

Table 1. Data and associated models available in KiMoSys repository.

Data EntryID	Organism	Type of data	Original Publication Title	Reference	Reference of associated model(s)
30	<i>Escherichia coli</i>	time-series data of metabolites	Dynamic modeling of the central carbon metabolism of <i>Escherichia coli</i> .	[1]	[1]
35	<i>Escherichia coli</i>	flux measurements	Multiple high-throughput analyses monitor the response of <i>E. coli</i> perturbations.	[2]	-
37	<i>Lactococcus lactis</i>	time-series data of metabolites	Is the glycolytic flux in <i>Lactococcus lactis</i> primarily controlled by the redox charge? Kinetics of NAD(+) and NADH pools determined <i>in vivo</i> by ¹³ C NMR.	[3]	[4,5]
38	<i>Escherichia coli</i>	time-series data of metabolites	Modeling and simulation of the main metabolism in <i>Escherichia coli</i> and its several single-gene knockout mutants with experimental verification	[6]	[6]
41	<i>Escherichia coli</i>	metabolites at steady-state	Multiple high-throughput analyses monitor the response of <i>E. coli</i> perturbations.	[2]	-
42	<i>Clostridium acetobutylicum</i>	time-series data of metabolites	A systems biology approach to investigate the effect of pH-induced gene regulation on solvent production by <i>Clostridium acetobutylicum</i> in continuous culture.	[7]	[7]
44	<i>Escherichia coli</i>	enzyme/protein concentrations	Multiple high-throughput analyses monitor the response of <i>E. coli</i> perturbations	[2]	-
51	<i>Escherichia coli</i>	time-series data of metabolites	Dynamic simulation of an <i>in vitro</i> multi-enzyme system.	[8]	-
52	<i>Escherichia coli</i>	metabolites at steady-state	Metabolic profiling of <i>Escherichia coli</i> cultivation: evaluation of extraction and metabolite analysis procedures.	[9]	-
54	<i>Escherichia coli</i>	metabolites at steady-state	Quantification of intracellular metabolites in <i>Escherichia coli</i> K12 using liquid chromatographic-electrospray ionization tandem mass spectrometric technique.	[10]	-
55	<i>Pichia pastoris</i>	metabolites at steady-state	Glucose-methanol co-utilization in <i>Pichia pastoris</i> studied by metabolomics and instationary ¹³ C flux analysis.	[11]	-
57	<i>Pichia pastoris</i>	flux measurements	Glucose-methanol co-utilization in <i>Pichia pastoris</i> studied by metabolomics and instationary ¹³ C flux analysis.	[11]	-
58	<i>Homo sapiens</i>	metabolites at steady-state	Dynamics and control of the central carbon metabolism in Hepatoma cells.	[12]	-

(continued from previous page)

Data EntryID	Organism	Type of data	Publication Title	Reference	Reference of associated model(s)
59	<i>Rattus</i>	time-series data of metabolites	The selective control of glycolysis, gluconeogenesis and glycogenesis by temporal insulin patterns.	[13]	-
61 and 62	<i>Saccharomyces cerevisiae</i>	time-series data of metabolites	Temporal system-level organization of the switch from glycolytic to gluconeogenic operation in yeast.	[14]	-
63	<i>Escherichia coli</i>	time-series data of metabolites	Metabolomics-driven quantitative analysis of ammonia assimilation in <i>E. coli</i> .	[15]	[15]
64	<i>Escherichia coli</i>	flux measurements	Impact of global transcriptional regulation by <i>arcA</i> , <i>arcB</i> , <i>cra</i> , <i>crp</i> , <i>cya</i> , <i>fnr</i> , and <i>mlc</i> on glucose catabolism in <i>Escherichia coli</i> .	[16]	-
65	<i>Escherichia coli</i>	flux measurements	Nonlinear dependency of intracellular fluxes on growth rate in miniaturized continuous cultures of <i>Escherichia coli</i> .	[17]	-
66	<i>Shewanella oneidensis</i>	flux measurements	Invariability of central metabolic flux distributions in <i>Shewanella oneidensis</i> MR-1 under environmental or genetic perturbation	[18]	-
67 and 68	<i>Escherichia coli</i>	flux measurements	An integrated computational and experimental study for overproducing fatty acids in <i>Escherichia coli</i> .	[19]	-
69 and 70	<i>Saccharomyces cerevisiae</i>	time-series data of metabolites	Characterization of glucose transport mutants of <i>Saccharomyces cerevisiae</i> during a nutritional upshift reveals a correlation between metabolite levels and glycolytic flux.	[20]	[20]
71	<i>Penicillium chrysogenum</i>	time-series data of metabolites	Generating short-term kinetic responses of primary metabolism of <i>Penicillium chrysogenum</i> through glucose perturbation in the bioscope mini reactor.	[21]	-
72	<i>Aspergillus niger</i>	flux measurements	Integration of in vivo and in silico metabolic fluxes for improvement of recombinant protein production.	[22]	-
73	<i>Aspergillus niger</i>	flux measurements	Overexpression of isocitrate lyase-glyoxylate bypass influence on metabolism in <i>Aspergillus niger</i> .	[23]	-
74	<i>Escherichia coli</i>	metabolites at steady-state	Catching prompt metabolite dynamics in <i>Escherichia coli</i> with the BioScope at oxygen rich conditions.	[24]	-
75	<i>Escherichia coli</i>	time-series data of metabolites	Changes in substrate availability in <i>Escherichia coli</i> lead to rapid metabolite, flux and growth rate response.	[25]	-
76	<i>Streptomyces coelicolor</i>	time-series data of metabolites	Intracellular metabolite pool changes in response to nutrient depletion induced metabolic switching in <i>Streptomyces coelicolor</i> .	[26]	-

(continued from previous page)

Data EntryID	Organism	Type of data	Publication Title	Reference	Reference of associated model(s)
77	<i>Shewanella oneidensis</i>	time-series data of metabolites	Integrating flux balance analysis into kinetic models to decipher the dynamic metabolism of <i>Shewanella oneidensis</i> .	[27]	[27]
78	<i>Escherichia coli</i>	time-series data of metabolites	Metabolic flux analysis in a nonstationary system: fed-batch fermentations of a high yielding strain of <i>E. coli</i> producing 1,3-propanediol.	[28]	-
79	<i>Escherichia coli</i>	metabolites at steady-state	Metabolic regulation analysis of wild-type and arcA mutant <i>Escherichia coli</i> under nitrate conditions using different levels of omics data.	[29]	-
80	<i>Escherichia coli</i>	flux measurements	Metabolic regulation analysis of wild-type and arcA mutant <i>Escherichia coli</i> under nitrate conditions using different levels of omics data.	[29]	-
81	<i>Mouse</i>	enzyme/protein concentrations	Division of labor by dual feedback regulators controls JAK2/STAT5 signaling over broad ligand range.	[30]	[30]
82	<i>Homo sapiens</i>	enzyme/protein concentrations	Network quantification of EGFR signaling unveils potential for targeted combination therapy.	[31]	-

Data EntryID: internal accession identifier.

Data EntryID without associated models is indicated by “-”.

Reference List

1. Chassagnole C, Noisommit-Rizzi N, Schmid JW, Mauch K, Reuss M: **Dynamic modeling of the central carbon metabolism of *Escherichia coli***. *Biotechnology and Bioengineering* 2002, **79**: 53-73.
2. Ishii N, et.al.: **Multiple high-throughput analyses monitor the response of *E. coli* to perturbations**. *Science* 2007, **316**: 593-597.
3. Neves AR, Ventura R, Mansour N, Shearman C, Gasson MJ, Maycock C *et al.*: **Is the glycolytic flux in *Lactococcus lactis* primarily controlled by the redox charge? Kinetics of NAD(+) and NADH pools determined in vivo by C-13 NMR**. *Journal of Biological Chemistry* 2002, **277**: 28088-28098.

4. Levering J, Musters MWJM, Bekker M, Bellomo D, Fiedler T, de Vos WM *et al.*: **Role of phosphate in the central metabolism of two lactic acid bacteria - a comparative systems biology approach.** *FEBS J* 2012, **279**: 1274-1290.
5. Vinga S, Neves AR, Santos H, Brandt BW, Kooijman SALM: **Subcellular metabolic organization in the context of dynamic energy budget and biochemical systems theories.** *Philosophical Transactions of the Royal Society B-Biological Sciences* 2010, **365**: 3429-3442.
6. Kadir TAA, Mannan AA, Kierzek AM, McFadden J, Shimizu K: **Modeling and simulation of the main metabolism in *Escherichia coli* and its several single-gene knockout mutants with experimental verification.** *Microbial Cell Factories* 2010, **9**: 88.
7. Haus S, Jabbari S, Millat T, Janssen H, Fischer RJ, Bahl H *et al.*: **A systems biology approach to investigate the effect of pH-induced gene regulation on solvent production by *Clostridium acetobutylicum* in continuous culture.** *Bmc Systems Biology* 2011, **5**.
8. Ishii N, Suga Y, Hagiya A, Watanabe H, Mori H, Yoshino M *et al.*: **Dynamic simulation of an *in vitro* multi-enzyme system.** *FEBS* 2007, **581**: 413-420.
9. Hiller J, Franco-Lara E, Weuster-Botz D: **Metabolic profiling of *Escherichia coli* cultivations: evaluation of extraction and metabolite analysis procedures.** *Biotechnology Letters* 2007, **29**: 1169-1178.
10. Buchholz A, Takors R, Wandrey C: **Quantification of intracellular metabolites in *E. coli* K12 using liquid chromatographic-electrospray ionization tandem mass spectrometric techniques.** *Analytical Biochemistry* 2001, **295**: 129-137.
11. Jorda J, Suarez C, Carnicer M, ten Pierick A, Heijnen JJ, van Gulik W *et al.*: **Glucose-methanol co-utilization in *Pichia pastoris* studied by metabolomics and instationary C-13 flux analysis.** *Bmc Systems Biology* 2013, **7**.
12. Maier K, Hofmann U, Reuss M, Mauch K: **Dynamics and control of the central carbon metabolism in hepatoma cells.** *BMC Systemes Biology* 2010, **4**: 54-82.
13. Noguchi R, Kubota H, Yugi K, Toyoshima Y, Komori Y, Soga T *et al.*: **The selective control of glycolysis, gluconeogenesis and glycogenesis by temporal insulin patterns.** *Molecular Systems Biology* 2013, **9**.
14. Zampar GG, Kummel A, Ewald J, Jol S, Niebel B, Picotti P *et al.*: **Temporal system-level organization of the switch from glycolytic to gluconeogenic operation in yeast.** *Molecular Systems Biology* 2013, **9**.

15. Yuan J, et al.: **Metabolomics-driven quantitative analysis of ammonia assimilation in *E. coli***. *Molecular Systems Biology* 2009, **5**.
16. Perrenoud A, Sauer U: **Impact of global transcriptional regulation by ArcA, ArcB, Cra, Crp, Cya, Fnr, and Mlc on glucose catabolism in *Escherichia coli***. *Journal of Bacteriology* 2005, **187**: 3171-3179.
17. Nanchen A, Schicker A, Sauer U: **Nonlinear dependency of intracellular fluxes on growth rate in miniaturized continuous cultures of *Escherichia coli***. *Applied and Environmental Microbiology* 2006, **72**: 1164-1172.
18. Tang YJ, Martin HG, Deutschbauer A, Huang R, Llorca X, Arkin A *et al.*: **Invariability of central metabolic flux distribution in *Shewanella oneidensis* MR-1 under environmental or genetic perturbations**. *Biotechnol Prog* 2009, **25**: 1254-1259.
19. Ranganathan S, Tee TW, Chowdhury A, Zomorodi AR, Yoon JM, Fu YF *et al.*: **An integrated computational and experimental study for overproducing fatty acids in *Escherichia coli***. *Metabolic Engineering* 2012, **14**: 687-704.
20. Bosch D, Johansson M, Ferndahl C, Franzen CJ, Larsson C, Gustafsson L: **Characterization of glucose transport mutants of *Saccharomyces cerevisiae* during a nutritional upshift reveals a correlation between metabolite levels and glycolytic flux**. *Fems Yeast Research* 2008, **8**: 10-25.
21. Nasution U, van Gulik WM, Proell A, van Winden WA, Heijnen JJ: **Generating short-term kinetic responses of primary metabolism of *Penicillium chrysogenum* through glucose perturbation in the bioscope mini reactor**. *Metabolic Engineering* 2006, **8**: 395-405.
22. Driouch H, Melzer G, Wittmann C: **Integration of in vivo and in silico metabolic fluxes for improvement of recombinant protein production**. *Metabolic Engineering* 2012, **14**: 47-58.
23. Meijer S, Otero J, Olivares R, Andersen MR, Olsson L, Nielsen J: **Overexpression of isocitrate lyase-glyoxylate bypass influence on metabolism in *Aspergillus niger***. *Metabolic Engineering* 2009, **11**: 107-116.
24. De Mey M, Taymaz-Nikerel H, Baart G, Waegeman H, Maertens J, Heijnen JJ *et al.*: **Catching prompt metabolite dynamics in *Escherichia coli* with the BioScope at oxygen rich conditions**. *Metabolic Engineering* 2010, **12**: 477-487.
25. Taymaz-Nikerel H, De Mey M, Baart G, Maertens J, Heijnen JJ, van Gulik W: **Changes in substrate availability in *Escherichia coli* lead to rapid metabolite, flux and growth rate responses**. *Metabolic Engineering* 2013, **16**: 115-129.

26. Wentzel A, Sletta H, Consortium S, Ellingsen TE, Bruheim P: **Intracellular metabolite pool changes in response to nutrient depletion induced metabolic switching in *Streptomyces coelicolor*.** *Metabolites* 2012, **2**: 178-194.
27. Feng X, Chen Y, Tang YJ: **Integrating flux balance analysis into kinetic models to decipher the dynamic metabolism of *Shewanella oneidensis* MR-1.** *PLOS Comput Biol* 2012, **8**: e1002376.
28. Antoniewicz MR, Kraynie DF, Laffend LA, Gonzalez-Lergier J, Kelleher JK, Stephanopoulos G: **Metabolic flux analysis in a nonstationary system: Fed-batch fermentation of a high yielding strain of *E. coli* producing 1,3-propanediol.** *Metabolic Engineering* 2007, **9**: 277-292.
29. Toya Y, Nakahigashi K, Tomita M, Shimizu K: **Metabolic regulation analysis of wild-type and *arcA* mutant *Escherichia coli* under nitrate conditions using different levels of omics data.** *Molecular Biosystems* 2012, **8**: 2593-2604.
30. Bachmann J, Raue A, Schilling M, Bohm ME, Kreutz C, Kaschek D *et al.*: **Division of labor by dual feedback regulators controls JAK2/STAT5 signaling over broad ligand range.** *Molecular Systems Biology* 2011, **7**.
31. Klinger B, Sieber A, Fritsche-Guenther R, Witzel F, Berry L, Schumacher D *et al.*: **Network quantification of EGFR signaling unveils potential for targeted combination therapy.** *Molecular Systems Biology* 2013, **9**.