheromatous plaque classification is limited due to its low spatial resolution and for the usual IVUS transducers (20 and 40 MHz) for which the axial resolution is 200 μm and the lateral 200–250 μm . Based on their visual appearance, not necessarily histological composition, atheromas have been classified in four categories by gray-scale IVUS: (1) soft plaque (lesion echogenicity less than the surrounding adventitia), (2) fibrous plaque (intermediate echogenicity between soft (echolucent) atheromas and highly echogenic calcified plaques), (3) calcified plaque (echogenicity higher than the adventitia with acoustic shadowing),

and (4) mixed plaques (no single acoustical subtype represents >80% of the plaques) [2].

IVUS based imaging modalities for tissue characterization

To overcome the limitations of qualitative visual interpretation of the IVUS images and to describe the coronary plaque morphology, several post-processing methods for computer-assisted quantification have been developed during the recent years. There are two basic different approaches: (1) Signal-based analysis (the so called raw radiofrequency analysis or RF-analysis) and (2) Image-based analysis. The different methods will be described below:

Tissue characterization using virtual histology IVUS (VH-IVUS)

The first commercial available RF-signal based tissue composition analysis tool was the so-called virtual histology (VH-IVUS, Volcano Therapeutics) software. It uses in-depth analysis of the backscattered RF-signal in order to provide a more detailed description of the atheromatous plaque composition and is performed with either a 20 MHz, 2.9F phased-array transducer catheter (Eagle EyeTM Gold, Volcano Therapeutics) or 45 MHz 3.2F rotational catheter (Revolution[®], Volcano Therapeutics) that acquires IVUS data electrocardiogram gated [21]. The main principle of this technique is that it uses not only the envelope amplitude of the reflected RF-signals (as gray-scale IVUS does), but uses also the underlying frequency content to analyze the tissue components present in coronary plaques. This combined information is processed using autoregressive models and thereafter in a classification tree that determines four basic plaque tissue components [15]: (1) Fibrous tissue (dark green), (2) Fibrofatty tissue (light green), (3) Necrotic core (red) and (4) dense calcium (white). The current software version assumes the presence of a media layer, which is artificially added, positioned just at the inside of the outer vessel contour. This technique has been compared in several studies against histology in humans and other species, see Table 1 for further details.

Table 1 Validation studies of IVUS and IVUS based imaging modalities

Author	Study settings	Year	Primary objective	Results
<i>Greyscale IVUS</i>				
Palmer [24]	In vitro	1999	IVUS for coronary atheromatous lesions compared to histology. Atheromatous plaque was classified as echodense, echolucent, heterogeneous or calcified by each observer and by one observer on separate occasions	Overall inter- and intra-observer reproducibility for plaque-type (Kappa 0.87[0.80–0.94] and 0.89[0.85–0.93 respectively]) and focal calcification (0.78[0.74–0.82] and 0.88[0.84–0.92]) was high Agreement for overall plaque type between intravascular ultrasound and histology occurred in 89% of sites (Kappa 0.73[0.69–0.77]). Specificity $\geq 90\%$ Lipid pools were observed by histology in 30 sections (25%). IVUS revealed the presence of lipid pools in 19 of these sections (16%; sensitivity 65%). Specificity $\geq 95\%$
Prati [25]	In vitro	2001	IVUS, high frequency transducer(40 MHz) for plaque composition compared to histomorphology Lipid/necrotic areas were defined by IVUS as large echolucent intraplaque areas surrounded by tissue with higher echodensity	Lipid pools were observed by histology in 30 sections (25%). IVUS revealed the presence of lipid pools in 19 of these sections (16%; sensitivity 65%). Specificity $\geq 95\%$
<i>VH IVUS</i>				
Nair [15]	Ex vivo	2002	Coronary plaque classification with intravascular ultrasound radiofrequency data analysis	Autoregressive classification schemes performed better than those from classic Fourier spectra with accuracies of 90.4% for fibrous, 92.8% for fibrolipidic, 90.9% for calcified, and 89.5% for calcified-necrotic regions in the training data set and 79.7, 81.2, 92.8, and 85.5% in the test data, respectively Predictive accuracy from all patients data: 87.1% for fibrous, 87.1% for fibro-fatty, 88.3% for necrotic core, and 96.5% for dense calcium regions, respectively Sensitivities: NC:67.3%, FT:86%, FF:79.3%, DC:50%. Specificities: NC:92.9%, FT:90.5%, FF:100%, DC:99%
Nasu [26]	In vivo	2006	Accuracy of in vivo coronary plaque morphology assessment: a validation study of in vivo virtual histology compared with in vitro histopathology	The overall predictive accuracies were 93.5% for FT, 94.1% for FF, 95.8% for NC, and 96.7% for DC Sensitivities: NC:91.7%, FT:95.7%, FF:72.3%, DC:86.5%. Specificities: NC:96.6%, FT:90.9%, FF:97.9%, DC:98.9%
Nair [16]	Ex vivo	2007	Automated coronary plaque characterisation with intravascular ultrasound backscatter: ex vivo validation	The overall predictive accuracies were 93.5% for FT, 94.1% for FF, 95.8% for NC, and 96.7% for DC Sensitivities: NC:91.7%, FT:95.7%, FF:72.3%, DC:86.5%. Specificities: NC:96.6%, FT:90.9%, FF:97.9%, DC:98.9%

Table 1 continued

Author	Study settings	Year	Primary objective	Results
Granada [27]	Ex vivo	2007	In vivo plaque characterization using intravascular ultrasound-virtual histology in a porcine model of complex coronary lesions	Compared with histology, IVUS-VH correctly identified the presence of fibrous, fibro-fatty, and necrotic tissue in 58.33, 38.33, and 38.33% of lesions, respectively Sensitivities: fibrous 76.1%, fibro-fatty 46%, and necrotic core 41.1%
Van Herk [28]	Ex vivo	2009	Validation of in vivo plaque characterisation by virtual histology in a rabbit model of atherosclerosis	VH-IVUS had a high sensitivity, specificity and positive predictive value for the detection of non-calcified thin cap fibroatheroma (88, 96, 87%, respectively) and calcified thin cap fibroatheroma (95, 99, 93%, respectively). These values were respectively 82, 94, 85% for non-calcified fibroatheroma and 78, 98, 84% for calcified fibroatheroma. The lowest values were obtained for pathological intimal thickening (74, 92, 70%, respectively). For all plaque types, VH-IVUS had a kappa-value of 0.79
Thim [29]	Ex vivo	2010	Unreliable assessment of necrotic core by VHTM IVUS in porcine coronary artery disease	No correlations were found between the size of the necrotic core determined by VH IVUS and histology. VH IVUS displayed necrotic cores in lesions lacking cores by histology
<i>IB IVUS</i>				
Kawasaki [22]	In vivo	2002	In vivo quantitative tissue characterization of human coronary arterial plaques by use of integrated backscatter intravascular ultrasound and comparison with angioscopic findings	r:0.954 for each category, DC, FF, FT, NC
Kawasaki [23]	In vivo	2006	Diagnostic accuracy of optical coherence tomography and integrated backscatter intravascular ultrasound images for tissue characterization of human coronary plaques	Sensitivities: DC:100% FT:94% Lipid pool:84% Specificities: DC:99% FT:84% Lipid pool:97%
Okubo [18]	Ex vivo	2008	Development of integrated backscatter intravascular ultrasound for tissue characterization of coronary plaques	IB classified fibrous, lipid-rich and fibrocalcific plaque components with a high accuracy of 93, 90 and 96%, respectively
<i>iMAP</i>				
Sathyanarayana [17]	In vivo	2009	Characterisation of atherosclerotic plaque by spectral similarity of radiofrequency intravascular ultrasound signals	Ex vivo validation demonstrated accuracies at the highest level of confidence as: 97, 98, 95, and 98% for necrotic, lipidic, fibrotic and calcified regions respectively

Table 1 continued

Author	Study settings	Year	Primary objective	Results
<i>Echogenicity</i> Bruining [19]	Ex vivo	2007	Three-dimensional and quantitative analysis of atherosclerotic plaque composition by automated differential echogenicity	Areas of hypoechogenicity correlated with the presence of smooth muscle cells. Areas of hyperechogenicity correlated with presence of collagen, and areas of hyperechogenicity with acoustic shadowing correlated with calcium

Tissue characterization using i-MAP-IVUS

Recently, another RF-based processing method has become commercially available for coronary plaque tissue characterization which is called iMAP-IVUS (Boston Scientific) [17]. In principle this software is comparable, from methodological point of view, in brief IVUS-RF parameters are processed using auto-regressive models and matched to a database of known RF-signal profiles containing the characteristics of four tissue types. The colour-coded images contain the different colors overlaying the gray-scale IVUS images. However, there are differences, to start with the applied colour scheme: (1) Fibrous tissue (light green), (2) Lipid tissue (yellow), (3) Necrotic core (pink) and (4) Calcium (blue) [17]. Furthermore, the applied IVUS catheter is a 40 MHz rotating single element catheter. Ex vivo validation demonstrated accuracies at the highest level of confidence as: 97%, 98%, 95%, and 98% for necrotic, lipidic, fibrotic and calcified regions respectively (Table 2) [17].

Tissue characterization by integrated backscatter IVUS analysis (IB-IVUS)

IB-IVUS analysis is an alternative approach, as compared to the 2 previous ones, using the RF-signals of the IVUS catheters to characterize coronary plaque tissue components. IB-IVUS analyses the RF-signals generated by the 40 MHz mechanically rotating IVUS catheters by applying a fast Fourier transformation of the frequency components of the backscattered signals calculating the intensity of the signal measured in decibels (dB). Different tissue components reflect the RF-signals at different power levels, which, according to the developers, could be used to differentiate various tissue components. Analogue to the other methods it also applies a colour coded overlay onto the gray-scale IVUS images. It comprises the following different tissue components: (1) Calcification, (2) Fibrous tissue, and (3) Lipidic [18, 22, 23].

Tissue characterization using, automated differential echogenicity (ADE)

Automated differential echogenicity is a post-processing imaging analysis method that uses IVUS

Table 2 Similarities and differences of IVUS and IVUS-based imaging modalities

	Gray-scale IVUS	VH	i-MAP	Integrated backscatter	Echogenicity
Type of device	Mecanical and electrical	Mechanical and electrical	Mechanical	Mechanical	As IVUS
Transducer frequency	20–40 MHz	20–45 MHz	40 MHz	40 MHz	As IVUS
Colour code	Gray-scale	Fibrous: green Necrotic core: red Calcium: white Fibrofatty: light green	Fibrous: light green Necrotic core: pink Calcium: blue Fibrofatty: yellow	Fibrous: green Necrotic core: blue Calcium: red Fibrofatty: yellow	Gray-scale
Backscatter radiofrequency signal analysis	Amplitude (dB)	Autoregressive model	Fast Fourier transformation	Fast Fourier transformation	Computer analysis

images to characterize atheromatous coronary plaque components [19]. At variance with the previous described RF-signal analysis methods, it makes use of the differences in gray-value intensities within the gray-scale IVUS images which are caused by the different acoustic properties (e.g. echogenicity) of the different tissue components. It is known that adventitia tissue contains mostly fibrotic tissue components. Using the relative gray-value of the adventitia layer surrounding the external elastic membrane (EEM), which within quantitative IVUS analysis is detected and defined as the outer vessel wall, tissue components can be differentiated as hypo- (lower gray-values than the adventitia) or hyperechogenic (higher gray-values than the adventitia). Bright structures (very high gray-levels) combined with acoustic shadowing can be identified as calcium. The method classifies structures within these acoustic shadowed areas as unknown as the ultrasound signal is mostly completely reflected by thick layers of calcium and is not able to penetrate it (Figs. 1 and 2). The method has not been developed, such as the other methods, to generate colour-coded IVUS images that try to mimic as close as possible images as generated by histopathology, but to quantify differences over time to evaluate possible changes of the plaque components induced by new therapeutic treatments. It is of outmost importance, in order to better define the images, to follow a certain algorithm that includes: (1) accurate plaque identification (e.g. quantitative IVUS), (2) tunica adventitia definition and grey-level intensity determination (3) normal distribution of the gray-level intensity histogram of the adventitia tissue (image quality control), (4)

acoustic shadowing and calcification identification [19]. The advantages of this method are that it can be performed retrospectively on already acquired IVUS data and it is independent of the applied IVUS systems and/or catheters. However, it is recommended to apply within a longitudinal study the same console type and catheter during baseline and follow-up examinations not only within this method but also with respect to the other methods.

Discussion

As above described, IVUS has become over the past 20 years an important clinical intracoronary imaging tool. It not only facilitates clinical practice but it is also a reference method of which its quantitative parameters are often used as endpoints in first-in-man studies and larger clinical trials. However, the limited resolution, as compared to histology, and gray-scale (256 shades of gray are used when the image is optimal) representation, for which we humans have only limited capabilities with our eyes to make a distinction between them (on average we can only discriminate 8–12 different gray-levels) combined with the large amount of images acquired during a pullback examination (in an IVUS study of 4 cm there are 2,400 individual cross-sections acquired), requires another representation of the information we are looking for. Of course quantification is mandatory for research purposes and thus automated or semi-automated computer-assisted methods are necessary. In addition, humans can interpret colour-information much better than gray-scale information alone and

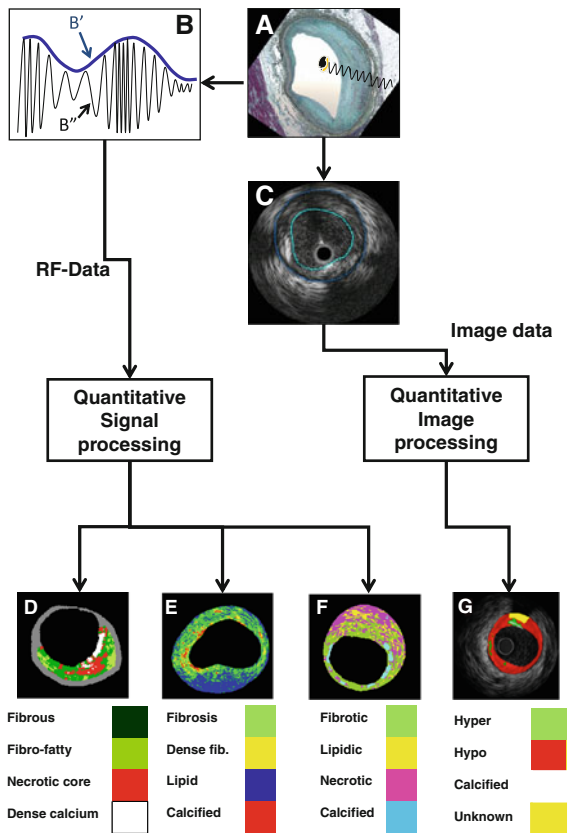


Fig. 1 A shows how an intravascular ultrasound signal is obtained from the vessel wall within an histology image. The *greyscale* IVUS image, as can be appreciated in C, is formed by the envelope (amplitude) (B') of the radiofrequency signal, which is illustrated in B. By *greyscale* atherosclerotic plaque can be classified into 3 categories: hypoechogenic, hyperechogenic and calcified. A 4th category is unknown, which is tissue that is acoustically showed (G). C shows a cross-sectional view of a grayscale image. The *blue lines* limit the actual atheroma. The frequency and power of the signal commonly differ between tissues, regardless of similarities in the amplitude. From the backscatter radiofrequency data (identified by B'' in B) different types of tissue information can be retrieved: virtual histology (D), integrated backscattered (IB) IVUS (E) and iMAP (F). Virtual histology is able to detect four tissue types: necrotic core, fibrous, fibro-fatty and dense calcium. The tissues characterized by integrated backscattered (IB) IVUS are lipidic, fibrous and calcified; and iMAP detects fibrotic, lipidic, necrotic and calcified

therefore the plaque compositional tools as described in this overview are valuable additions to the use of standard IVUS gray-scale imaging alone. Until recently, there was only one tool (e.g. VH IVUS)

commercially and thus widely available. This has changed, now more methods are available which could potentially lead to different outcomes as they are using different mathematical methods and software algorithms. Certainly, in the near future the different methods will possibly be applied side-by-side in multi-center studies and it is yet unknown if this could result in possible different study outcomes, this needs to be explored in future research.

The name giving of virtual histology IVUS (VH-IVUS) could possibly lead to expectations that the results derived by this method are one-to-one comparable to histology, which is unfortunately not the case. The development of the RF-signal based methods have been performed empirically, in brief: explanted vessels are imaged by IVUS, the pathologist performs histology and the histology results are cross-correlated to the IVUS images. The derived and correlated RF-signal patterns have been used to build the signal pattern databases which are later used to identify the plaque tissue components *in vivo*. The crux in this development process is the cross-correlation between histology and IVUS, which due to the large differences in image resolution is difficult (by example the lateral resolution of IVUS is 200–250 μm while that for histology is 5 μm). Although histology is considered the golden standard it is also not free from possible artifacts, by example during the fixation and staining process. Interpretation of the images by different pathologists could also result in interobserver related-biases. It is therefore not realistic to expect similar results from the IVUS derived tissue compositional tools as from quantitative histology. However, if appropriately used, and with justified expectations, these quantitative plaque tissue compositional tools, with all of their limitations, can be of great additional value to investigate plaque compositional changes which cannot be performed with any other imaging method. Great care must thus be taken when trying to prove a hypothesis using these methods which could be out of range of the capabilities of the methods and the basic method, e.g. IVUS itself. An example of expectations which could lead to disappointing results when the RF-based methods are applied is within metallic stented segments. The RF-based methods do not have the signal profile of metallic stent struts in their tissue signal profile databases and this will thus result in an

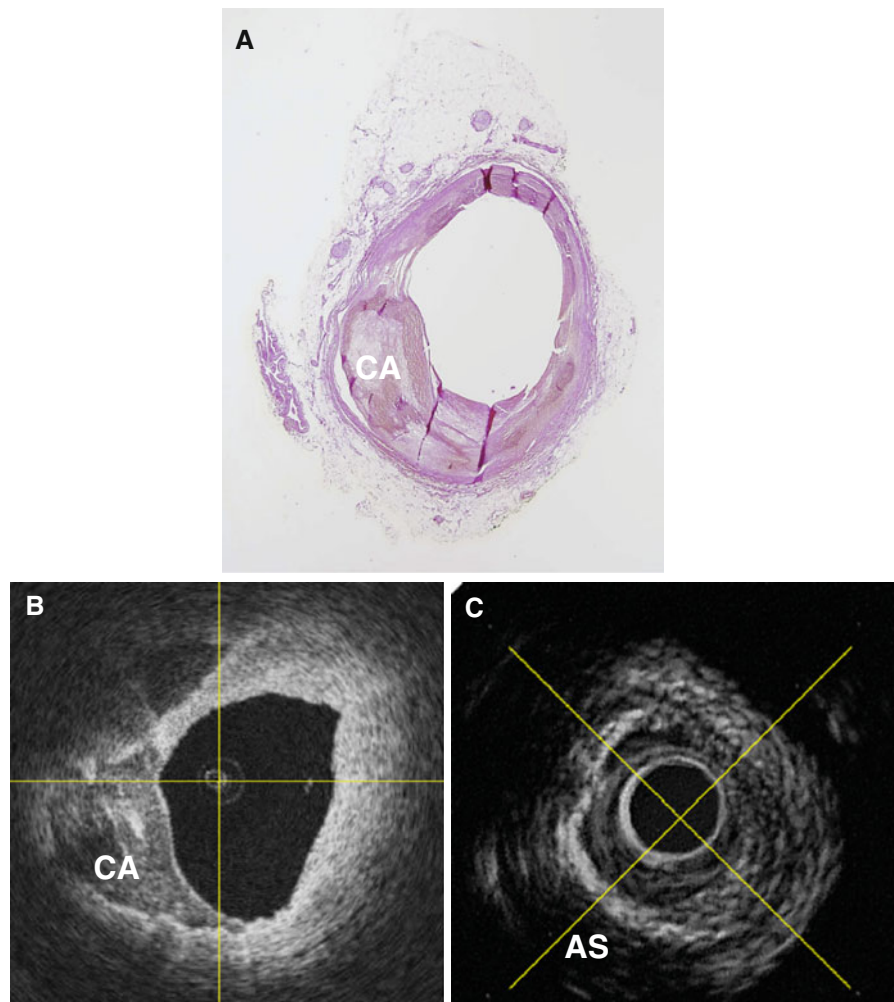


Fig. 2 One of the main limitations of processing the RAW radiofrequency data only, not taking into account other information available, such as visible acoustic shadowing, is that these signal processing methods identify tissue components in these acoustic *shadowed* regions while IVUS does not return any information about it. The ultrasound waves are reflected at the calcium interface as can be appreciated in this figure. **A** presents a histology sample of an explanted human coronary artery of which in **B** the OCT image is presented and in **C** the IVUS equivalent. It can be appreciated that OCT

presents the calcium very well between 6 and almost 12 o'clock. Also within the IVUS image it can be appreciated that between 6 and 12 o'clock there is a calcium layer present due to the *white* and bright interface but everything behind it is *shadowed* and thus IVUS does not contain any information deeper than this interface. When using IVUS as the only source to estimate the plaque composition tissue within these *shadowed* areas should therefore be classified as unknown tissue

artificial identification (in some research papers this is identified as misuse). This could later lead to unpredictable results and thus care must be taken applying these methods for research for which they are not designed.

However, encouraging results have been achieved applying the tissue composition methods to detect and to quantify as a surrogate the absorption of the

recently introduced bioabsorbable BVS stent (BVS, Abbott Vascular, Santa Clara, CA, USA). This information could help to explore into more detail the overall performance of this new stent platform.

A major limitation of the RF-based methods, as compared to the image-based echogenicity method, is that they do not take into account acoustic shadowed areas. They divide every individual RF-beam into

small regions of interest and compare the found signal profiles to their databases of known tissue profiles without taking into account what signal is found in front or behind that window (e.g. the larger picture is missing). This results in detection of tissue behind calcified areas, even if on the gray-scale IVUS image there is only shadow visible, thus the RF-signal will most likely contain noise (thick layers of calcium will reflect all acoustic energy back to the transducer causing the typical bright white appearance on gray-scale IVUS images) (Fig. 2). This could lead to two potential biases: (1) an observer related bias as the outer vessel boundaries are shadowed and thus the outer vessel contour must be interpolated by the observer and (2) a software related bias as the signal processing tools are not taking into account these shadowed areas and will assign the pixels in these regions to any of the tissue components within their database based on the “noise” of the ultrasound waves related to that area. In contrast, the image-based method of echogenicity at first examines every cross-sectional IVUS image if it contains acoustic shadowed areas and classifies tissue within these areas as unknown preventing possible software related deviations.

Although IVUS is proven to be safe and is a well-established method widely available, there are other promising imaging methods, such as optical coherence tomography, but for the time being they are not capable identifying quantitatively tissue components comparable to the described methods in this paper.

In-depth post-hoc analysis of IVUS data can be applied to quantify coronary plaque composition and possible changes of this composition over time which can be applied to evaluate new therapeutic treatment methods. However, there is still a large scientific debate how these different analysis methods exactly relate to each other and to that of the golden standard of histopathology. If these methods are also capable of identifying possible vulnerable segments is still under investigation.

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Conflict of interest None.

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