



# Biosynthesis, characterization and antimicrobial activity of gold nanoparticles from leaf extracts of *Annona muricata*

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## Abstract

The need to divulge the rapid synthesis, non-hazardous, cost effectiveness and eco-friendly methods for the synthesis of nanoparticles utilizing plants is of great importance. This is as a result of high toxicity associated with the chemical method of synthesizing nanoparticles. The aim of this study was to investigate the potency of the synthesized gold nanoparticle against selected human pathogens. Gold nanoparticles were synthesized by reacting 1 mM gold chloride solution with leaf extract of *Annona muricata*. The synthesized gold nanoparticles were characterized with UV–visible spectrophotometer, transmission electron microscope (TEM) and Fourier transformed infrared spectroscopy (FTIR). The antibacterial and antifungal activities of the synthesized gold nanoparticles were also investigated. The morphology, size, and structural properties of synthesized gold nanoparticles were determined with TEM analysis which showed spherical mono-dispersed structure with an average particle size of 25.5 nm. FTIR analysis reveal band at 3271.14, 2111.91 and 1637.82 cm<sup>-1</sup> corresponding to –N–H, –C=C, and –C–N functional groups that are responsible for the capping and stabilization of synthesized gold nanoparticles. The effectiveness of the gold nanoparticle against the test pathogens increases as the concentration of gold nanoparticle increases. The percentage of zones of inhibition of synthesized gold nanoparticle against test fungi and bacteria ranges from 30 to 66% and 40 to 54%, respectively. The potency of the synthesized gold nanoparticle against the selected fungi and bacteria increases with increase in concentration of gold nanoparticle. Therefore, the antibacterial and antifungal investigation revealed that the synthesized gold nanoparticles exhibited good antimicrobial activity.

**Keywords** Characterization · Antimicrobial · Gold nanoparticles · *Annona muricata*

## Introduction

In modern materials sciences nanotechnology is one of the imperative approach used to detect nano-substance which has found numerous applications in various fields such as pharmaceutical and biomedical sciences [1]. Previously, nanoparticles were synthesized by chemical method where chemicals act as the reducing agents. But the toxicity and probable risk on human health and environment are major limitations to this approach, the biological methods which use plants is more preferable because of its nontoxicity, tranquil scaling-up, and reproducibility in making [2, 3]. The greener method of phytomediated synthesis of gold nanoparticles using dalspinin at room temperature has been reported not to toxic [4]. The development of an eco-friendly procedure for the synthesis of nanoparticle via the biological method using plants arises as a result of the hazardous effect of the chemical and physical methods of synthesizing

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nanoparticles. The bioactive compounds present in plants act as reducing agents which helps in the reduction of metal ions to metal nanoparticles with distinct size, shapes and significant antimicrobial efficiency [5]. The antimicrobial potency of nanoparticles and its small size motivate researchers in evaluating different methods for the synthesis of highly mono-dispersed nanoparticles with verified antimicrobial activities [6]. The antimicrobial properties of the nanoparticles synthesized from *Annona muricata* extract showed a great inhibition against Gram-positive and Gram-negative bacteria, the growing resistant strains developed by microbes against metal ions and antibiotics coupled with the good antibacterial properties of gold nanoparticles are the major interest of researchers in green synthesis of gold nanoparticles [7, 8]. Among other metal nanoparticles gold nanoparticle exhibits effective antimicrobial property due to its tremendously large surface area, which provides better contact with microorganisms [9]. The high chemical stability, optical properties, oxidation resistance and biocompatibility properties of gold nanoparticles and their applications in different field of study such as biology, optics and medicine make synthesis of gold nanoparticles an interesting object of study [10]. The advantage of synthesizing gold nanoparticles from plant extract is not limited to reduction of environmental toxicity, the production of nanoparticles in large quantities, simplicity of reducing metal salts, rapid, cost effective, and safety for clinical research to nanoparticles are other advantages associated with this method of synthesis, the green synthesized gold nanoparticle can be used in development of catalysts, sensors, nano medicine and biomarker applications [11]. Gold nanoparticles exhibit properties that are different from that of bulk gold and can be manufactured into various shapes such as nanorods, nanostars, nanobelts, nanospheres, nanocages and nanoprisms [12]. The size, high surface area, and shapes of gold nanoparticles have influenced its chemical properties and have increased its applications especially in the field of medicine [13]. From traditional age to modern age entire part of *A. muricata* plant have been reported to provide vast medicinal benefits to human by curing ailments such as bacteria, fungi, cancer, tumor, arthritic, diabetics, inflammation, sedative, diarrheal and diuretic infections

[14]. This study was aimed at the investigation of stable and efficient process for the synthesis of gold nanoparticle using *A. muricata* leaf via the characterization and pharmacological activities evaluation.

## Materials and methods

### Preparation of leaf extract

The leaves of *A. muricata* were collected from the premises of botany department of Obafemi Awolowo University, Ile-Ife, Nigeria and were authenticated. The leaves were properly washed, air dried and pulverized with blender grinder. 20 g of the pulverized leaves sample of *A. muricata* was transferred into a 200 ml Erlenmeyer flask, 150 ml of de-ionized water was added and boiled for 20 min. The boiled plant extract was cooled at room temperature and filtered with (Whatman filter paper No. 1). This extract was further used for the synthesis of gold nanoparticles. The fresh, air-dried and pulverized leaves of *A. muricata* are showed in Fig. 1.

### Biosynthesis of gold nanoparticles

The method of [15] was adopted in the synthesis of the gold nanoparticles. 1 ml of the obtained leaf extract was added to 1 mM solution of gold chloride and stirred properly at room temperature until the yellow color of the extract changed to purple color that confirmed the synthesis of gold nanoparticles. The mixture was incubated for 22 h for thorough bio-reduction of gold chloride to gold nanoparticles from leaf extract. The reduction of gold chloride to gold nanoparticles was monitored with UV–Vis spectrometer by recording the UV–Vis absorption spectrum as a function of time.

### UV–visible spectroscopy

UV–Vis spectrometer (UV-245 Shimadzu) was used at various time intervals to ascertain the optimum concentration of plant extracts. The purple color confirmed the formation of the gold nanoparticle. The formation of the



Fig. 1 Fresh, air-dried and pulverized leaves sample of *Annona muricata*



gold nanoparticle was confirmed by scanning the absorption maxima of freshly prepared sample of gold nanoparticle using 1 cm path length quartz cuvettes at temperature range of 24–28 °C at wavelength 450–850 nm.

### FTIR spectroscopy

The FTIR analysis was determined with FTIR spectrometer, Nicolet iS50 (Thermo Fisher Scientific, Waltham, MA, USA). The freeze-dried gold nanoparticle extract was mixed with KBR (FTIR grade) and scanned on FTIR over the range of 4000–380  $\text{cm}^{-1}$  at a resolution of 4  $\text{cm}^{-1}$ . FTIR analysis was used for the identification of functional group present and to study the interactions of the functional groups as a source of reducing agents on the surface of synthesized nanoparticles.

### Transmission electron microscopy

The particle size and surface morphology of synthesized gold nanoparticle were measured with transmission electron microscopy (TEM) JEOL model 1200EX, at an accelerating voltage of 80 kilo voltage. The synthesized gold nanoparticle samples were prepared by dropping few drops of the gold nanoparticle solution on Lacey carbon grids, 300 mesh and allowed the grid to dry before measurement.

### Antibacterial investigation of gold nanoparticles

The investigation of the antibacterial potency of the synthesized gold nanoparticle against some clinical isolates bacteria was conducted following the standard well diffusion procedures [16]. Freshly cultures prepared overnight were spread on sterilized nutrient agar plates. Sterile cork borer was used to bore 8 mm diameter wells in the agar plates. The wells were loaded with 50  $\mu\text{l}$  of 2 mg/l and 4 mg/l concentration of gold nanoparticle solution and 2 mg/l of an antibiotic (Streptomycin) was used as positive control. The incubation of the plates at 37 °C for 28 h was done. The zones of inhibition around the well impregnated with gold nanoparticle were measured and the percentage zones of inhibition were calculated to determine the antibacterial activity of the synthesized gold nanoparticle. The antibacterial investigation was on triplicate analysis. The formula below was used for the calculation of the percentage growth inhibition.

$$\text{PGI} = (\text{BDC} - \text{BDT})/\text{BDC} \times 100,$$

PGI = Percent growth inhibition,

BDC = Bacteria colony diameter in control,

BDT = Bacteria colony diameter in treatment.

### Antifungal investigation of gold nanoparticles

The antifungal effectiveness of the synthesized gold nanoparticle from *A. muricata* was investigated following the procedures described by [17]. Sterilized SDA was transferred into sterile Petri dish. After media setting, sterile gel puncture was used to puncture 8 mm diameter wells in the agar plates used. Forty microliters of 2 mg/l and 4 mg/l concentrations of the gold nanoparticle were introduced into the wells. Each well containing the fungal discs was inoculated in inverted position. The plates were then incubated at 28 °C for 70–94 h. Amphotericin B was used as control. The diameter of the fungal colony was measured after incubation at 28 °C and the percentage growth inhibition was determined relatively to the fungal diameter in control. The antifungal investigation was carried out on triplicate analysis. The formula below was used for the calculation of the percentage growth inhibition:

$$\text{PGI} = (\text{FDC} - \text{FDT})/\text{FDC} \times 100,$$

PGI = Percent growth inhibition,

FDC = Fungal colony diameter in control,

FDT = fungal colony diameter in treatment.

## Results and discussion

### UV analysis of gold nanoparticles

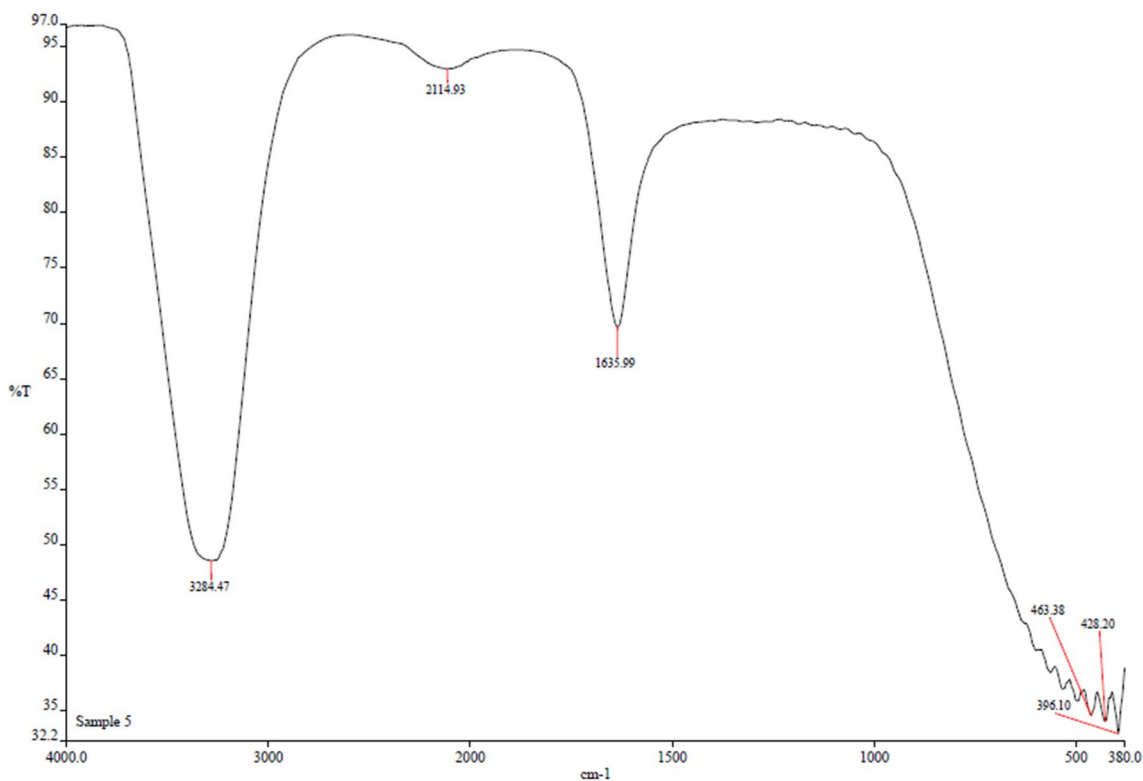
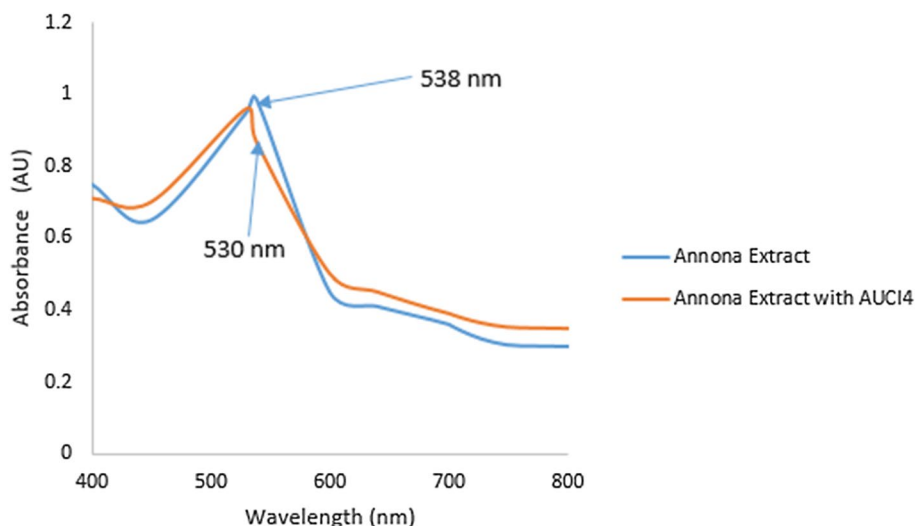
The purple color that appeared after mixing the leaf extract of *A. muricata* with gold chloride solution confirmed the formation of gold nanoparticles. The resulting colloidal solutions and *A. muricata* extract were scanned with UV–Vis spectroscopy and the spectrum showed in Fig. 2 was obtained. The occurrence of the peak at wavelength 530 and 538 nm corresponds to gold absorption peaks. The variation in the absorbance–wavelength of *A. muricata* extract and the colloidal solutions with gold chloride confirmed the synthesis of gold nanoparticles from the leaf of *A. muricata*.

### FTIR analysis of gold nanoparticles

FTIR analysis was carried out to determine the possible biomolecules in leaf extract of *A. muricata* that was responsible for capping that lead to the efficient stabilization of the gold nanoparticles. The bands observed at 3284.47, 2114.93, 1635.99  $\text{cm}^{-1}$  on the FTIR spectrum of *A. muricata* extract as shown in (Fig. 3) and bands at 3271.14, 2111.91 and 1637.82  $\text{cm}^{-1}$  on the FTIR spectrum of the synthesized gold nanoparticles from *A. muricata* (Fig. 4). The sharp bands at wavelength 3271.14  $\text{cm}^{-1}$  corresponding to –N–H, –C–N and –C=C– functional groups. These functional groups are also present in the studies [18, 19]. The variation in



**Fig. 2** UV spectrum before and after reduction of gold nanoparticles



**Fig. 3** FTIR spectrum of *Annona muricata* extract

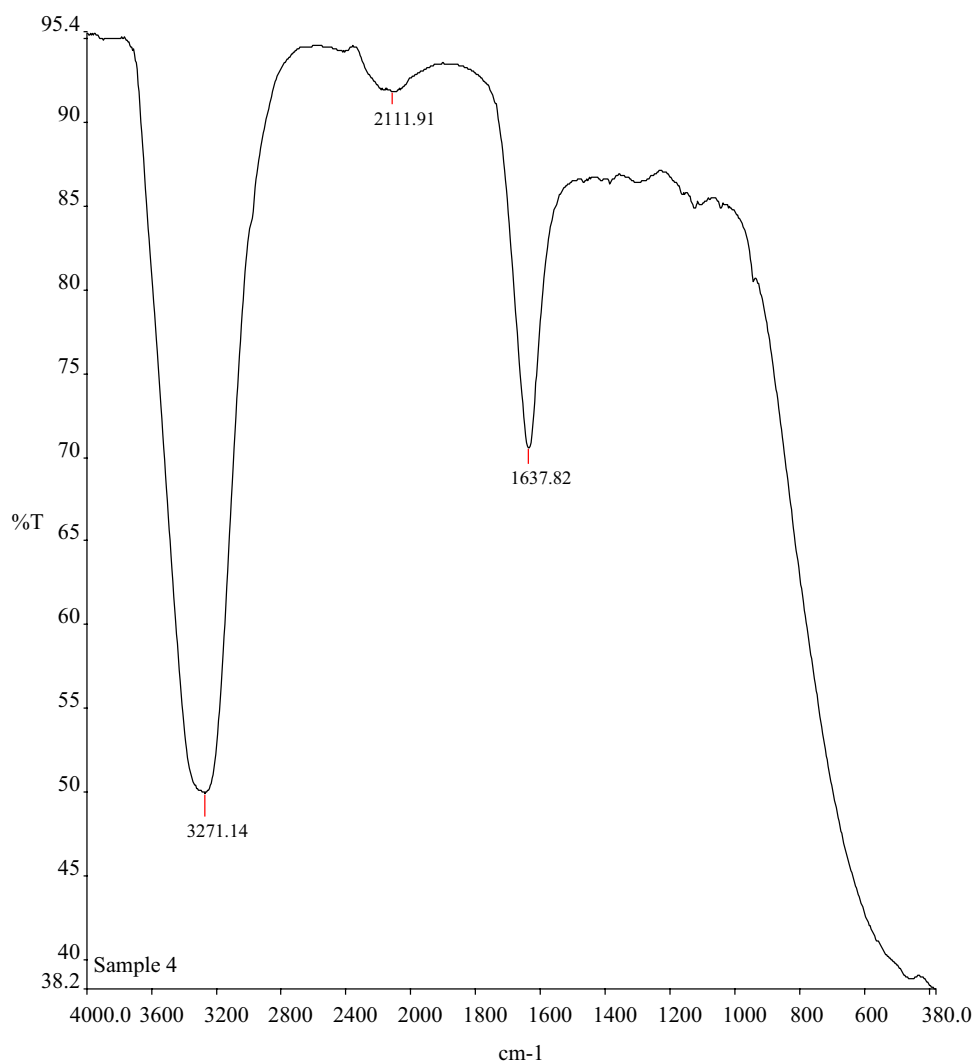
the absorption bands observed on both spectra of the FTIR might be responsible for the reduction of gold ions, formation and stabilization of the synthesized nanoparticles from *A. muricata* leaves. The vibrational bands corresponding to bonds such as  $-N-H$ ,  $-C=C-$ , and  $-C-N$  had being reported to be peculiar with flavonoids, terpenoids and protein compound [20]. This could signify that the extract has flavonoids, terpenoids and protein base compounds.

### TEM analysis of gold nanoparticles

Transmission electron microscope was used to determine the size, shape and distribution of synthesized gold nanoparticles. TEM micrograph of the synthesized gold nanoparticles from *A. muricata* leaves shows the presence of spherical mono-dispersed nanoparticles with particle size ranging from 27 to 32 nm with average particle size of



**Fig. 4** FTIR spectrum of the synthesized gold nanoparticles from *Annona muricata*



25.5 nm as shown in Fig. 5. This finding is in line with the result obtained by [20] on the TEM characterization of gold nanoparticle synthesized from *Periploca aphylla* plant. The particle size distribution in this study is lower than that obtained by [21] but found higher than that obtained from [22] findings.

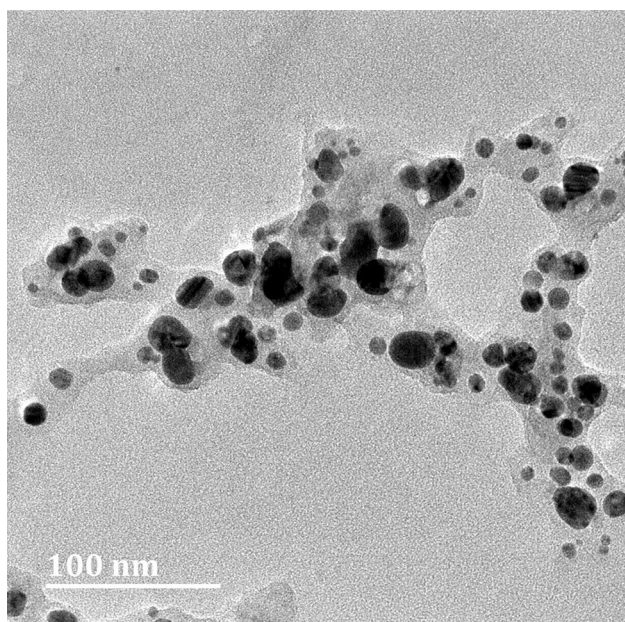
### Antibacterial investigation of gold nanoparticles

The antibacterial activities of synthesized gold nanoparticles from *A. muricata* leaf extract against selected bacterial showed in Table 1. It was obvious from the obtained data that the synthesized gold nanoparticles were toxic against all selected bacteria at different concentrations. *Clostridium sporogenes* was the most susceptible bacterium to AuNps at both concentrations of 2 mg/l and 4 mg/l with maximum percentages of zones of inhibition of 50% and 54%, respectively, whereas *Staphylococcus aureus* at both concentrations and *Enterococcus faecalis* at 2 mg/l concentration

of synthesized gold nanoparticles were the least susceptible bacteria with 40% percentages of zones of inhibition as shown in Fig. 6. The efficacy of the gold nanoparticle against the test bacteria increases as the concentration of gold nanoparticle increases. The increasing order of potency of the synthesized gold nanoparticle against the selected pathogenic human bacteria at both concentration of gold nanoparticle was *Staphylococcus aureus* > *Enterococcus faecalis* > *Klebsiella pneumonia* > *Clostridium sporogenes*. Previous studies revealed that plant-mediated metal nanoparticles play a crucial role in drug delivery and can be used against many microbes because of the efficiency antimicrobial properties they possess [21–23]. This suggested that the synthesized gold nanoparticle has good efficacy against bacteria and could be used in treating bacteria-infected diseases.



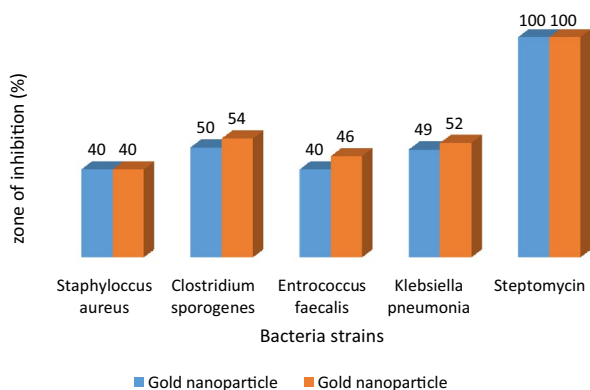




**Fig. 5** TEM micrograph of the synthesized gold nanoparticles from *Annona muricata*

**Table 1** Pathogenic bacteria and fungal used

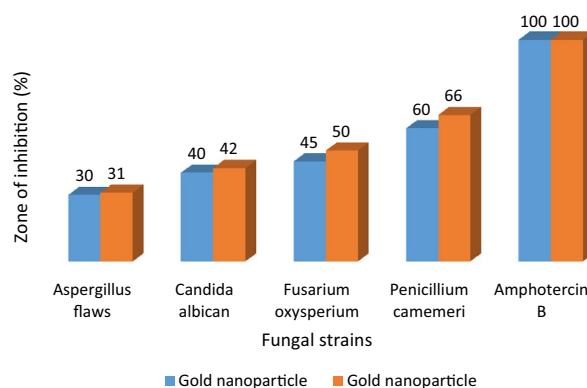
Bacteria	Fungi
<i>Staphylococcus aureus</i>	<i>Aspergillus flaws</i>
<i>Clostridium sporogenes</i>	<i>Candida albican</i>
<i>Entrococcus faecalis</i>	<i>Fusarium oxysperium</i>
<i>Klebsiella pneumonia</i>	<i>Penicillium camemeri</i>



**Fig. 6** Antibacterial activity of synthesized gold nanoparticle against selected bacteria strains

### Antifungal investigation of gold nanoparticles

Figure 7 shows the antifungal potency of synthesized gold nanoparticle against selected fungal species *Aspergillus flaws*, *Candida albican*, *Fusarium oxysperium* and



**Fig. 7** Antifungal activity of synthesized gold nanoparticle against selected fungal strains

*Penicillium camemeri* as shown in Table 1. Highest percentage of zone of inhibition was recorded against *Penicillium camemeri* (66%) at 4 mg/l gold nanoparticle while the lowest percentage of zone of inhibition was recorded against *Aspergillus flaws* (30%) at concentration of 2 mg/l gold nanoparticle. The efficacy of the gold nanoparticle against the test fungal increases as the concentration of gold nanoparticle increases. The increasing order of potency of the synthesized gold nanoparticle against the selected pathogenic human fungi was *Aspergillus flaws* > *Candida albican* > *Fusarium oxysperium* > *Penicillium camemeri*. The antifungal investigation of the synthesized gold nanoparticle from *A. muricata* leaves in this study was in line with previous study on antifungal activities of gold nanoparticle from different studied plants against different pathogenic human fungi where notable activity were observed [24, 25].

### Conclusion

The simplicity in synthesizing gold nanoparticle from *A. muricata* leaf extract offers a valuable contribution to green synthesis and nanotechnology because of its rapid synthesis, economic effectiveness, reproducible method and large amount of gold nanoparticles was attained without any toxic reagents been released to the environment. The synthesized gold nanoparticles from *A. muricata* leaf extract show broad inhibition against test bacterial and fungal there by making the synthesized gold nanoparticles a good antimicrobial agent. The information obtained from this study can be adopted by drug agencies in production of antibiotics drugs which will help in combating microbes that have developed resistance against existing antibiotic drugs.

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