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Permalink https://escholarship.org/uc/item/438355mn

Journal Magnetic Resonance in Medicine, 71(2)

ISSN 0740-3194

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Publication Date

2014-02-01

DOI

10.1002/mrm.24689

Peer reviewed

Time-Resolved Noncontrast Enhanced 4-D Dynamic Magnetic Resonance Angiography Using Multibolus TrueFISP-Based Spin Tagging with Alternating Radiofrequency (TrueSTAR)

Lirong Yan,¹ Noriko Salamon,² and Danny J. J. Wang^{1,2*}

Purpose: The goal of this study was to introduce a new noncontrast enhanced 4D dynamic MR angiography (dMRA) technique termed multibolus TrueFISP-based spin tagging with alternating radiofrequency (TrueSTAR).

Methods: Multibolus TrueFISP-based spin tagging with alternating radiofrequency was developed by taking advantage of the phenomenon that the steady-state signal of TrueFISP is minimally disturbed by periodically inserted magnetization preparations (e.g., spin tagging) that are sandwiched by two $\alpha/2$ RF pulses. Both theoretical analysis and experimental studies were carried out to optimize the proposed method which was compared with both pulsed and pseudo-continuous arterial spin labeling-based dMRA in healthy volunteers. Optimized multibolus dMRA was also applied in a patient with arteriovenous malformation to demonstrate its potential clinical utility.

Results: Multibolus dMRA offered a prolonged tagging bolus compared to the standard single-bolus dMRA, and allowed improved visualization of the draining veins in the arteriovenous malformation patient. Compared to pseudo-continuous arterial spin labeling-based dMRA, multibolus dMRA provided visualization of the full passage of the labeled blood with the flexibility for both static and dynamic magnetic resonance angiography.

Conclusion: By combining the benefits of pulsed and pseudocontinuous arterial spin labeling-based dMRA, multibolus True-FISP-based spin tagging with alternating radiofrequency can prolong and enhance the tagging bolus without sacrificing imaging speed or temporal resolution. **Magn Reson Med 71:551–560, 2014.** © **2013 Wiley Periodicals, Inc.**

Key words: dynamic magnetic resonance angiography (dMRA); non-contrast enhanced MRA; steady-state free precession (SSFP); true fast imaging with steady state precession (TrueFISP); arterial spin labeling (ASL); arteriovenous malformation (AVM)

DOI 10.1002/mrm.24689

Published online 25 February 2013 in Wiley Online Library (wileyonlinelibrary.com).

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Arterial spin labeling (ASL) techniques were originally introduced and developed for noncontrast enhanced MR angiography (MRA) in the 1980s (1-3). Since the seminal study by Detre et al. in 1992 (4,5), ASL has evolved into a class of MRI techniques for noninvasive measurements of microvascular perfusion of the brain and body organs (6). Recently, there has been a resurgent trend for the development and applications of ASL-based MRA techniques. This emerging trend may be attributed to the advent of spin tagging techniques with improved signal-to-noise ratio (SNR), such as pseudo-continuous arterial spin labeling (pCASL) (7,8) with the capability for vessel-selective labeling (9,10). Compared to pulsed ASL (PASL), pCASL-based MRA demonstrated not only increased SNR but also reduced sensitivity to variations in delay time (inversion time) and repetition time (TR) for imaging carotid arteries (11). At the meantime, various fast imaging techniques became mature which offer adequate spatial resolution and volumetric imaging coverage for performing ASL-based MRA within a short time period. A recent study combined pCASL with accelerated 3D radial acquisition (vastly undersampled isotropic projection imaging) to achieve whole-brain MRA with submillimeter 3D isotropic resolution (12). The drawbacks for the use of pCASL for MRA, however, include relatively long labeling durations (a few seconds) that may compete with image acquisition.

Another notable trend in ASL-based MRA involves the development of time-resolved noncontrast enhanced 4D dynamic MRA (dMRA) (13,14). In 4D dMRA, spin labeling is combined with a cine multiphase balanced steady-state free precession readout, offering both high temporal (a few ten milliseconds) and spatial resolution (a few cubic millimeter) for depicting the dynamic flow pattern through the vasculature. Initial clinical evaluations of 4D dMRA showed promises in patients with arteriovenous malformation (AVM) (15,16) and stenoocclusive disease (17), as compared to the gold standard of digital subtraction angiography. In 4D dMRA, the apparent blood T_1 may be shorter than the true blood T_1 depending on the blood flow velocity and imaging parameters of the balanced steady-state free precession readout such as the flip angle. This apparent blood T_1 is a primary limiting factor for the length of the tagging bolus (approximately,1-2 s) in dMRA methods. This relatively narrow dynamic temporal window may limit the clinical utility of 4D dMRA, e.g., for visualizing draining veins of AVM which is a critical factor in clinical evaluations (15,16).

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Grant sponsor: NIH; Grant numbers: R01-MH080892; R01-NS081077; R01-B014922.

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Received 21 September 2012; revised 7 January 2013; accepted 22 January 2013



FIG. 1. Diagram of multibolus TrueSTAR sequence that employs a train of intermittent HS inversion pulses for spin labeling during the multiphase-segmented trueFISP readout. Spin tagging is implemented using the STAR scheme. Each inversion pulse (except the first one) is sandwiched by two $\alpha/2$ pulses and interleaved by a number of phases of balanced SSFP acquisitions. A presaturation pulse is applied at the beginning of the sequence to saturate the background signal in the imaging slab.

To date, most existing dMRA techniques have used PASL for spin tagging, given its short duration that allows the visualization of the full passage of labeled blood through arteries. Pseudo-continuous arterial spin labeling has been attempted for dMRA at the cost of prolonged scan time, and a few initial phases of arterial inflow may be sacrificed (10,18). It will be ideal to combine the benefits of both PASL, including short RF duration and technical simplicity, and pCASL, including higher SNR and prolonged labeling bolus, in 4D dMRA to improve the image quality as well as to increase the temporal window for visualizing the full passage of dynamic blood flow. The primary purpose of this study was to introduce such a dMRA technique termed multibolus TrueFISP-based spin tagging with alternating radiofrequency (TrueSTAR). Both theoretical analysis and in vivo experiments were carried out in this study to demonstrate the feasibility of the proposed approach, which was compared with both PASL- and pCASL-based dMRA methods.

THEORY

As shown by Scheffler et al. (19), the steady-state signal of the TrueFISP sequence is minimally disturbed by periodically inserted magnetization preparation such as fat saturation, which is achieved by an $\alpha/2$ flip-back pulse before the magnetization preparation to temporally store the established steady-state signal as pure longitudinal magnetization. After the magnetization preparation, this longitudinal magnetization is excited by another $\alpha/2$ pulse to continue the TrueFISP acquisition.

The proposed multibolus TrueSTAR technique took advantage of this phenomenon, in which a number of magnetization preparations using PASL were applied during the continuous multiphase segmented TrueFISP acquisition without significant modifications of the steady-state signal. Figure 1 shows the pulse sequence diagram of multibolus TrueSTAR. Spin tagging is implemented using the STAR scheme (20) by applying a hyperbolic secant (HS) inversion pulse inferior to the imaging slab. Each inversion pulse (except the first one) is sandwiched by two $\alpha/2$ pulses and interleaved by a number of phases of balanced steady-state free precession acquisitions. A presaturation pulse is applied at the beginning of the sequence to saturate the background signal in the imaging slab.

The temporal profile of labeled blood bolus at the tagging site using pulsed ASL, W'(t), can be described as a boxcar function convoluted with a dispersion kernel (21). By incorporating relaxation with the T_1 of blood (T_{1b}) as the label travels from the tagging region to the imaging region, the magnetization of labeled arterial blood, $M_a(t)$, for PASL can be expressed as

$$M_{\rm a}(t) = 2M_{\rm 0b}e^{-t/T_{\rm 1b}}W'(t)$$
[1]

$$W'(t) = k(t) \otimes W(t)$$
[2]

where $M_{\rm ob}$ is the equilibrium magnetization of arterial blood, W'(t) is the temporal profile of the labeled bolus with dispersion, k(t) is the dispersion convolution kernel, and W(t) is the boxcar function. The gaussian dispersion model proposed by Hrabe and Lewis (22) was used in this study

$$k(t) = \frac{1}{\sqrt{2\pi\sigma}} e^{-t^2/2\sigma^2}$$
 [3]

where σ is the standard deviation of the gaussian normal distribution. Hrabe and Lewis (22) further suggested that the trailing edge (τ_2) of the labeled bolus should be more dispersed than the leading edge (τ_1), resulting in

$$W'(t) = \frac{1}{2} \left[\operatorname{erf}\left(\frac{t - \tau_1}{\sqrt{2\sigma_1}}\right) - \operatorname{erf}\left(\frac{t - \tau_2}{\sqrt{2\sigma_2}}\right) \right]$$
[4]

where $\operatorname{erf}(t)$ is the error function evaluated at t, σ_1 , and σ_2 denote the standard deviations of the leading (τ_1) and trailing edge (τ_2) of the labeled bolus, respectively. In brief, σ_2 is generally set to $\sigma_1 \sqrt{\tau_2/\tau_1}$. Although alternative dispersion models may be applied, no significant differences have been shown between gaussian and other dispersion models such as gamma variate function for modeling dMRA signals (21).

In the case of more than one tagging bolus, the observed labeled blood signal will be the simple summation of individual boluses, given no interactions between the successive boluses. However, if the leading edge of the subsequent bolus occurs before the trailing edge of the preceding bolus, the overlapping part of the two



FIG. 2. Illustration of the steps for simulating longitudinal magnetizations of labeled arterial blood of two boluses by using the boxcar function (**a**), then including T_1 relaxation (**b**) and dispersion effects (**c**). Here, two time intervals of 300 and 400 ms between boluses were used with the assumption of v = 20 cm/s. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

boluses needs to be taken into account. Figure 2 shows the steps for simulating two labeled boluses with and without overlap, respectively. For the sake of simplicity, only the case that the interaction happens between two consecutive labeled boluses was considered in this study. The labeled blood signal with multibolus PASL can be expressed as:

$$M_{\rm a}(t) = \sum_{i=1}^{n} M_{{\rm a}(i)}(t - (i-1)\Delta t) + \sum_{i=1}^{n-1} M_{{\rm a}(i,i+1)}(t - i\Delta t) \quad [5]$$

$$M_{a(i,i+1)}(t) = \begin{cases} 2M_{0b} \left(1 - e^{-\Delta t/T_{1b}}\right) e^{-t/T_{1b}} W_{i,i+1}'(t) & v\Delta t < D\\ 0 & v\Delta t \ge D \end{cases}$$
[6]

where *i* is the bolus number, Δt is the time interval of two consecutive boluses, $M_{a(i)}$ is the arterial blood mag-

netization of the *i*th bolus according to Eq. [1]. $M_{a(i,i+1)}$ is the modified bolus caused by the interaction of the *i*th and (i+1)th labeling boluses (Fig. 2b,c), *D* is the width of tagging slab, and *v* is flow velocity. The parameters of τ_1 and τ_2 in W'_i and $W_{i,i+1}'$ were determined by the tagging slab thickness, the gap between the tagging and imaging slab, and flow velocity.

One advantage of using multiphase TrueFISP for dMRA is that it causes minimal perturbation of the longitudinal magnetization of flowing spins, as demonstrated by both Bloch equation simulation and phantom experiments (23,24). Therefore, the potential saturation effect of multiphase TrueFISP on labeled blood signal was ignored in this study, and multibolus TrueSTAR dMRA signal can be approximated by the simulated $M_a(t)$ signal.

Table 1							
Imaging	Parameters	Used	in	the	Simulation	and	Experiments

	Aim1: optimiza parameters of mu	ition of imaging Itibolus TrueSTAR	Aim2: evaluation of optimized multibolus TrueSTAR
Parameters of multibolus dMRA	Simulation	Exp 1	Exp 2
Inversion recovery thickness (cm)	8	8	8
Gap between tagging and imaging slab (cm)	2	2	2
Time interval between inversion recoveries (ms)	200/300/400/500	210/315/420/525	420
Bolus number	3	3	2, 3, 4
Flow velocity (cm/s)	15, 20, 25, 30	16–30	

METHODS

Simulations of Multibolus TrueSTAR

Multibolus and single-bolus TrueSTAR dMRA signals were simulated using in-house Matlab programs (Mathworks, Natick, MA), with parameters matching those used in experiments. The STAR tagging slab was 80 mm with a gap of 20 mm to the imaging slab. Four flow velocities of 15, 20, 25, and 30 cm/s were chosen for the stimulation, according to the measured mean flow velocity of 21 cm/s with a range of 16–30 cm/s in internal carotid arteries from all the subjects. The leading and trailing edge (τ_1 and τ_2) of the (single) labeling bolus were determined by the tagging slab thickness, gap between the tagging region to imaging slab, and assumed mean flow velocity. For example, with an 80-mm tagging slab, 20 mm gap, and 20 cm/s flow velocity, τ_1 and τ_2 were 100 and 500 ms, respectively. In addition, σ_1 of 0.1 was used for the simulation, which was derived from curve-fitting of the time course of experimental singlebolus dMRA data. Simulation was performed for singleand three-bolus TrueSTAR. The time interval between two successive inversion pulses was varied from 200 to 500 ms with a step of 100 ms. The rest of the parameters used in the simulation were $T_{1b} = 1600$ ms, temporal resolution = 100 ms.

Experiments

All experiments were performed on a Siemens TIM Trio 3T scanner. Eight healthy volunteers $(24.6 \pm 3.6 \text{ years})$ three males) without history of cerebrovascular diseases participated in this study after providing written informed consents. Time-of-flight-MRA images were first acquired with the following parameters: field of view-= 200 \times 232 mm², resolution = 0.8 \times 0.8 \times 0.8 mm³, $TR\,{=}\,20$ ms, echo time ${=}\,3.6$ ms, flip angle ${=}\,18^\circ.$ For single- and multibolus TrueSTAR scans, a 3D imaging slab of 40 slices with 1.5-mm thickness was scanned to cover the Circle of Willis and its main branches. The remaining imaging parameters were field of view = 220×165 mm², resolution = 1 \times 1 \times 1.5 mm³, TR/echo time-= 4.24/2.12 ms, rate-2 GRAPPA, centric ordering k-space acquisition with 25 lines per segment, 22 phases from 150 to 2355 ms with a step of 105 ms, 500 ms delay time between the last temporal phase and the following presaturation pulse, and a total scan time of 7 min.

Two experiments were conducted: (1) Exp 1: Experimental optimization of imaging parameters for multibolus dMRA; (2) Exp 2: Evaluation of optimized multibolus dMRA by comparison with standard single-bolus PASL dMRA and pCASL dMRA.

The main imaging parameters of multibolus dMRA in Exp 1 and Exp 2 are listed in Table 1.

Exp 1: Experimental Optimization of Multibolus TrueSTAR

A three-bolus TrueSTAR sequence was implemented with HS inversion pulses (except the first pulse) inserted after the phase numbers of [2, 4], [3, 6], [4, 8] or [5, 10] during multiphase TrueFISP acquisitions, corresponding to the time interval of 210, 315, 420, and 525 ms, respectively. The HS inversion pulses (duration, 15 ms) were applied to an 80-mm slab inferior to the image slab with a 20-mm gap. A single-slice phase contrast MRI was performed at approximately the center of the tagging slab to measure blood flow velocities in internal carotid and vertebral arteries. The imaging parameters were slice thickness = 5 mm, field of view = 20 cm, matrix size = 256×256 , echo time/TR = 5/18 ms, flip angle = 15° , 80 cm/s velocity encoding.

Exp 2: Evaluation of Optimized Multibolus dMRA

The optimal parameters of multibolus TrueSTAR were determined based on Exp 1 and simulation results: 400 ms was chosen as the optimal bolus interval for an 80mm tagging slab with flow velocities from 15 to 30 cm/s. Therefore, the phase interval of 4 (corresponding to 420 ms) was chosen for Exp 2, with all other parameters identical to the protocol used in Exp 1. Three TrueSTAR dMRA scans were performed with 2, 3, and 4 inversion pulses inserted after the phase numbers of 4, [4, 8], and [4, 8, 12] during multiphase TrueFISP acquisitions, respectively. For comparison, a standard single-bolus TrueSTAR sequence was performed using otherwise identical imaging parameters. Multibolus TrueSTAR can achieve a continuous labeling bolus, analogous to pCASL, by intermittently tagging inflowing blood during data acquisition. For comparison, a pCASL-based dMRA sequence was also performed using pCASL with balanced gradients between label and control acquisitions (7). Three pCASL dMRA scans were performed with the labeling durations of 300, 600, and 900 ms, respectively. The imaging parameters were closely matched to those of multibolus TrueSTAR, except that the total phase number of TrueFISP acquisitions was decreased to 15, 15, and 12, respectively, depending on the labeling duration of pCASL.



FIG. 3. Simulated average labeled blood signal by standard (single) and three-bolus TrueSTAR with different bolus intervals (averaged over flow velocities from 15 to 30 cm/s) (a), and with bolus intervals of 300 ms (b) and 400 ms (c) at different flow velocities, respectively. The experimental dMRA time courses within global arterial ROI (d) are consistent with simulated dMRA signals (a).

Patient Study

One AVM patient (42 years, female) underwent singlebolus and four-bolus dMRA scans using the same imaging parameters as Exp 2. After dMRA scans, a time-offlight-MRA scan was performed after the injection of gadolinium contrast agent (0.1 mmol/kg).

Data Analysis

For both PASL- and pCASL-based dMRA scans, dMRA images were generated by complex subtraction between label and control acquisitions, followed by maximum intensity projection (MIP) for each temporal phase along three directions (axial, sagittal, and coronal), respectively. The MIP images along each direction can be displayed as a movie to visualize the dynamic blood flow through the Circle of Willis and its main branches.

Collapsed axial MIP images across all phases were generated for each scan. A mask containing the Circle of Willis and all of its branches was manually drawn to remove the skull and background signal outside the brain. Within this mask, a global arterial region-of-interest (ROI) was defined by user-specified threshold to isolate the main branches of the Circle of Willis from the tissue. Dynamic time courses were derived from the global arterial ROI of each subject. For each subject, the background noise level was measured from four manually drawn ROIs from background regions without apparent artifacts. Apparent SNR was calculated from the mean signal of the MIP image in the global arterial ROI divided by the standard deviation of the signal in the background ROIs for each temporal phase. Time-dependent SNR was defined by averaging the apparent SNR every three temporal phases to show its variation with time, whereas average SNR was calculated by averaging the apparent SNR across temporal phases of which the dMRA signal in the arterial ROI was 10% higher than the baseline (the lowest signal across all temporal phases).

Statistical Analysis

Statistical analyses were performed using the SPSS 19.0 software package (SPSS Inc., Chicago, IL). The SNR values of standard single-bolus TrueSTAR, multibolus TrueSTAR and pCASL dMRA were compared using paired *t*-tests. A two-tailed *P*-value of 0.05 or less was considered to indicate a significant difference.

RESULTS

Simulation of Multibolus TrueSTAR

Figure 3a shows the average time courses of simulated dMRA signals using single- and three-bolus TrueSTAR using flow velocities from 15 to 30 cm/s with different bolus intervals from 200 to 500 ms, respectively. Compared to the standard single-bolus TrueSTAR, a prolonged bolus of labeled blood can be achieved using multibolus TrueSTAR. The width of the integrated bolus increases with prolonged time interval between successive inversion pulses in multibolus TrueSTAR, however, with greater signal variations across boluses. As shown in Figure 3a, when the bolus interval is short (e.g., 200 ms), there is relatively large overlap between two consecutive boluses, resulting in an integrated bolus with



FIG. 4. Axial dMRA MIP images acquired using standard TrueSTAR (a), TrueSTAR with two-bolus (b), three-bolus (c), and four-bolus (d) with bolus interval of 420 ms from a representative subject. The average time between two phases was 105, 106, 107, or 108 ms for single-, two-, three-, or four-bolus cases, respectively (owing to the insertion of additional HS inversion pulses). The passages of labeled blood flowing through the Circle of Willis (arrow) into the main branches can be well visualized in both standard and multibolus TrueSTAR. A prolonged bolus was observed by multibolus TrueSTAR. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

reduced peak intensities. With increased bolus interval, there is less overlap between consecutive boluses, resulting in a prolonged bolus in multibolus dMRA (300 and 400 ms intervals, Fig. 3a). However, signal variations across boluses are also increased as each bolus is more separated in time (500-ms interval, Fig. 3a). From the simulation, the optimal bolus interval should be around 300–400 ms for achieving a prolonged bolus with a relatively stable plateau, assuming a flow velocity range of 15–30 cm/s.

Figure 3b,c shows simulated multibolus dMRA signals with the bolus intervals of 300 and 400 ms using various flow velocities, respectively. The plateau of the integrated bolus with 400-ms interval was relatively stable across the flow velocity range of 15–30 cm/s. However, the peak signal of multibolus dMRA with 300-ms interval dropped at the lower flow velocity of 15 cm/s. Overall, the bolus interval of 400 ms yielded optimal results for achieving a prolonged bolus with a relatively stable plateau.

Exp 1: Experimental Optimization of Multibolus TrueSTAR

Figure 3d shows the average dMRA time courses in the arterial ROI using three-bolus TrueSTAR with different bolus intervals, respectively. The experimental results are consistent with the simulation result shown in Figure 3a. The duration of the integrated bolus increased with prolonged time interval between successive inversion pulses. With a short bolus interval of 210 ms (two phases), the peak dMRA signal was decreased compared to that of single-bolus dMRA, owing to the cancellation

of two consecutive boluses. With increased bolus interval, a prolonged continuous bolus was achieved by integrating individual boluses. However, with the longest bolus interval of 525 ms, large dMRA signal variations across boluses were observed. Based on both the simulation and the experimental results shown in Figure 3, 420 ms (four phases) was chosen as the optimal bolus interval to achieve a prolonged and continuous bolus of labeled blood signal in multibolus dMRA.

Exp 2: Evaluation of Optimized Multibolus dMRA

Comparison of Single-Bolus and Multibolus TrueSTAR

Figure 4 shows axial MIP images of dMRA signals from a representative subject, acquired using standard singlebolus TrueSTAR (top row), TrueSTAR with two (second row), three (third row), and four boluses (bottom row), respectively. In all cases, the passage of labeled blood flowing through the Circle of Willis into the main branches can be visualized with both high spatial and temporal resolution. As shown in Figure 5 of the mean dMRA time courses in the arterial ROI, a prolonged bolus with a relatively constant plateau was achieved using multibolus TrueSTAR. As shown in Figure 6a of the average SNR values of single- and multibolus dMRA from five healthy volunteers, the average SNR of multibolus TrueSTAR was $13.6 \pm 2.2\%$ higher than that of single-bolus TrueSTAR although statistical significance was achieved only for two-bolus TrueSTAR (P = 0.029). Figure 6b shows the time-dependent SNR averaged every



FIG. 5. Normalized averaged time courses of dMRA signals in the arterial ROI by dividing the maximum dMRA signal with that of the standard TrueSTAR. A prolonged bolus was achieved by multibolus dMRA.

three temporal phases between single- and two-bolus TrueSTAR. The time-dependent SNR of two-bolus TrueSTAR was significantly higher between 1 and 1.75 s than that of single-bolus TrueSTAR (P < 0.05).

Comparison Between Multibolus TrueSTAR and pCASL-dMRA

Figure 7 shows pCASL-based dMRA images with three labeling durations (300, 600, and 900 ms) from the same subject of Figure 4. The average SNR of pCASL dMRA with 900 ms labeling duration was 27.5 ± 5.9 and $13.6 \pm 6.7\%$ higher than that of single- (P = 0.006) and two-bolus TrueSTAR (P = 0.007), respectively (Fig. 6a). As shown in Figure 7, pCASL-based dMRA provided better delineation of fine distal arteries compared to single- or multibolus PASL-based dMRA. However, the arterial inflow phases were missing in pCASL-based dMRA.

Evaluation of Imaging Artifacts in Multibolus TrueSTAR

Potential disturbances of the steady-state TrueFISP signal caused by inserting PASL magnetization preparations were investigated. As shown in Figure 8, compared to the TrueFISP images acquired before the inversion pulse, there were slight but visible signal fluctuations in the TrueFISP images acquired after the inversion pulse (arrow). However, the signal fluctuations were largely suppressed in dMRA images by the pairwise subtraction between control and label acquisitions.

Multibolus TrueSTAR dMRA on AVM

Figure 9 shows representative time frames of axial dMRA MIP images as well as collapsed MIP image with multibolus (a) and standard single-bolus (b) TrueSTAR of the AVM patient. Dynamic flow passages from the feeding artery (anterior parietal branch of right middle cerebral artery, red arrow), into the nidus ($4 \times 6 \text{ mm}^2$ in the right supra-marginal gyrus, yellow arrow), and draining veins (right parietal cortical vein draining into the right vein of Labbe) can be clearly visualized using multibolus TrueSTAR. Compared to standard single-bolus TrueSTAR, better delineation of the nidus and draining veins with improved contrast were achieved using multibolus TrueSTAR. The apparent SNR of the draining vein in the collapsed MIP image was 41.2 and 34.4 for fourbolus and standard TrueSTAR, respectively.

DISCUSSION

A new dMRA technique termed multibolus TrueSTAR was introduced in this study by taking advantages of the phenomenon that the steady-state signal of the TrueFISP sequence is minimally disturbed by periodically inserted magnetization preparations sandwiched by two $\alpha/2$ pulses (Fig. 8). Compared to single-bolus dMRA, a prolonged bolus with a relatively stable plateau was achieved by multibolus TrueSTAR with an optimal time interval between successive labeling pulses. The peak



FIG. 6. **a**: Average SNR of dMRA signals in the arterial ROI by standard, multibolus TrueSTAR and pCASL dMRA. **b**: Time-dependent SNR of dMRA signals in the arterial ROI by standard and two-bolus TrueSTAR.

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FIG. 7. Axial dMRA MIP images acquired using pCASL dMRA with the labeling duration of 300 (a), 600 (b), and 900 ms (c) from the same subject of Figure 4.

intensities of dMRA signals, however, were comparable between single- and multibolus TrueSTAR. pCASL-based dMRA with a sufficiently long tag (labeling duration, >600 ms) offered increased SNR compared to that of single-bolus TrueSTAR. However, a few temporal phases of arterial inflow were missing using pCASL-based dMRA.



FIG. 8. Fluctuations caused by the inserted PASL magnetization preparation during TrueFISP acquisition. A representative slice of label, control, subtraction, and MIP images before (a) and after (b) the inversion pulse during the TrueFISP acquisition are shown. Red arrows indicate signal fluctuations in the TrueFISP images acquired after the inversion pulse.



FIG. 9. Representative temporal phases of axial dMRA MIP images and collapsed MIP image acquired using four-bolus TrueSTAR (a) and standard TrueSTAR (b) from the AVM patient. Time-of-flight-MRA-MIP image after the injection of gadolinium contrast agent is shown on the right side. Multibolus dMRA provided improved visualization of the nidus and draining veins.

Overall, the proposed multibolus TrueSTAR technique may offer a novel approach by combining the benefits of PASL and pCASL-based dMRA.

Pros and Cons of Multibolus TrueSTAR

The proposed multibolus TrueSTAR was able to achieve a continuous bolus with a relatively stable plateau of labeled blood, analogous to CASL and pCASL. Since its introduction (25), pCASL has drawn growing interests, given its high labeling efficiency and SNR, as well as compatibility with MRI hardware. Besides quantitative perfusion imaging, pCASL has recently been applied for both static and dynamic MRA with extensions for vesselselective labeling (10,12,26). Potential drawbacks of pCASL, however, include long labeling durations during which image acquisition is prohibited. In existing pCASL dMRA studies, a tradeoff had to be made between the duration of labeling pulses and thus SNR and the number of arterial inflow phases that may be sacrificed (10). Compared to pCASL dMRA, multibolus TrueSTAR offers the benefit for visualizing the full dynamic passage of labeled blood without penalties in scan time. In addition, multibolus dMRA images acquired during the bolus plateau may be averaged to improve SNR and the delineation of distal arteries. As the duration of the bolus plateau can be as long as required, there should be minimal concern for missing the peak bolus signal as in existing PASL-based MRA.

One critical issue in multibolus TrueSTAR regards the timing of multiple inversion pulses during multiphase TrueFISP acquisitions. If the time interval between two consecutive inversion pulses is too short, the two boluses will partially cancel each other, leading to a loss of tagging efficiency. On the contrary, if the time interval is too long, there will be obvious signal drop before the next inversion pulse is applied. With a mean flow velocity of 20 cm/s in carotid arteries, both theoretical analysis and experimental results indicated that multibolus TrueSTAR with a time interval of approximately 400 ms between consecutive inversion pulses can provide a rela-

tively continuous bolus of labeled blood. The optimal bolus interval of multibolus dMRA for clinical populations (e.g., steno-occlusive diseases), however, awaits to be investigated in future studies. Another potential shortcoming of multibolus TrueSTAR is that the True-FISP readout is sensitive to field inhomogeneity effects, which may cause banding artifacts around the orbitofrontal cortex. A phase cycling approach may be necessary to address this issue in future studies.

Multibolus TrueSTAR dMRA on AVM

Multibolus dMRA was piloted in an AVM patient, which depicted the entire dynamic blood flow from the arterial feeder to the nidus and draining veins of the AVM. Compared to standard single-bolus dMRA, multibolus dMRA provided improved visualization of the nidus and draining veins. Although the peak intensity of multibolus TrueSTAR was comparable with that of the standard single-bolus TrueSTAR in healthy volunteers, it may be increased in distal vessels and/or draining veins of AVM owing to greater dispersion (or smoothing) effects, leading to an enhanced integrated bolus. The capability of multibolus dMRA for the identification of feeding arteries of an AVM is helpful for standard microsurgical operation in which the arterial feeders are generally occluded first, followed by excision of the nidus and finally resection of the draining veins. Information of feeding arteries can also act as referable factors for determining the sequence of arteries to be managed. Nevertheless, the clinical value of multibolus TrueSTAR versus existing techniques such as digital subtraction angiography awaits to be evaluated in future studies with larger cohorts of patients.

Multibolus dMRA and Perfusion Imaging

Past studies have employed multiple inversion pulses in ASL experiments for perfusion imaging (27), with an approximately twofold SNR gain in perfusion signal compared to the standard FAIR technique at 3T. The drawbacks for using multibolus approaches for perfusion multibolus PASL remains challenging. The proposed multibolus TrueSTAR may be more suitable for dMRA than perfusion imaging as no delay is required between the labeling and the image acquisition and the arterial input magnetization of labeled blood can be visualized with both high spatial and temporal resolution in 4D space. However, unlike brain tissue that can serve as a reservoir for labeled blood water, the labeled blood flows through an arterial voxel with limited dispersion. As shown by both simulation and experimental results, the integrated bolus of multibolus dMRA was primarily prolonged rather than enhanced compared to that of singlebolus dMRA. In the future, it may be possible to combine multibolus dMRA and perfusion imaging to improve the SNR and accuracy of quantitative perfusion MRI using PASL.

CONCLUSIONS

In conclusion, a novel time-resolved 4D dMRA technique termed multibolus TrueSTAR was introduced in this study, by combining the advantages of both PASLand pCASL-based dMRA. Its potential clinical utility, such as the delineation of feeding arteries, nidus, and draining veins of AVM was initially demonstrated in this study.

ACKNOWLEDGEMENT

The authors are grateful to Dr. Alessandra Gorgulho and Dr. Antonio De Salles for helping with patient scanning, and Cheng Li for his assistance with the manuscript.

REFERENCES

- Dixon WT, Du LN, Faul DD, Gado M, Rossnick S. Projection angiograms of blood labeled by adiabatic fast passage. Magn Reson Med 1986;3:454–462.
- Nishimura DG, Macovski A, Pauly JM. Considerations of magnetic resonance angiography by selective inversion recovery. Magn Reson Med 1988;7:472–484.
- 3. Nishimura DG, Macovski A, Pauly JM, Conolly SM. MR angiography by selective inversion recovery. Magn Reson Med 1987;4:193–202.
- Detre JA, Leigh JS, Williams DS, Koretsky AP. Perfusion imaging. Magn Reson Med 1992;23:37–45.
- Williams DS, Detre JA, Leigh JS, Koretsky AP. Magnetic resonance imaging of perfusion using spin inversion of arterial water. Proc Natl Acad Sci USA 1992;89:212–216.
- Detre JA, Rao H, Wang DJ, Chen YF, Wang Z. Applications of arterial spin labeled MRI in the brain. J Magn Reson Imaging 2012;35:1026– 1037.
- Wu WC, Fernandez-Seara M, Detre JA, Wehrli FW, Wang J. A theoretical and experimental investigation of the tagging efficiency of pseudocontinuous arterial spin labeling. Magn Reson Med 2007;58:1020– 1027.
- Dai W, Garcia D, de Bazelaire C, Alsop DC. Continuous flow-driven inversion for arterial spin labeling using pulsed radio frequency and gradient fields. Magn Reson Med 2008;60:1488–1497.

- 9. Wong EC. Vessel-encoded arterial spin-labeling using pseudocontinuous tagging. Magn Reson Med 2007;58:1086–1091.
- Okell TW, Chappell MA, Woolrich MW, Gunther M, Feinberg DA, Jezzard P. Vessel-encoded dynamic magnetic resonance angiography using arterial spin labeling. Magn Reson Med 2010;64:430–438.
- Koktzoglou I, Gupta N, Edelman RR. Nonenhanced extracranial carotid MR angiography using arterial spin labeling: improved performance with pseudocontinuous tagging. J Magn Reson Imaging 2011;34:384–394.
- 12. Wu H, Block WF, Turski PA, Mistretta CA, Johnson KM. Noncontrast-enhanced three-dimensional (3D) intracranial MR angiography using pseudocontinuous arterial spin labeling and accelerated 3D radial acquisition. Magn Reson Med 2013;69:708–715.
- Bi X, Weale P, Schmitt P, Zuehlsdorff S, Jerecic R. Non-contrastenhanced four-dimensional (4D) intracranial MR angiography: a feasibility study. Magn Reson Med 2010;63:835–841.
- 14. Yan L, Wang S, Zhuo Y, Wolf RL, Stiefel MF, An J, Ye Y, Zhang Q, Melhem ER, Wang DJ. Unenhanced dynamic MR angiography: high spatial and temporal resolution by using true FISP-based spin tagging with alternating radiofrequency. Radiology 2010;256:270–279.
- 15. Yu S, Yan L, Yao Y, Wang S, Yang M, Wang B, Zhuo Y, Ai L, Zhao J, Wang DJ. Evaluation of non-contrast dynamic MRA in intracranial arteriovenous malformation (AVM): comparison with time of flight (TOF) and digital subtraction angiography (DSA). Magn Rason Imaging 2012;30:869–877.
- Xu J, Shi D, Chen C, Li Y, Wang M, Han X, Jin L, Bi X. Noncontrastenhanced four-dimensional MR angiography for the evaluation of cerebral arteriovenous malformation: a preliminary trial. J Magn Reson Imaging 2011;34:1199–1205.
- 17. Lanzman RS, Kropil P, Schmitt P, Bi X, Gliem M, Miese FR, Hanggi D, Kamp M, Scherer A, Turowski B, et al. Nonenhanced ECG-gated time-resolved 4D steady-state free precession (SSFP) MR angiography (MRA) for assessment of cerebral collateral flow: comparison with digital subtraction angiography (DSA). Eur Radiol 2011;21:1329–1338.
- Okell TW, Chappell MA, Schulz UG, Jezzard P. A kinetic model for vessel-encoded dynamic angiography with arterial spin labeling. Magn Reson Med 2012;68:969–979.
- Scheffler K, Heid O, Hennig J. Magnetization preparation during the steady state: fat-saturated 3D TrueFISP. Magn Reson Med 2001;45:1075–1080.
- 20. Edelman RR, Siewert B, Darby DG, Thangaraj V, Nobre AC, Mesulam MM, Warach S. Qualitative mapping of cerebral blood flow and functional localization with echo-planar MR imaging and signal targeting with alternating radio frequency. Radiology 1994;192:513–520.
- Chappell MA, Woolrich MW, Kazan S, Jezzard P, Payne SJ, Macintosh BJ. Modeling dispersion in arterial spin labeling: validation using dynamic angiographic measurements. Magn Reson Med 2013;69:563–570.
- Hrabe J, Lewis DP. Two analytical solutions for a model of pulsed arterial spin labeling with randomized blood arrival times. J Magn Reson 2004;167:49–55.
- Yan L, Li C, Kilroy E, Wehrli FW, Wang DJ. Quantification of arterial cerebral blood volume using multiphase-balanced SSFP-based ASL. Magn Reson Med 2012;68:130–139.
- 24. Wu WC, Jain V, Li C, Giannetta M, Hurt H, Wehrli FW, Wang DJ. In vivo venous blood T1 measurement using inversion recovery true-FISP in children and adults. Magn Reson Med 2010;64:1140–1147.
- 25. Garcia DM, de Bazelaire C, Alsop D. Pseudo-continuous flow driven adiabatic inversion for arterial spin labeling. In Proceedings of the ISMRM Annual Meeting, Miami, Florida, USA, 2005. Abstract 37.
- Robson PM, Dai W, Shankaranarayanan A, Rofsky NM, Alsop DC. Time-resolved vessel-selective digital subtraction MR angiography of the cerebral vasculature with arterial spin labeling. Radiology 2010;257:507–515.
- 27. Fujiwara Y, Kimura H, Miyati T, Kabasawa H, Matsuda T, Ishimori Y, Yamaguchi I, Adachi T. MR perfusion imaging by alternate slab width inversion recovery arterial spin labeling (AIRASL): a technique with higher signal-to-noise ratio at 3.0 T. MAGMA 2012;25:103–111.